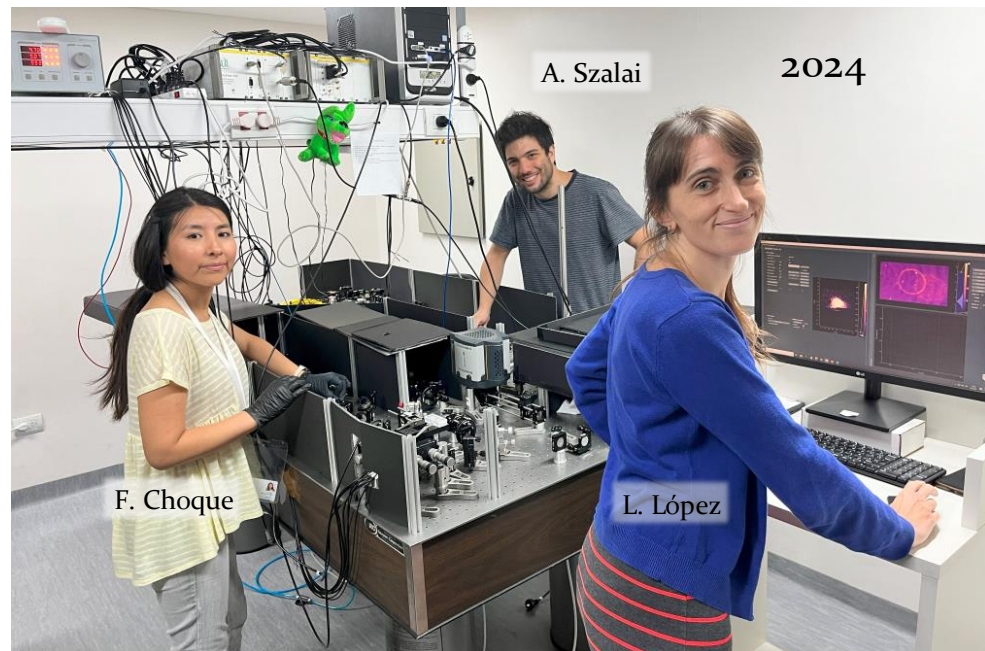
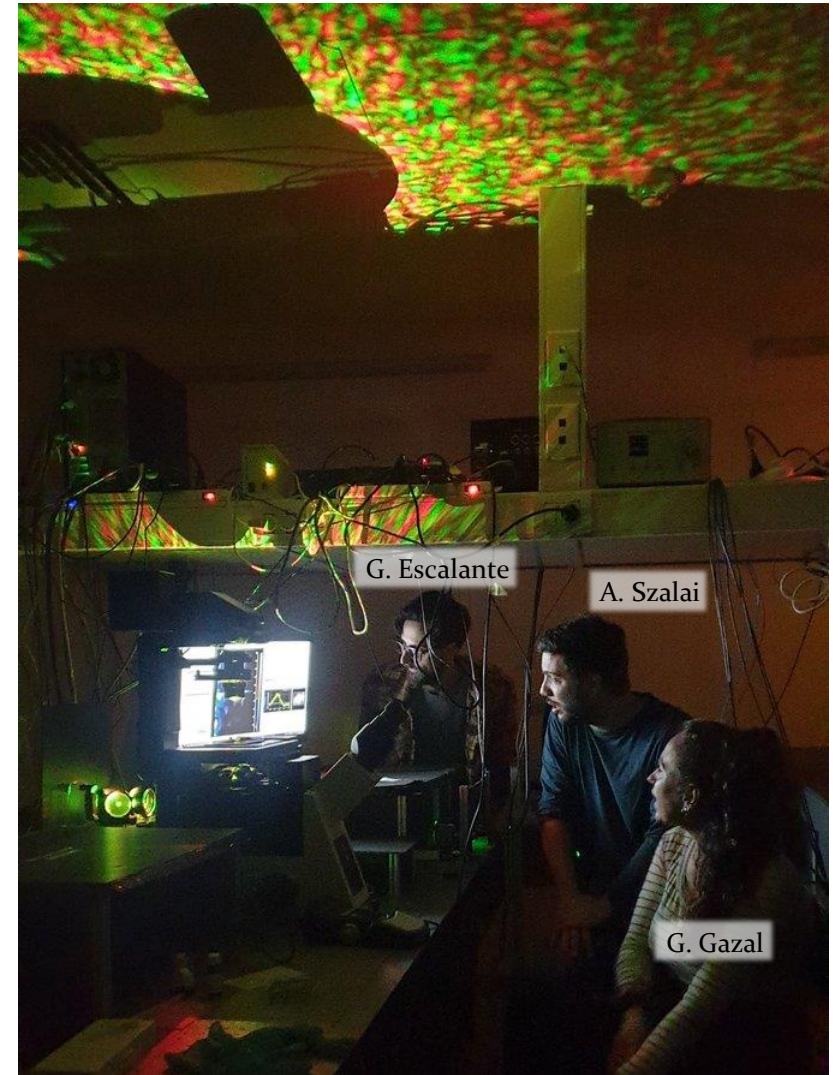


Advanced Fluorescence Imaging

Prof. Dr. Fernando D. Stefani

2023 / 2024



@FerStefaniLab

<https://stefani-lab.ar/>

Acknowledgements

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FIL Buenos Aires

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Advanced Fluorescence Imaging

Day 1: Fluorophores, Fluorescence
 Fluorescence Microscopy
 Intro to Single-Molecule Localization Microscopy
 Lab experience and Q&A

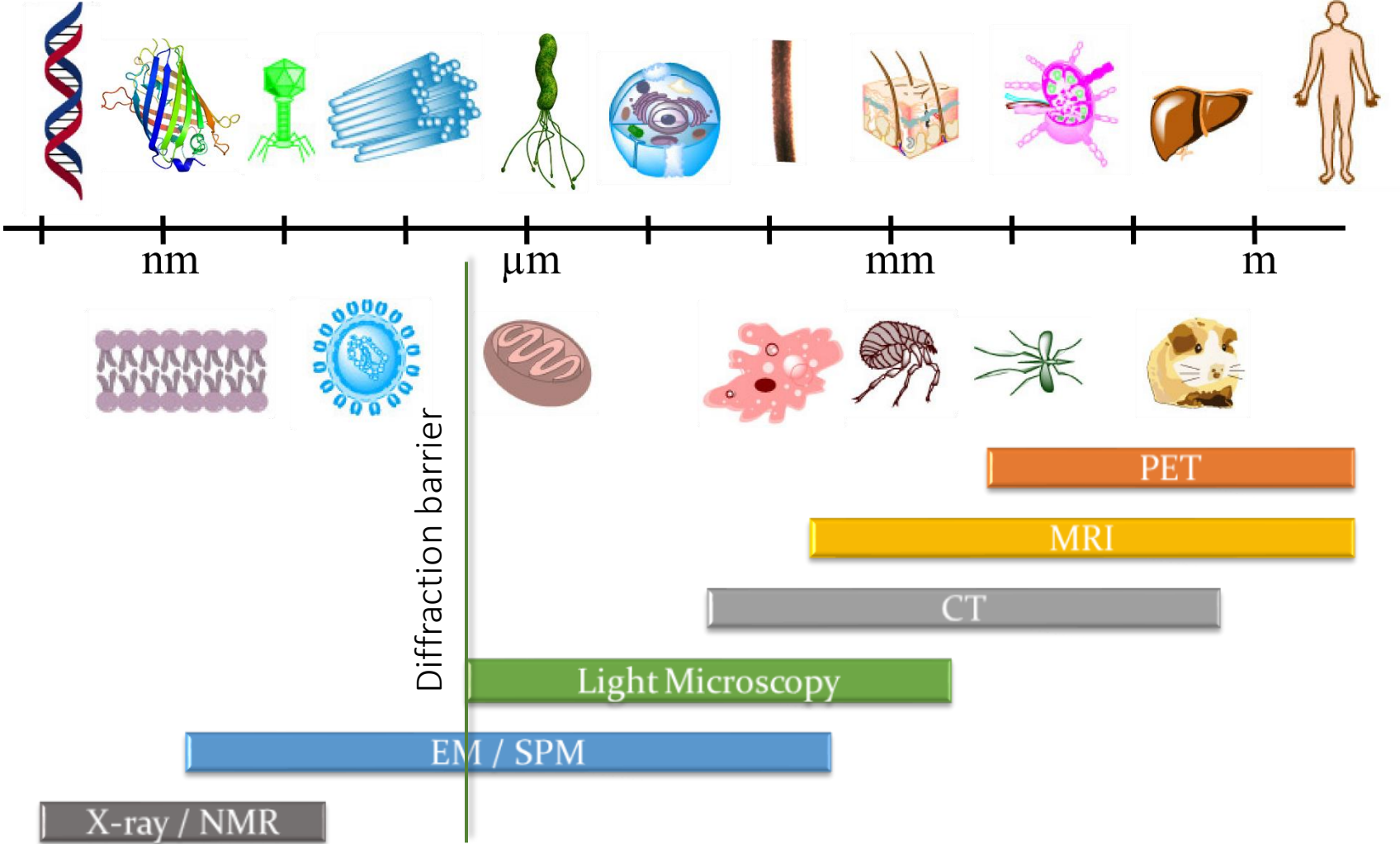
Day 2: Diffraction-limited imaging
 Super-resolution
 Coordinate-stochastic methods: SMLM
 Coordinate-targetted methods: STED
 New methods for sub-10 nm resolution
 Discussion

Super-resolution Fluorescence Microscopy

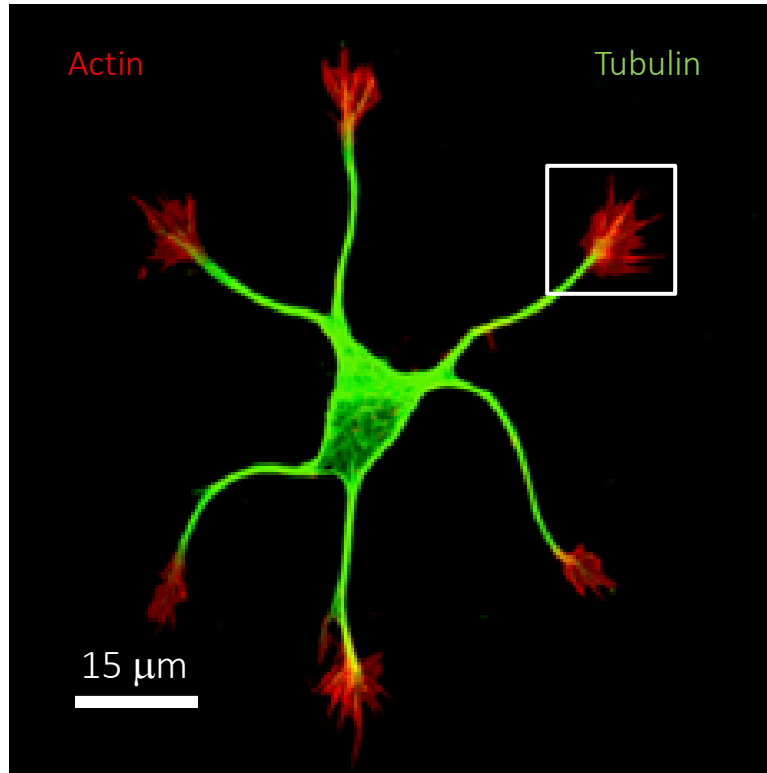
a.k.a.

Fluorescence Nanoscopy

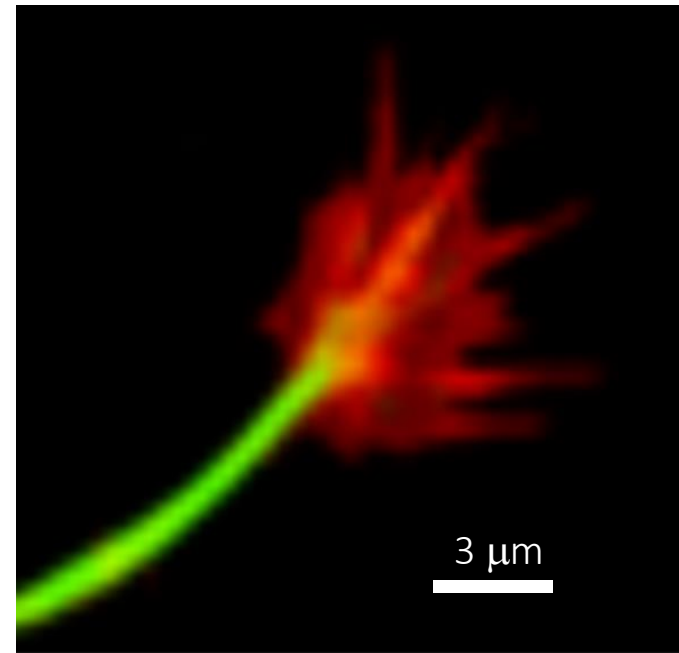
Imaging tools and their spatial resolution



Fluorescence microscopy

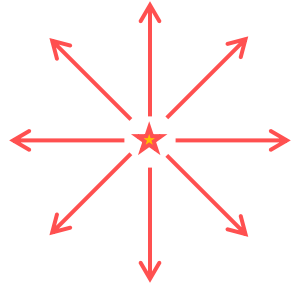


Diffraction-limited imaging

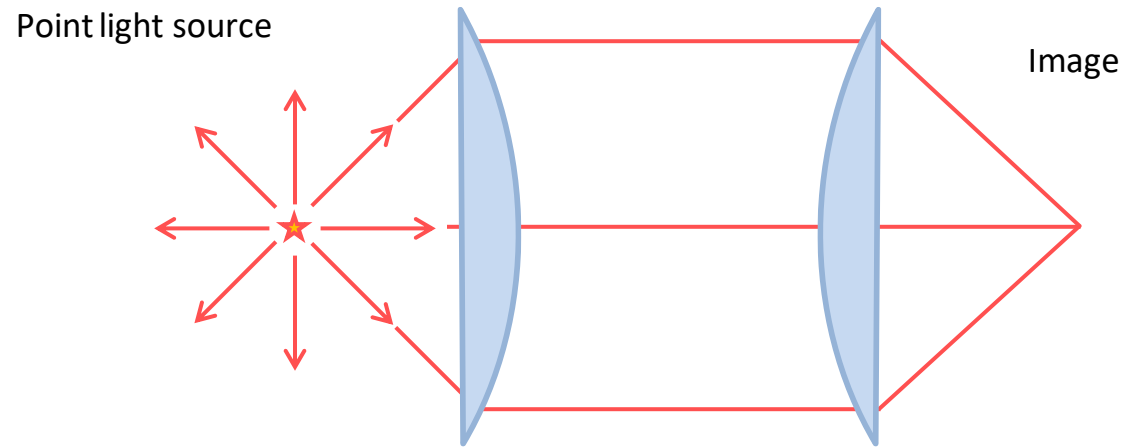


Diffraction-limited imaging

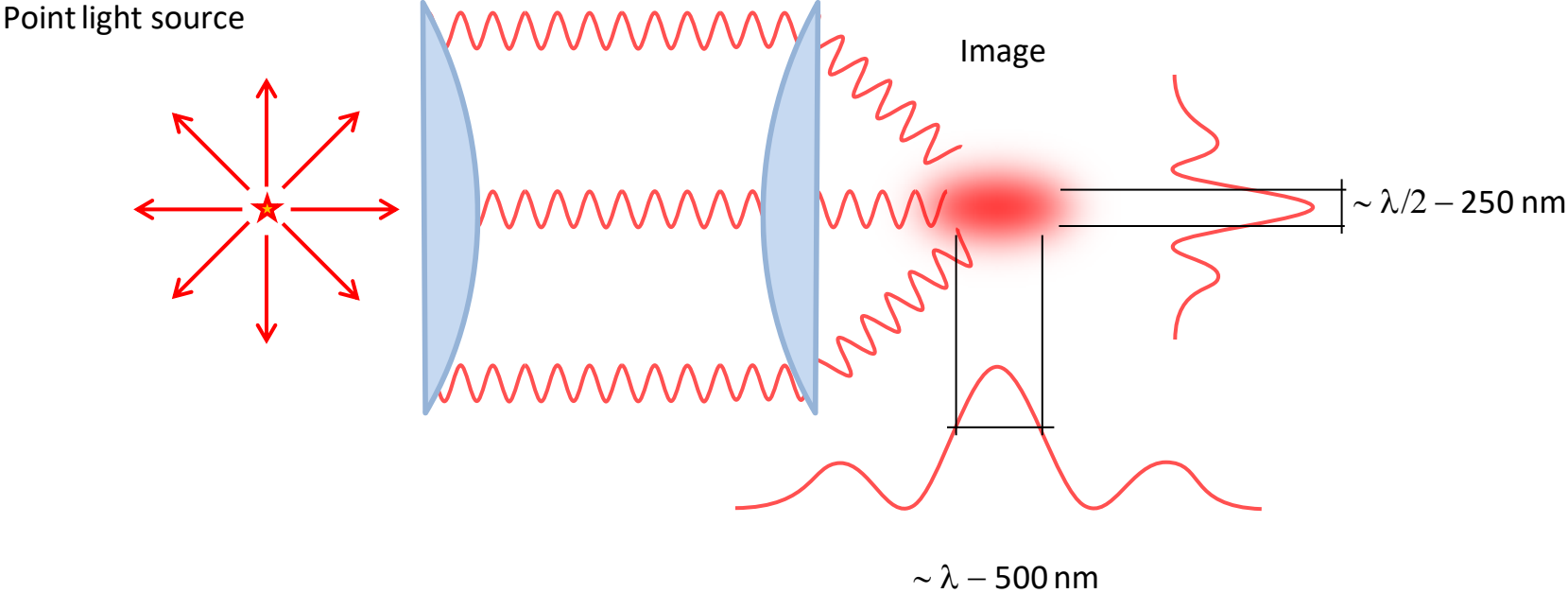
Point light source



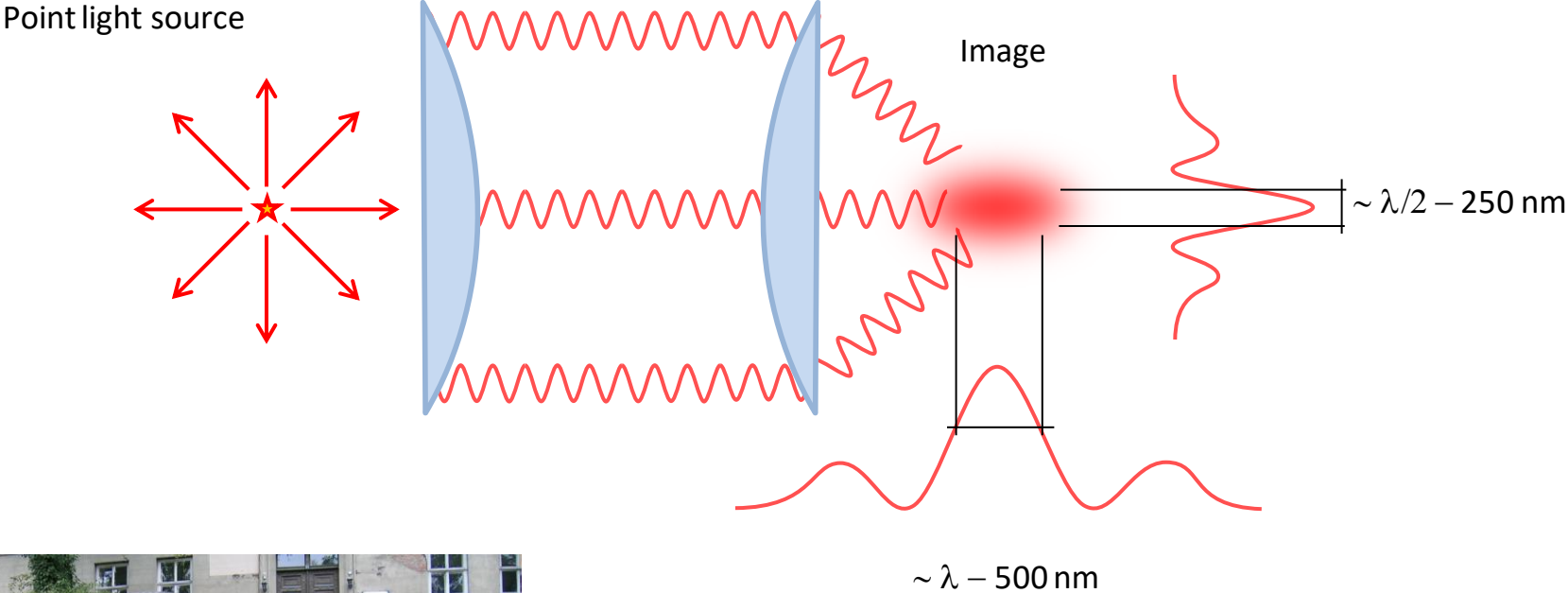
Diffraction-limited imaging



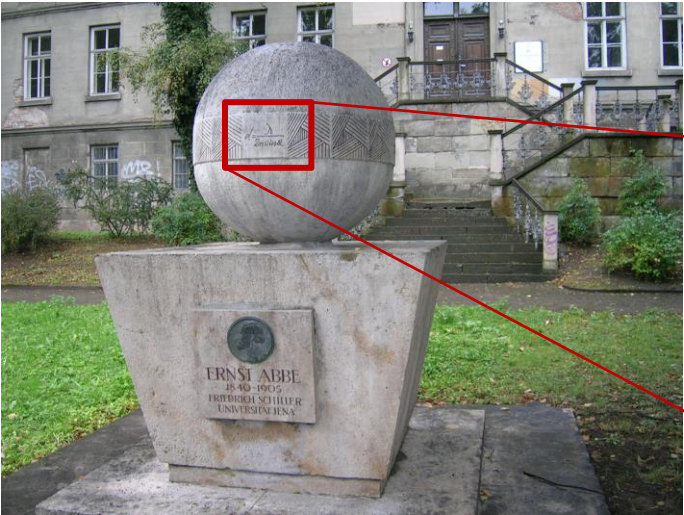
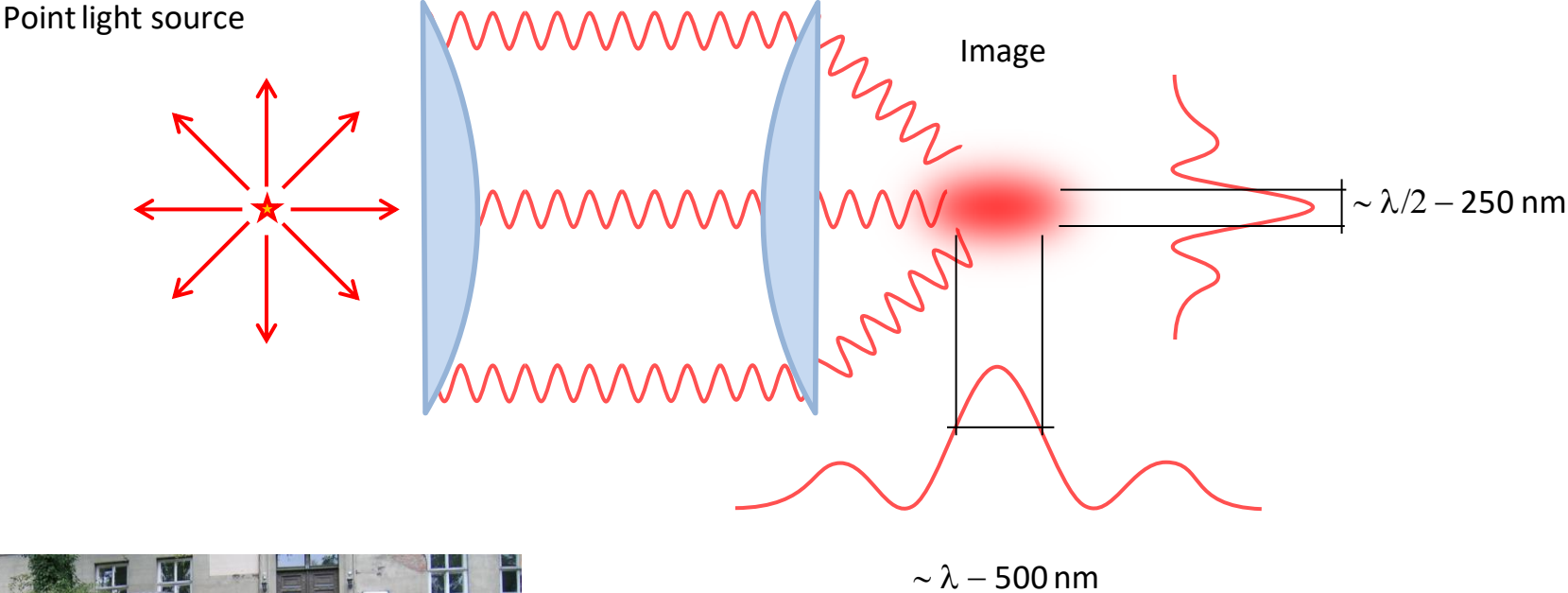
Diffraction-limited imaging



Diffraction-limited imaging



Diffraction-limited imaging



Diffraction-limited imaging



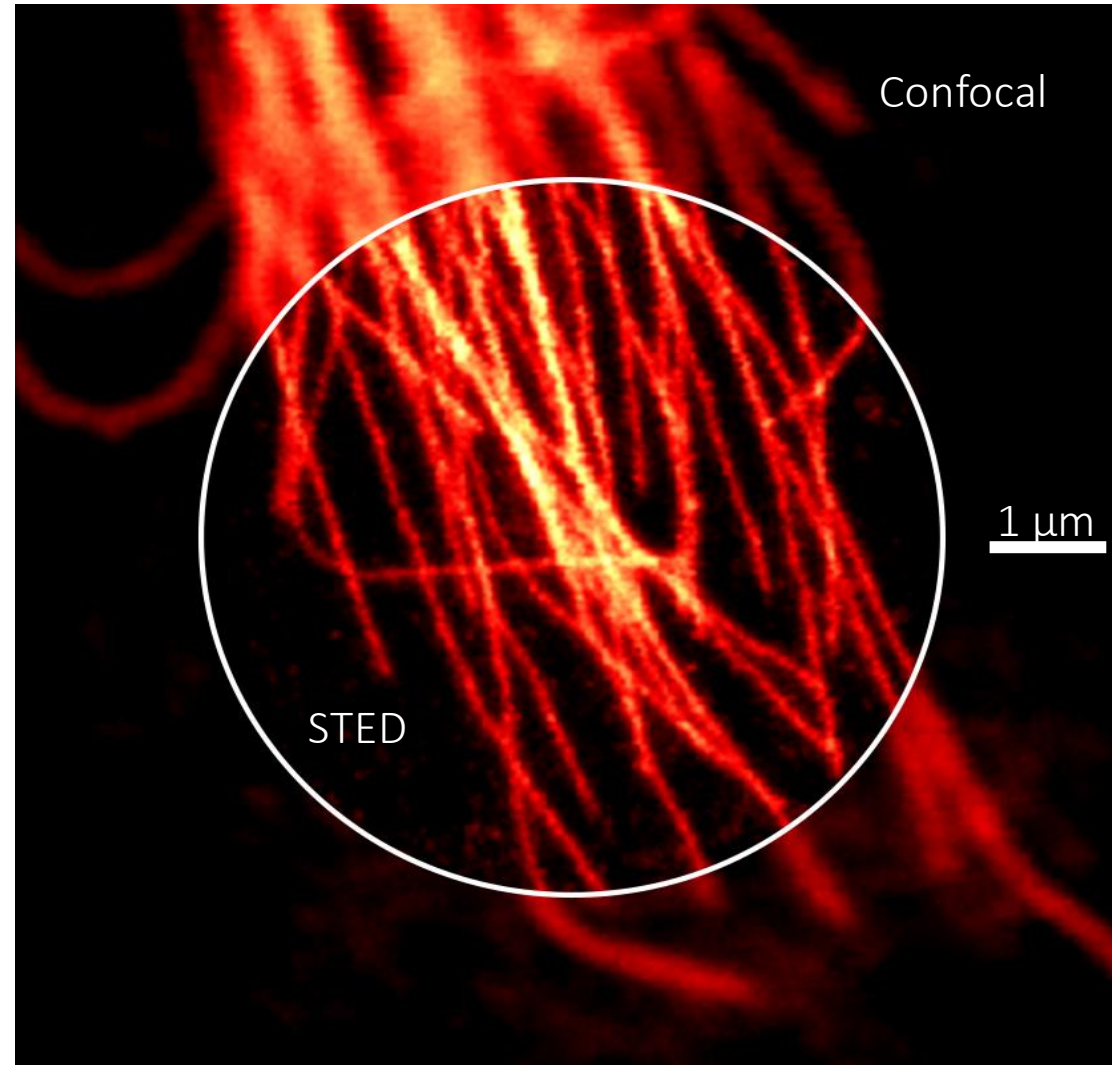
Diffraction-limited imaging



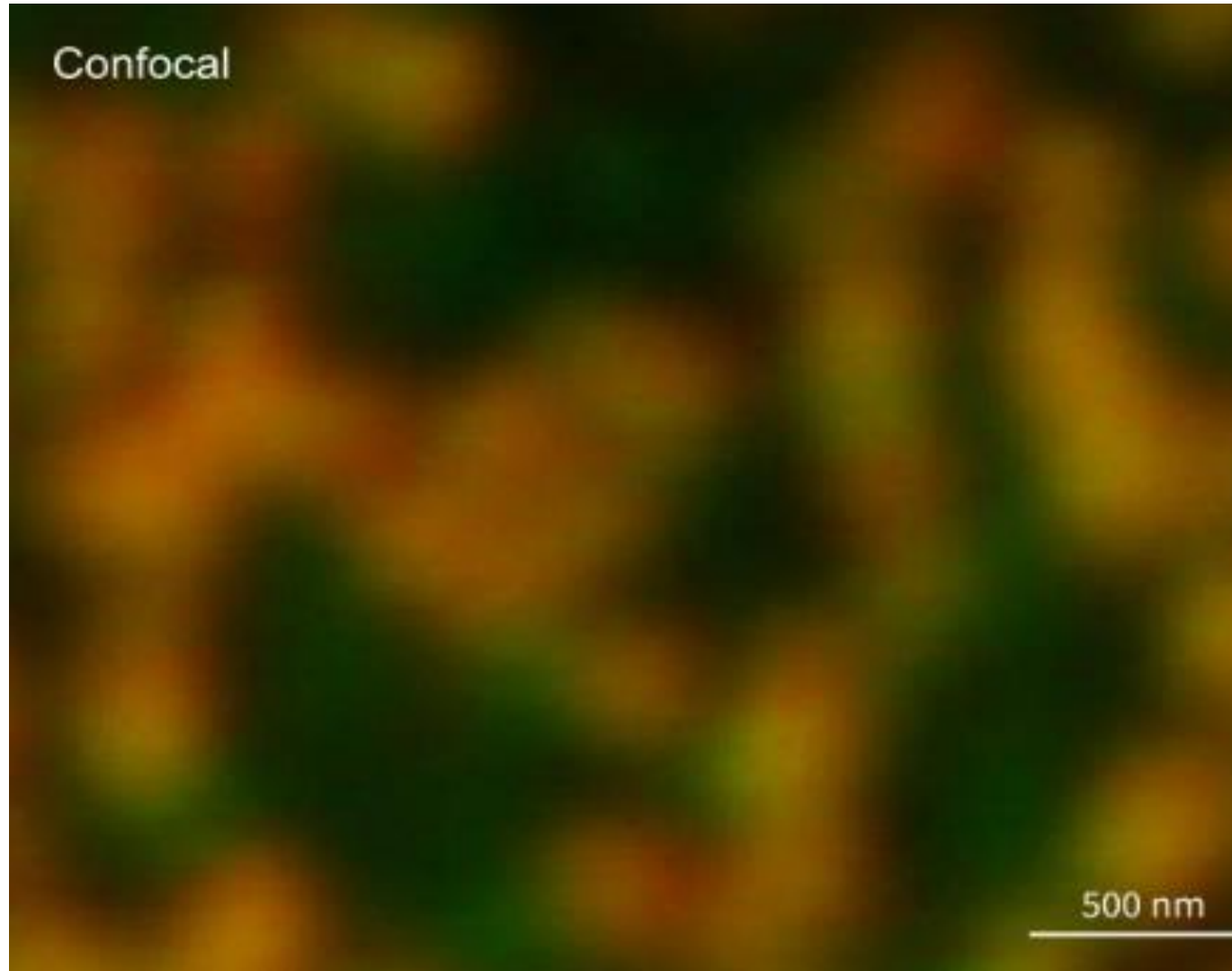
Diffraction-limited imaging



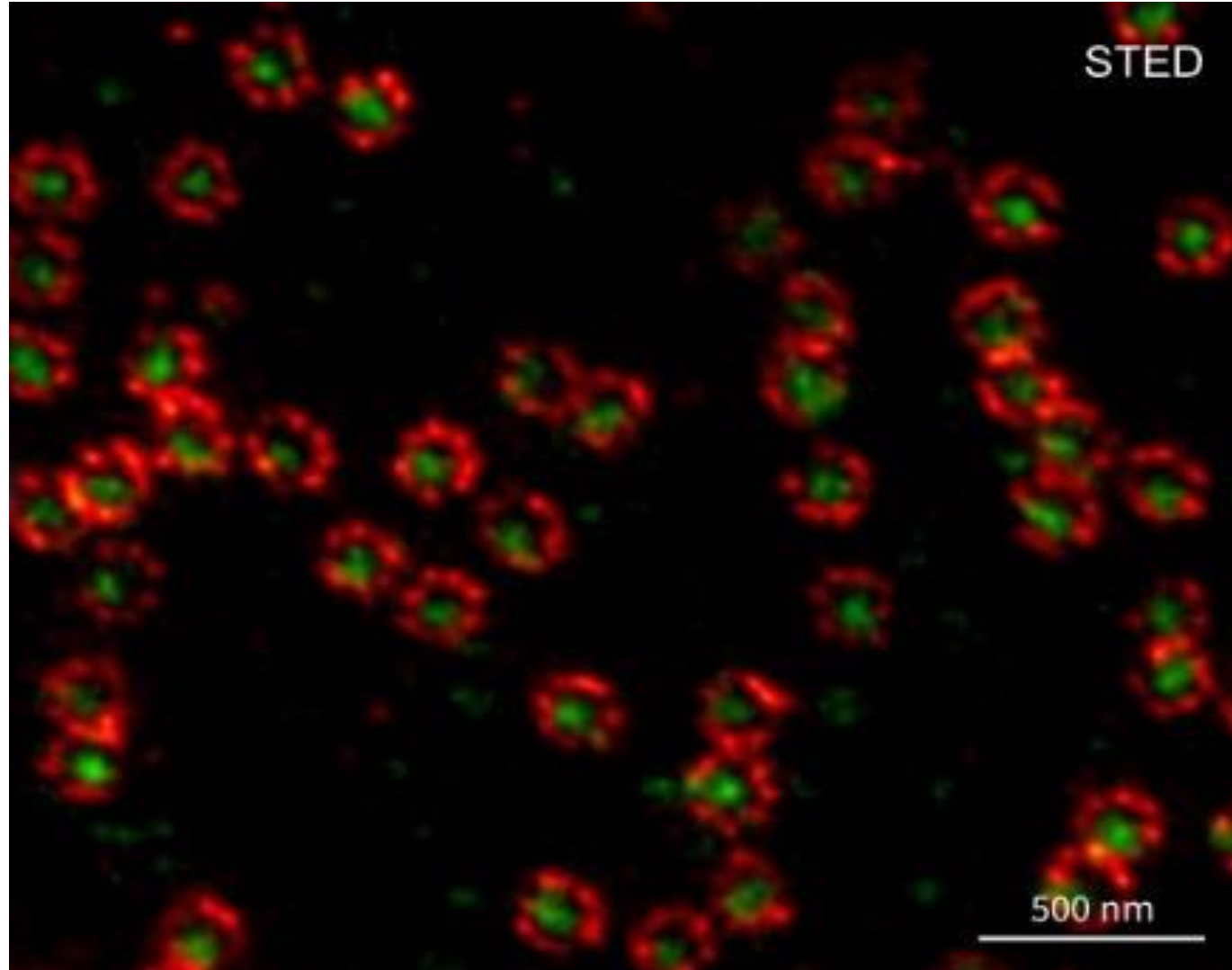
Super-resolution fluorescence microscopy



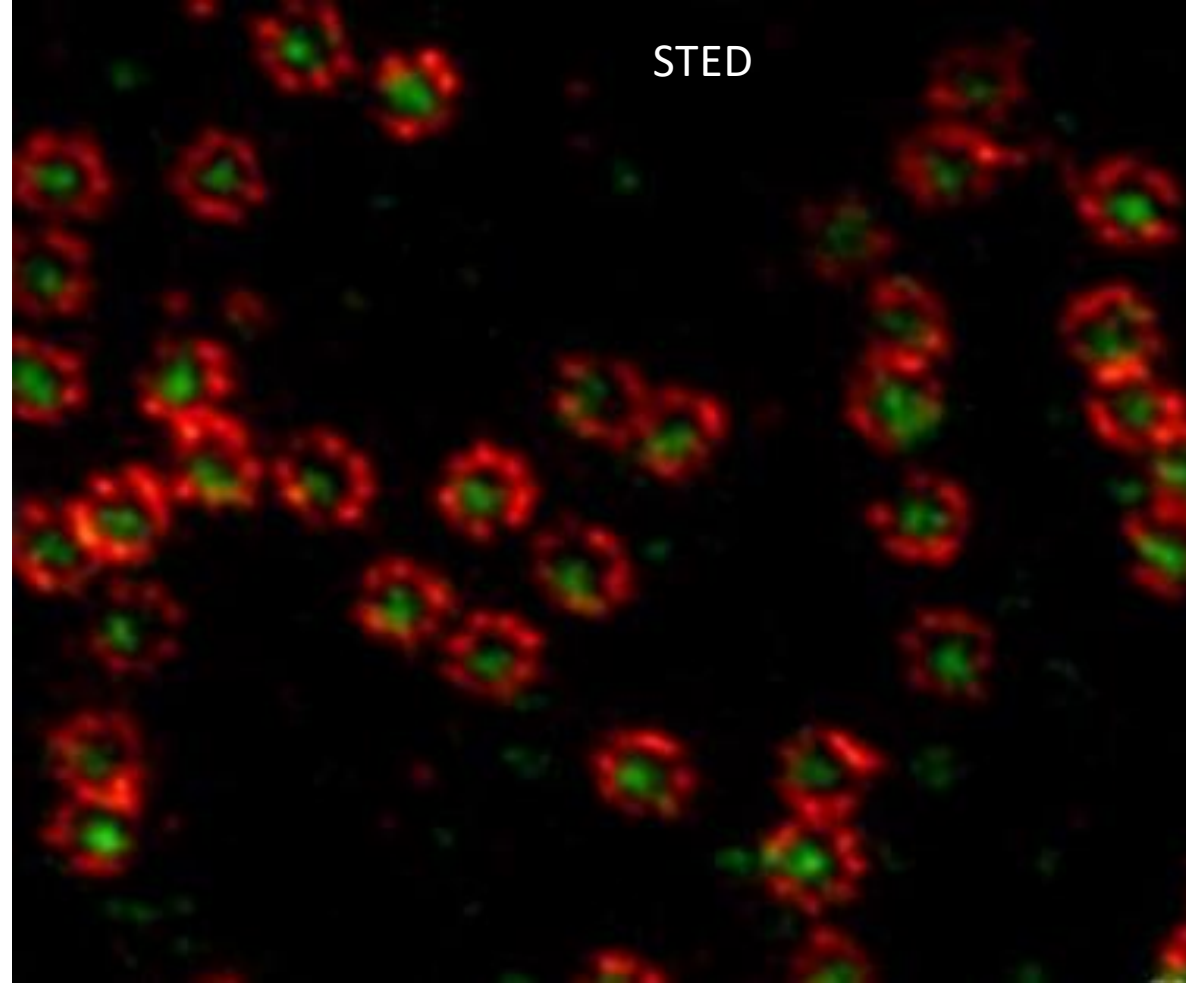
Super-resolution fluorescence microscopy



Super-resolution fluorescence microscopy

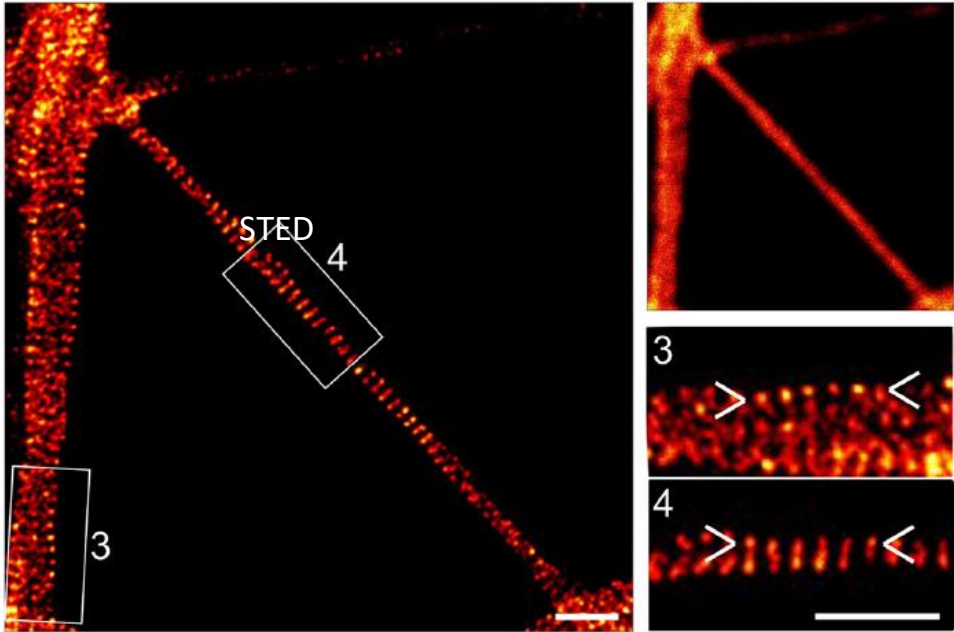
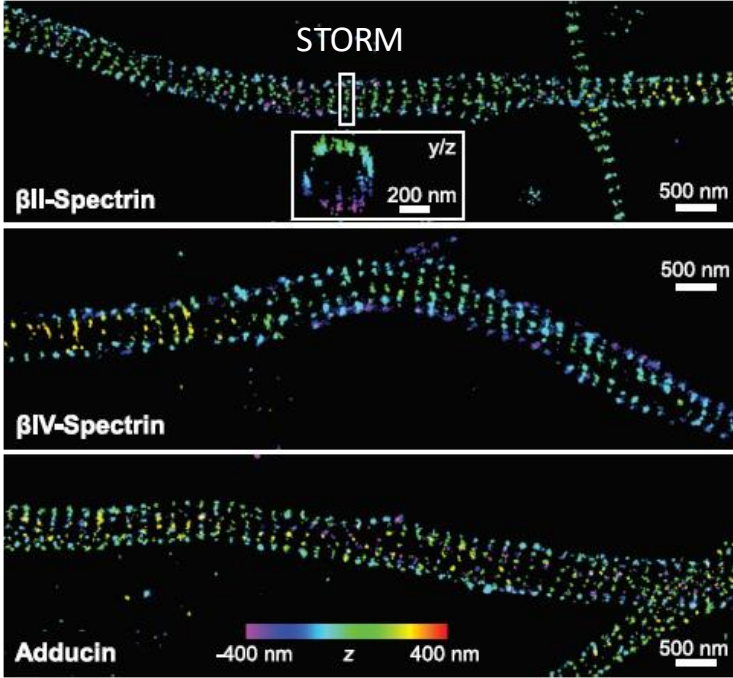


Super-resolution Fluorescence Microscopy

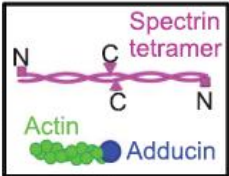
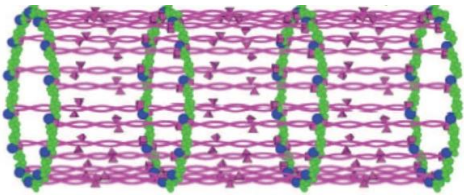


Super-resolution fluorescence microscopy

Membrane-associated Periodic Skeleton (MPS) of neurons

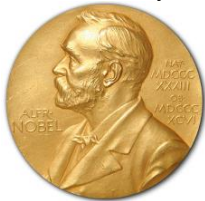


Unsain et al. *Scientific Reports* 2018 18 3007



Xiaowei Zhuang Lab
 Xu et al. *Science* 2013, 339, 452–456.

2014
 Chemistry



Stefan W. Hell



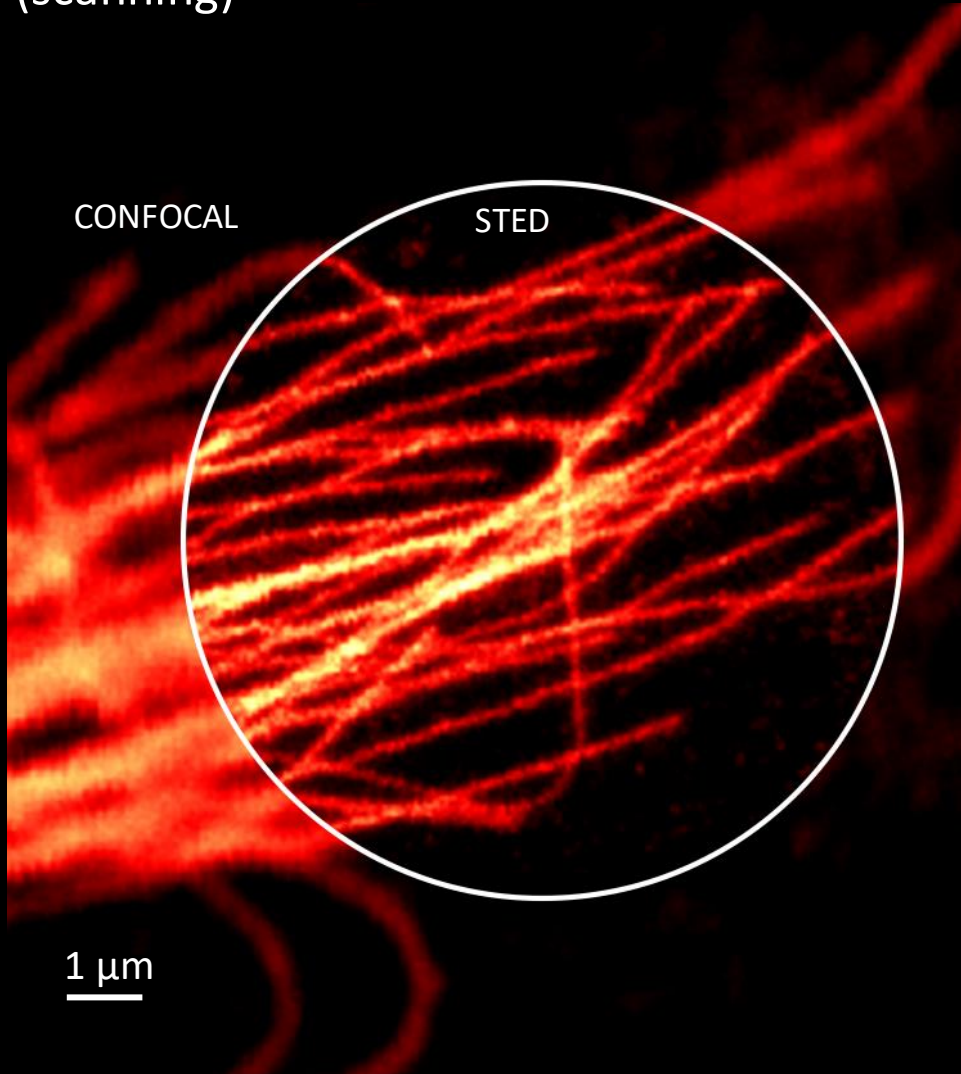
Eric Betzig



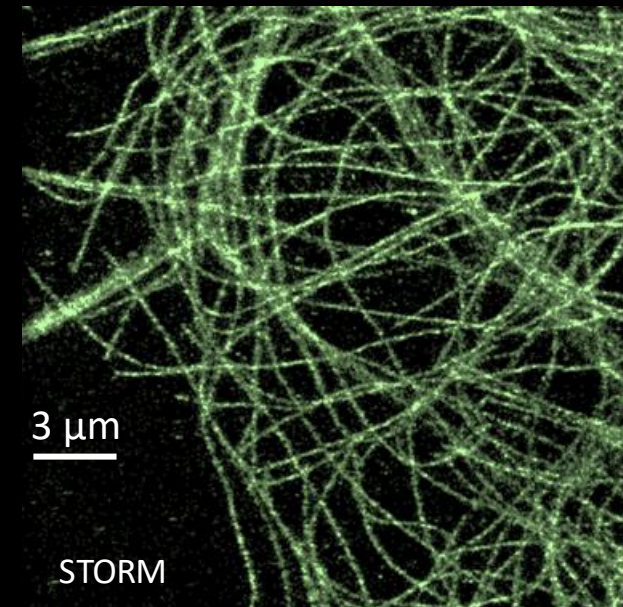
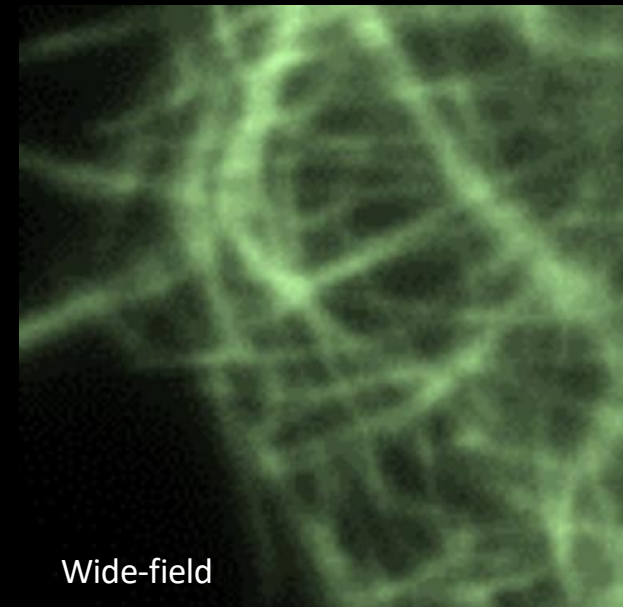
William E. Moerner

Super-resolution fluorescence microscopy

Coordinate-targeted nanoscopy
(scanning)



Coordinate-stochastic nanoscopy
(wide-field)

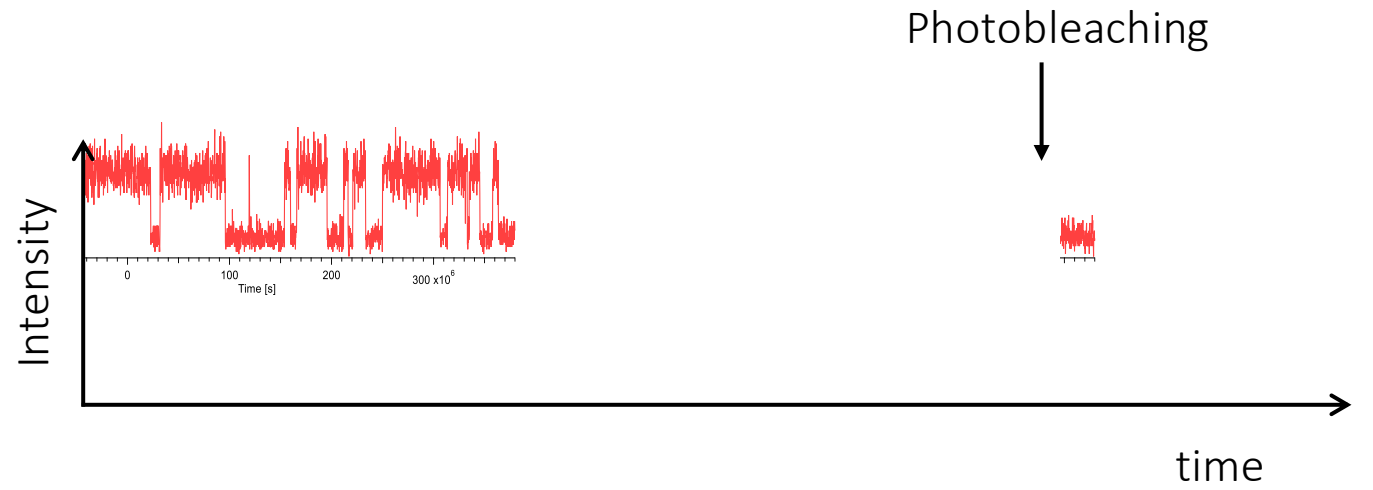
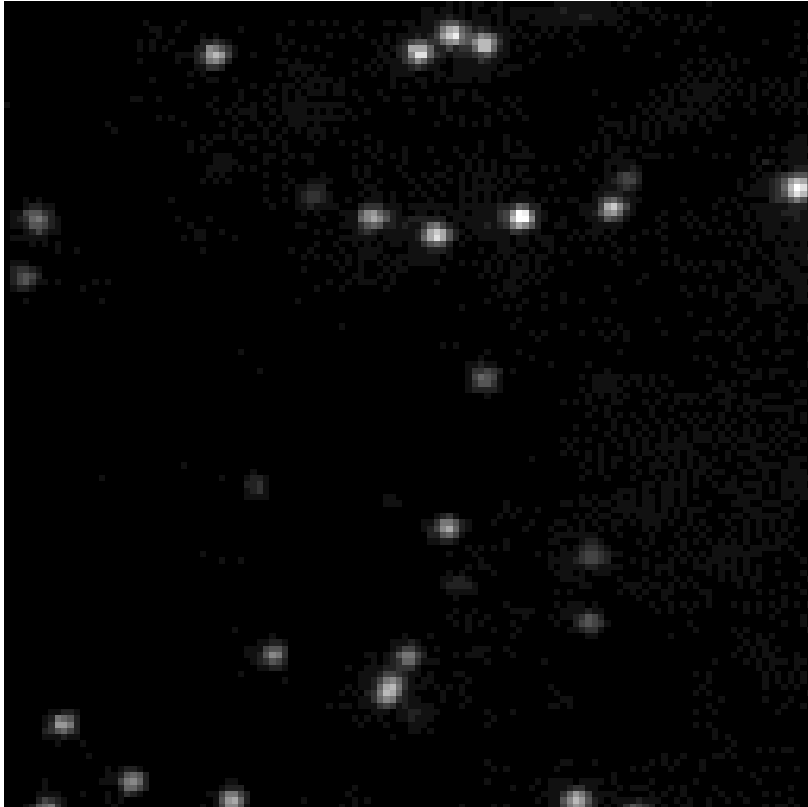


Coordinate-Stochastic Super-resolution Microscopy

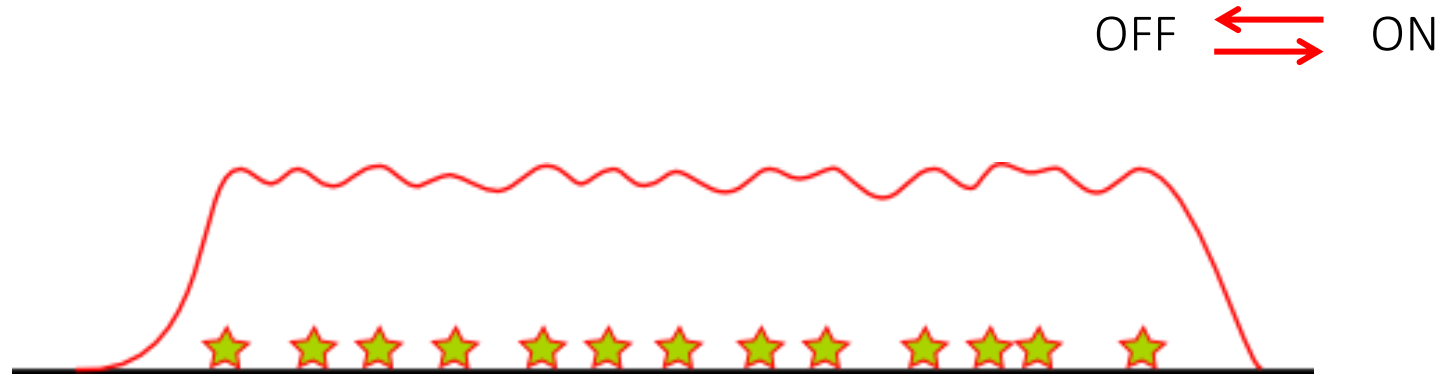
a.k.a.

Single-Molecule Localization Microscopy (SMLM)

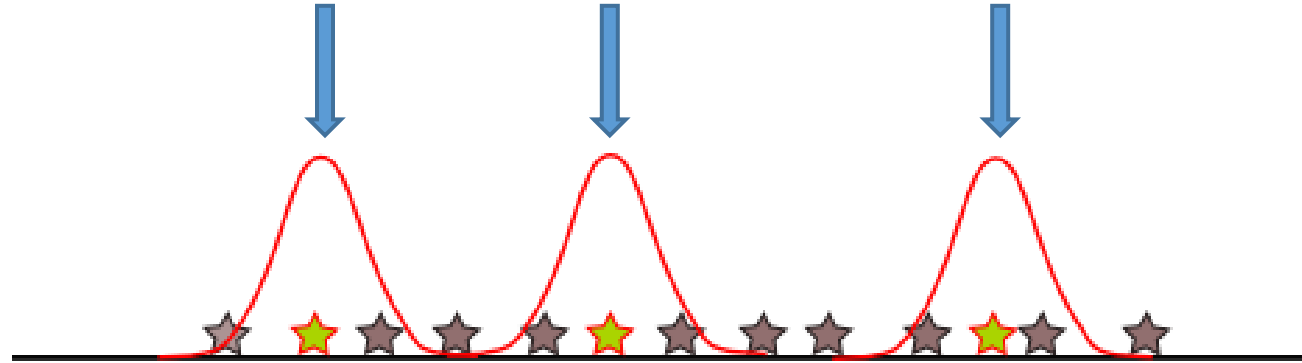
Single-molecule fluorescence blinking



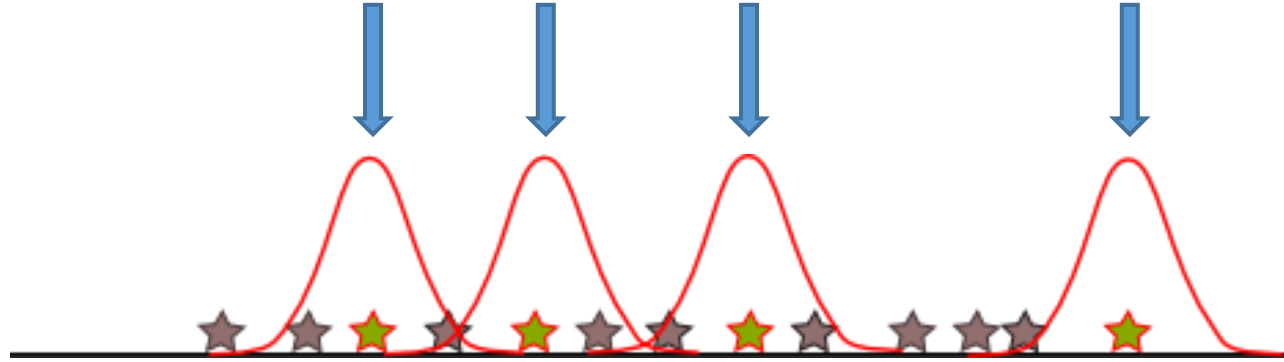
Single-Molecule Localization Microscopy (SMLM)



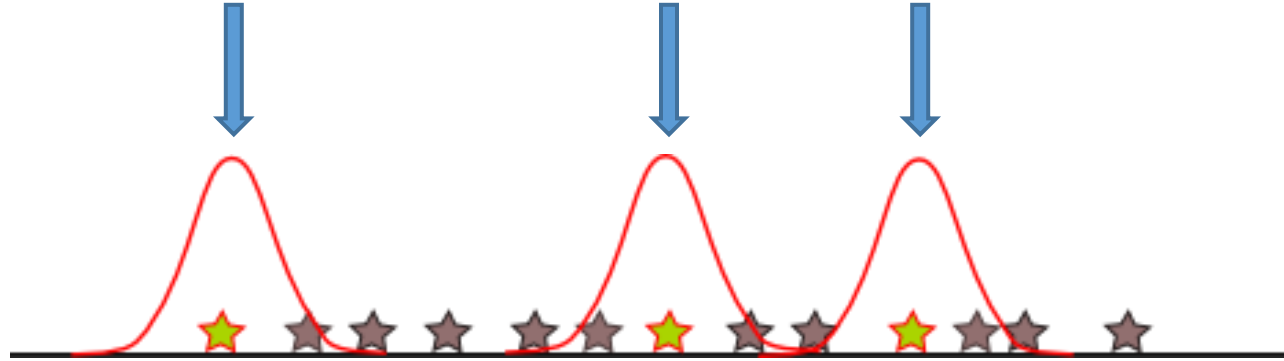
Single-Molecule Localization Microscopy (SMLM)



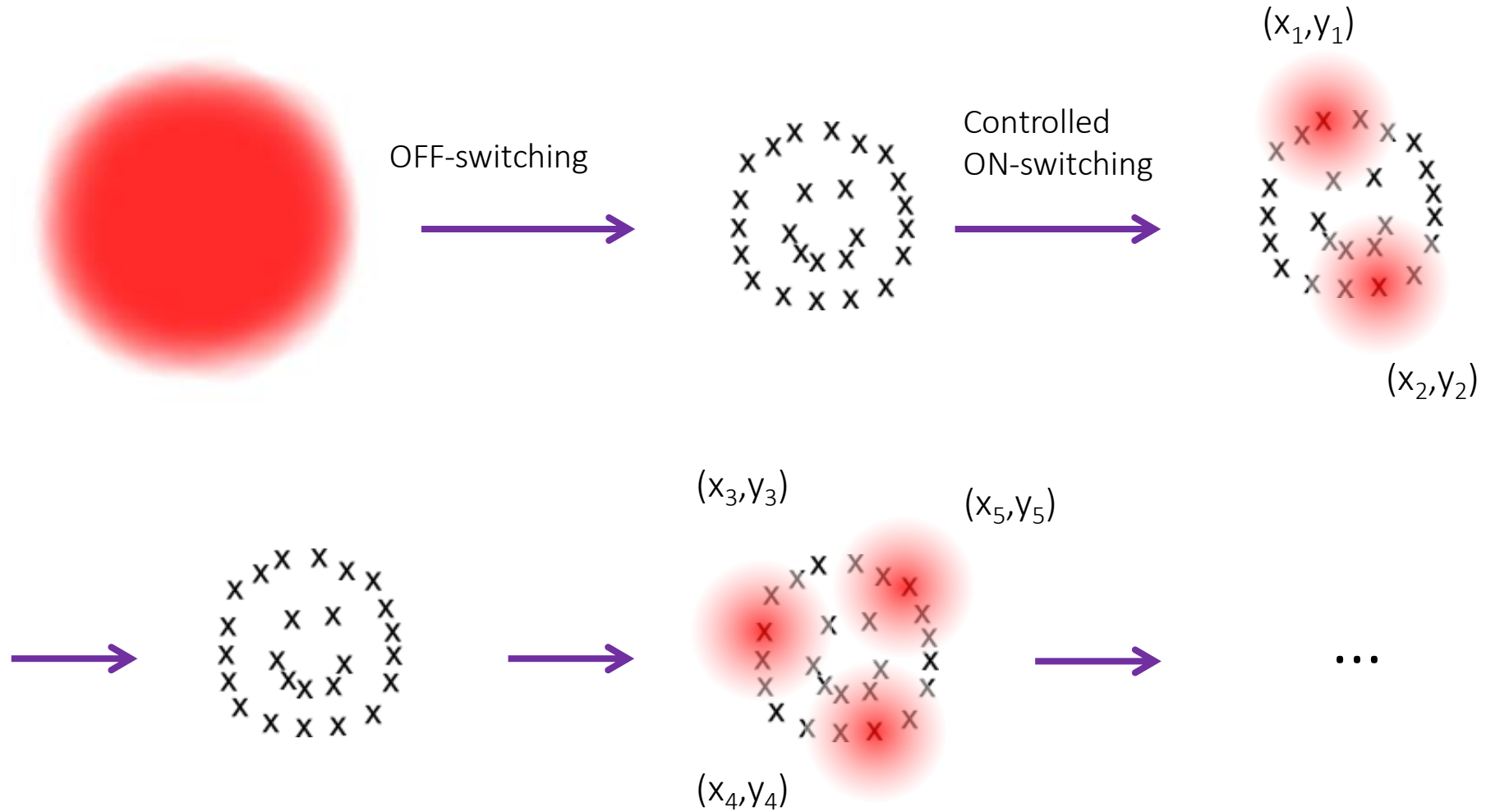
Single-Molecule Localization Microscopy (SMLM)



Single-Molecule Localization Microscopy (SMLM)



Single-Molecule Localization Microscopy (SMLM)



Single-Molecule Localization Microscopy (SMLM)

(x_1, y_1)

(x_2, y_2)

(x_3, y_3)

(x_4, y_4)

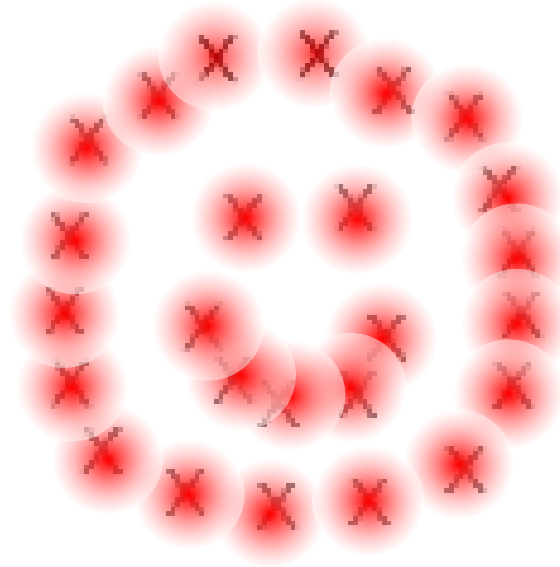
(x_5, y_5)

(x_6, y_6)

(x_7, y_7)

(x_8, y_8)

...

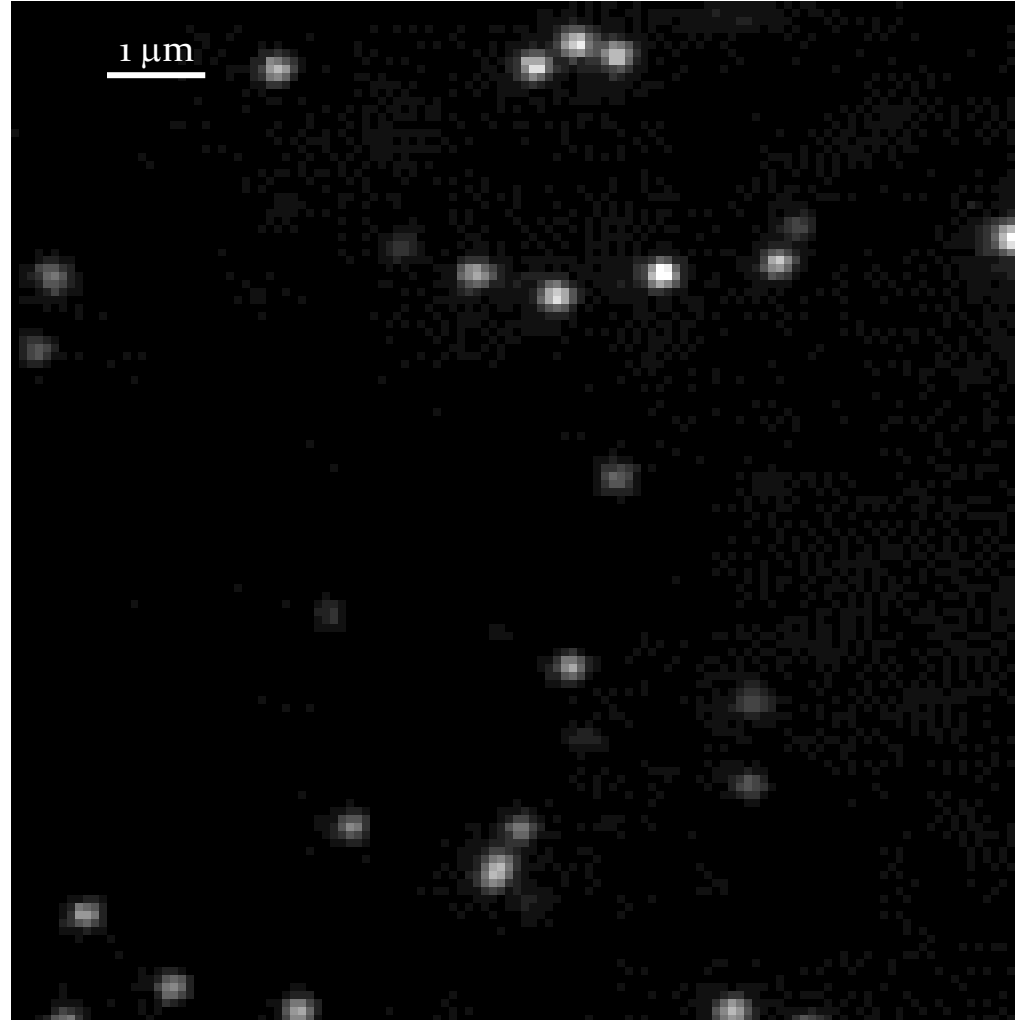


STORM – Stochastic Optical Reconstruction Microscopy

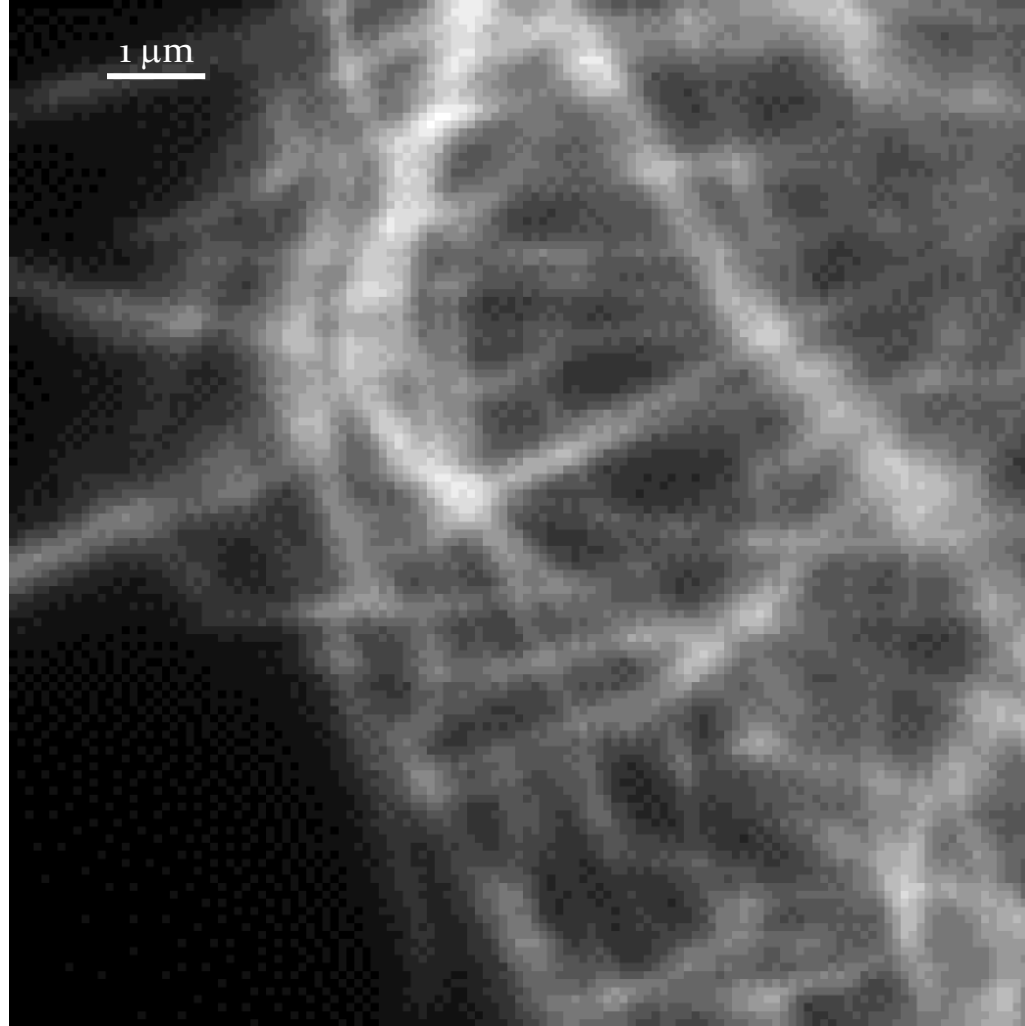
PALM – Photo-Activated Localization Microscopy

PAINT – Points Accumulation for Imaging in Nanoscale Topography

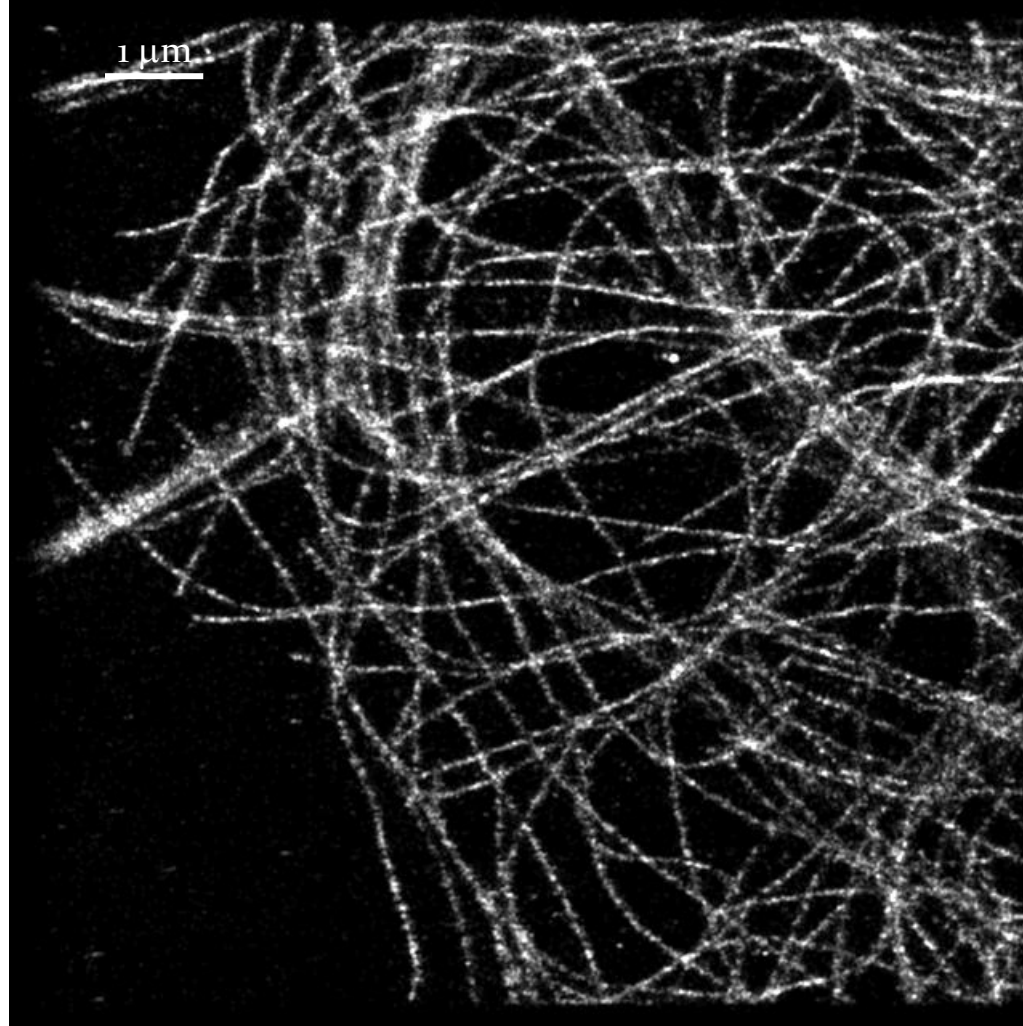
Single-Molecule Localization Microscopy (SMLM)



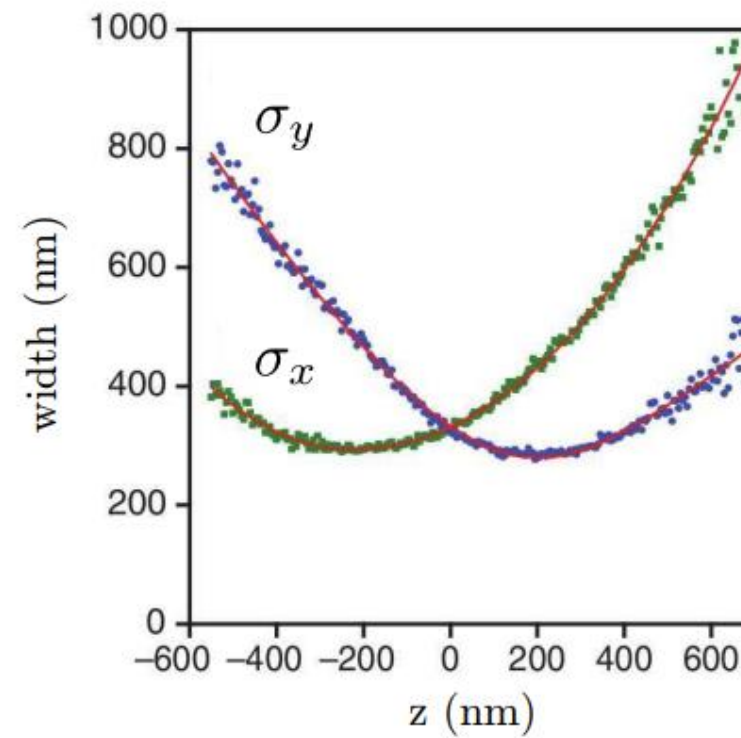
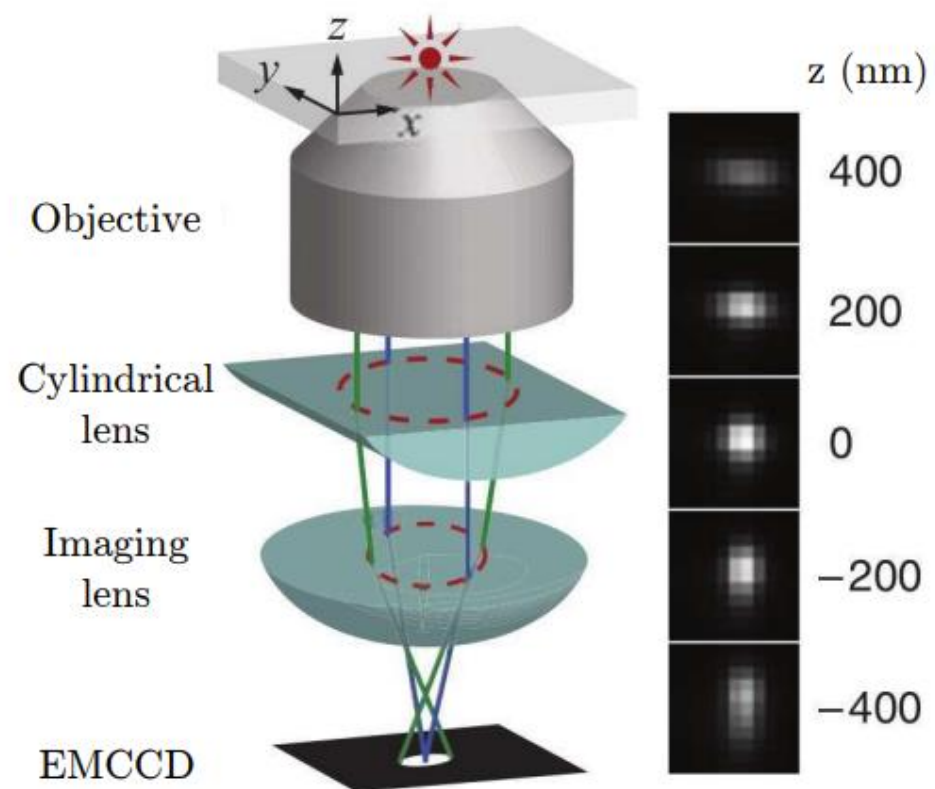
Single-Molecule Localization Microscopy (SMLM)



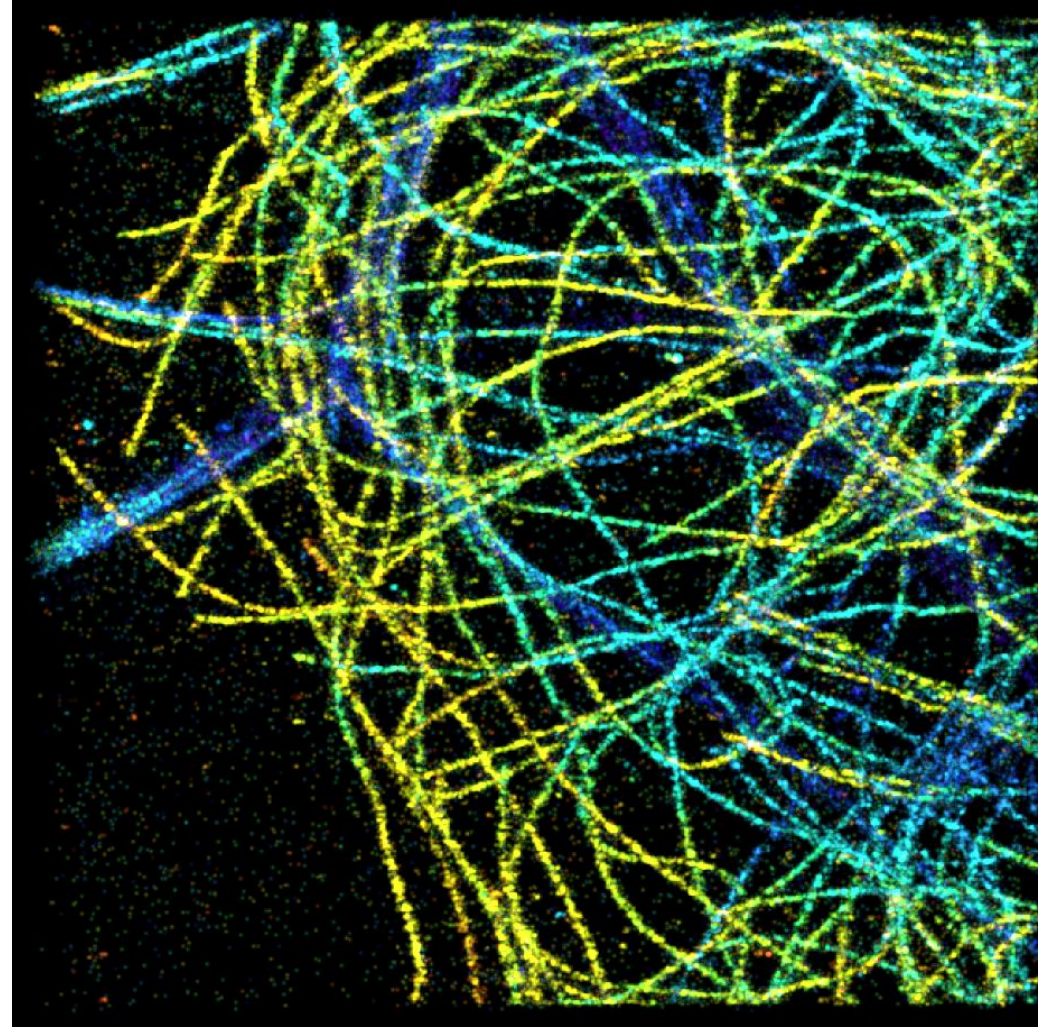
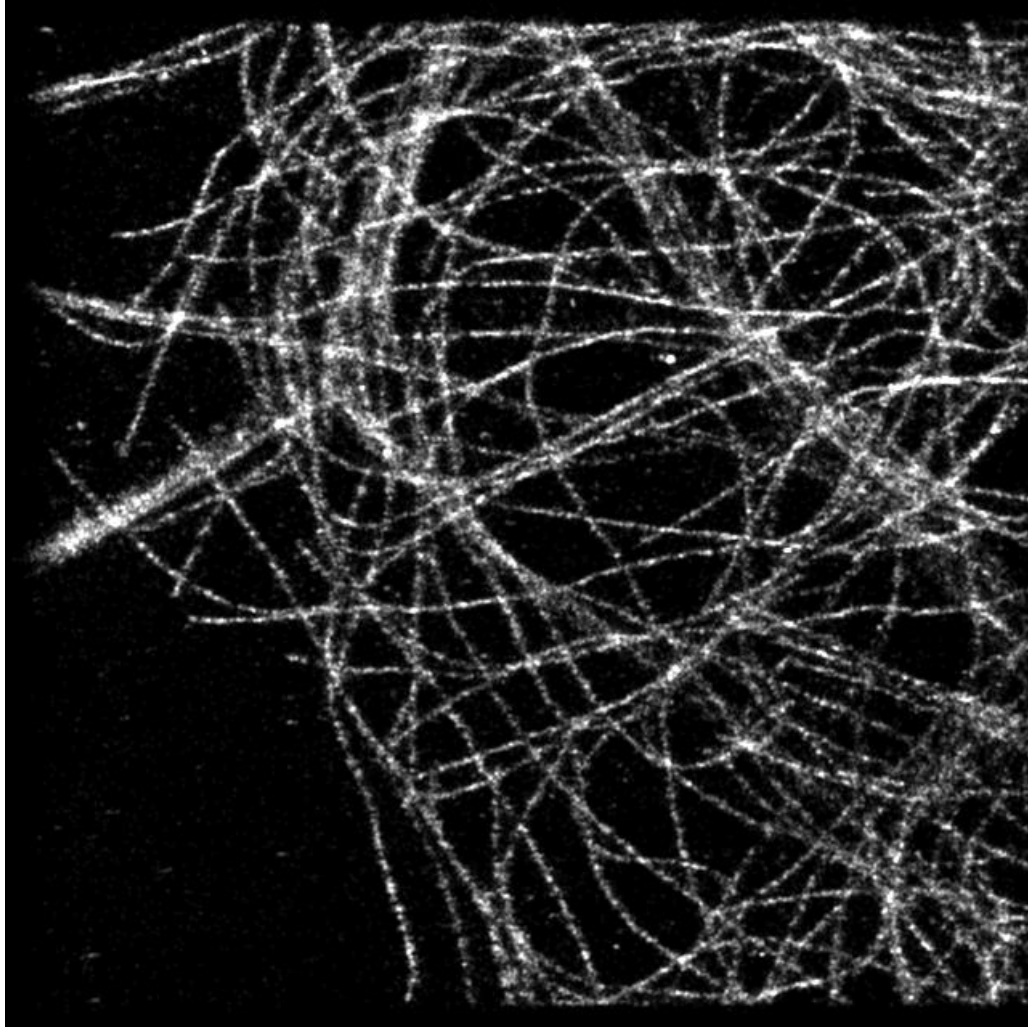
Single-Molecule Localization Microscopy (SMLM)



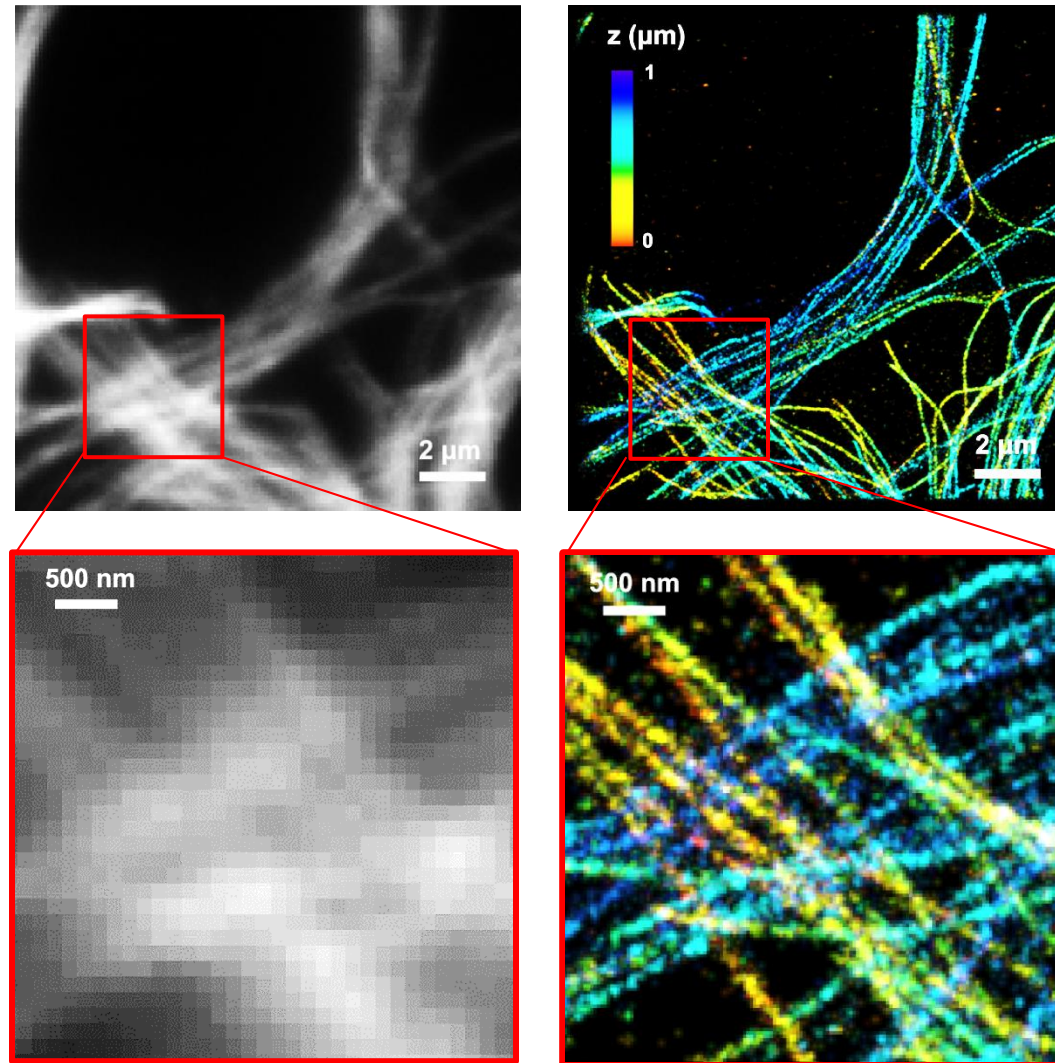
SMLM in 3D



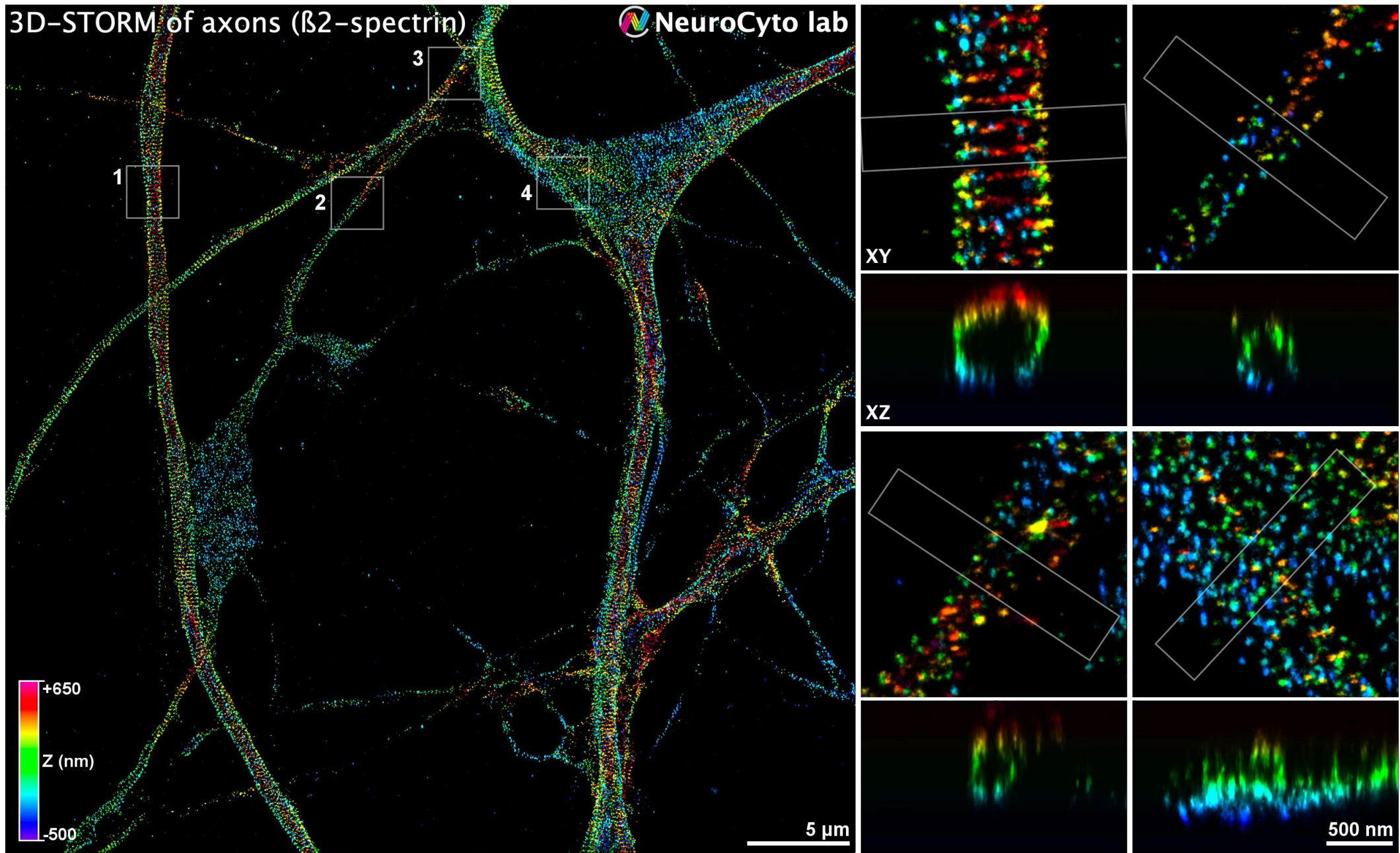
SMLM in 3D



SMLM in 3D



SMLM in 3D

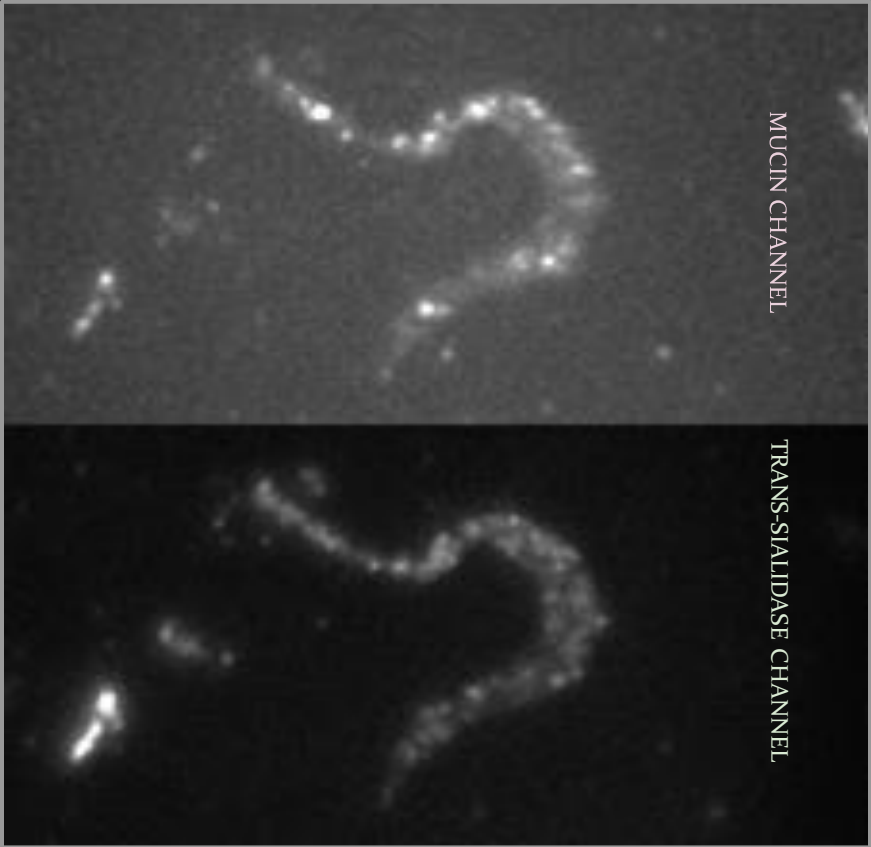


Multicolor SMLM

DIFFRACTION-LIMITED

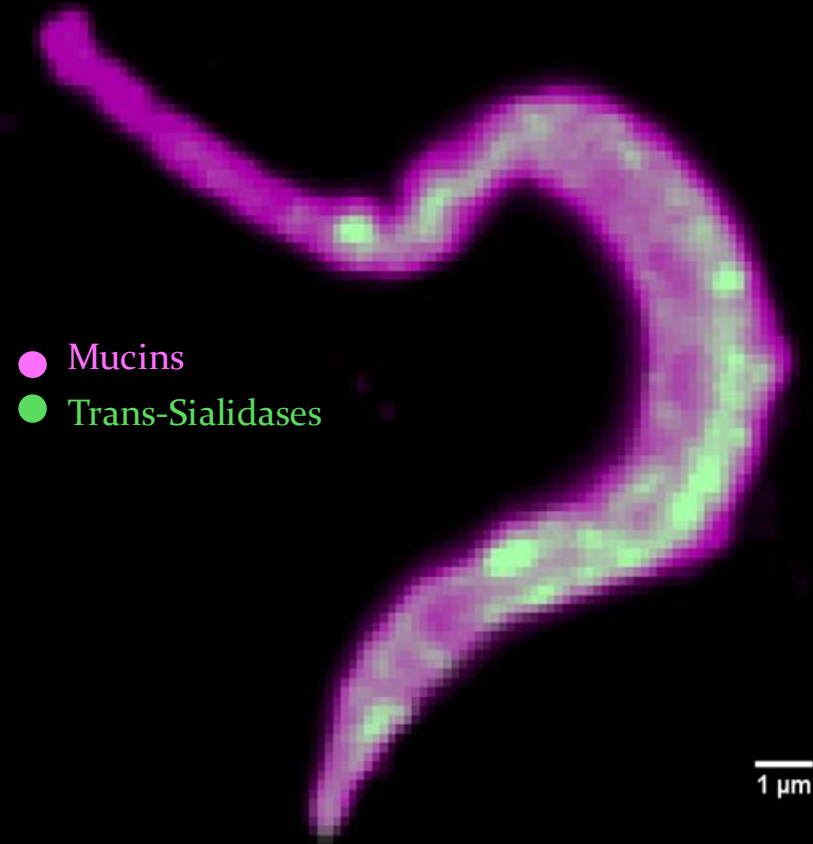


BLINKING SEQUENCE



Multicolor SMLM

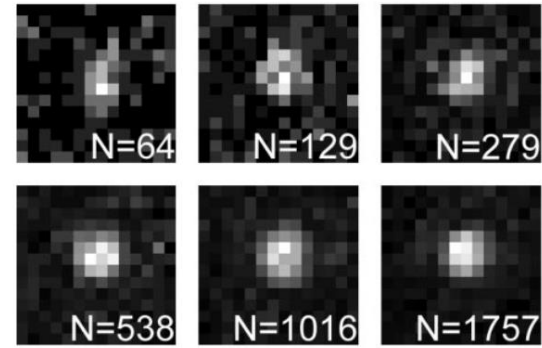
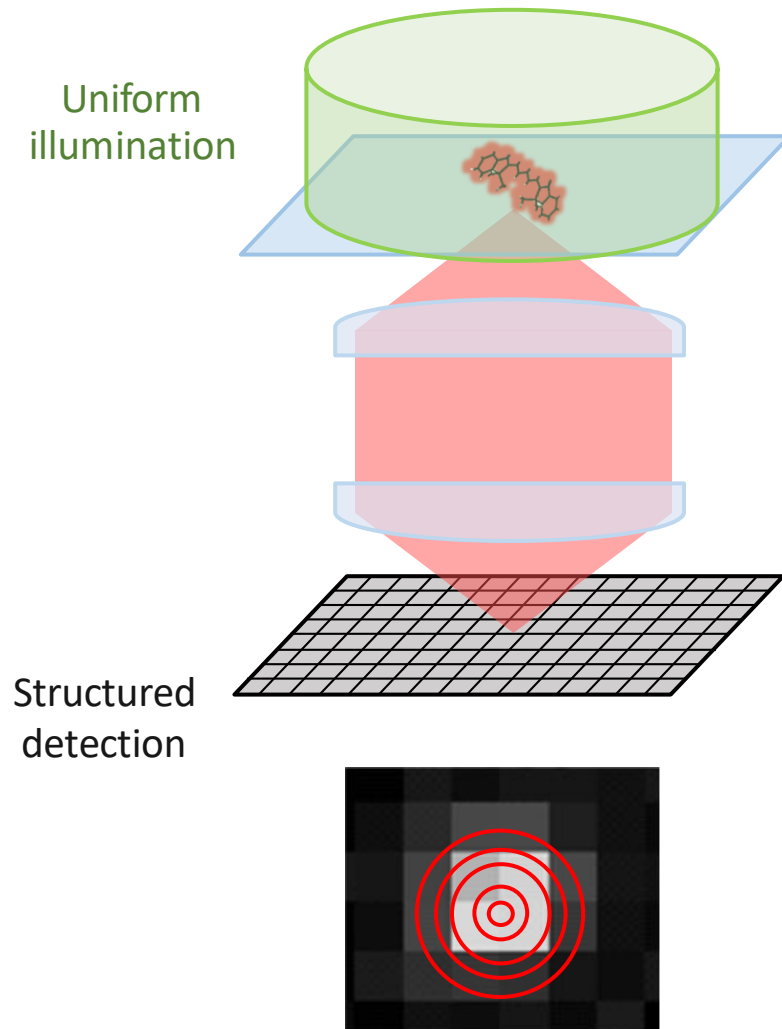
DIFFRACTION-LIMITED



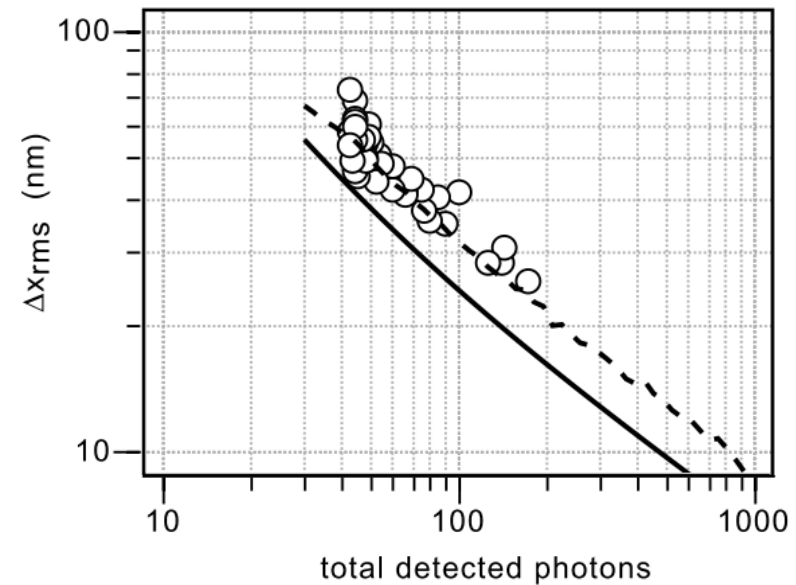
2-COLOR STORM



Camera-based single-molecule localization

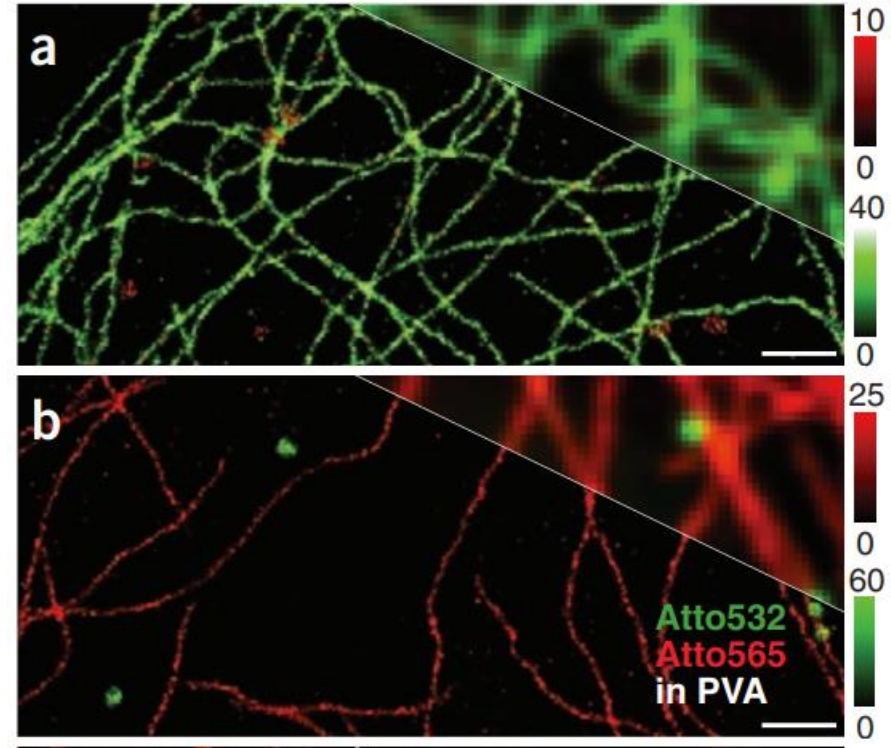
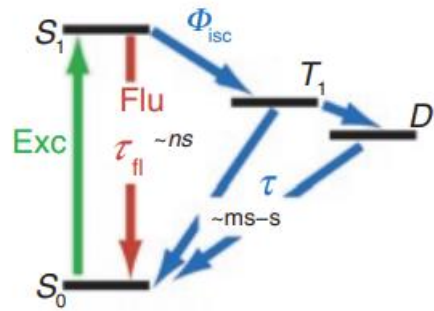


$$\sigma \approx PSF / \sqrt{N}$$

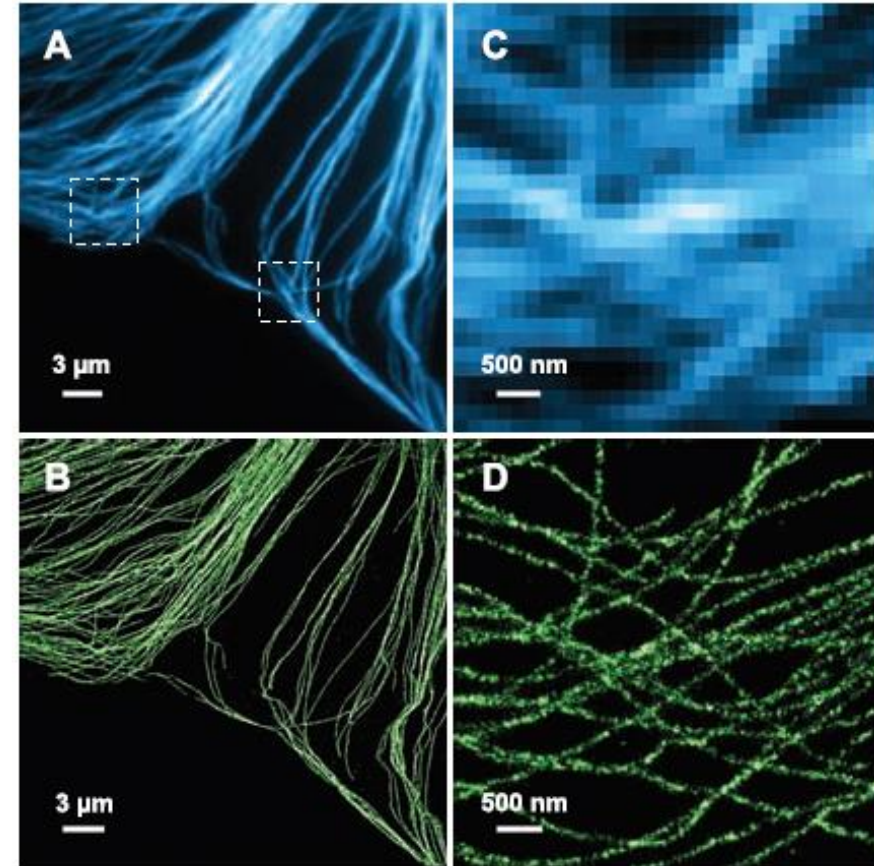
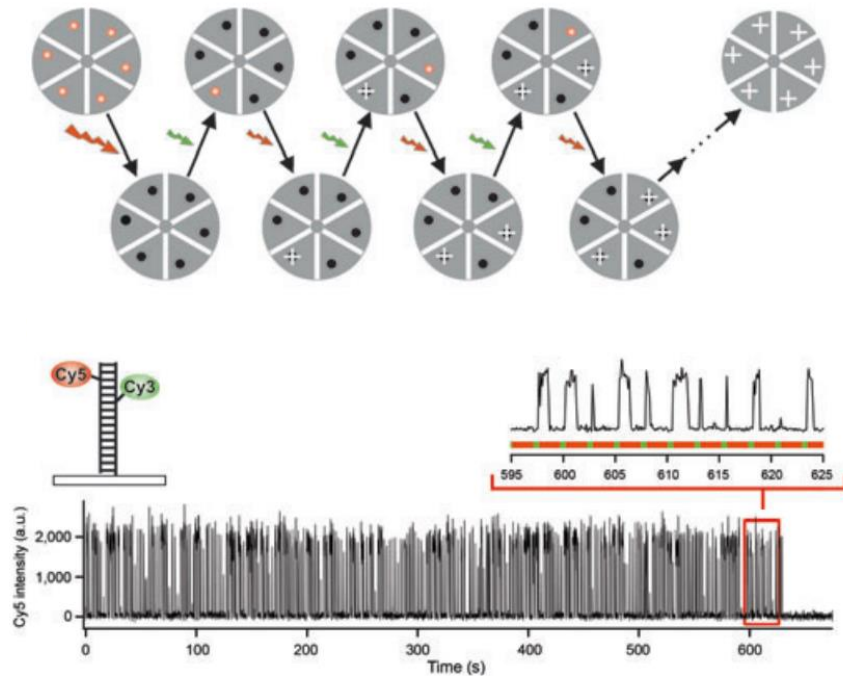


Various SMLM : excursion to the triplet state

ON-OFF by excursion to triplet states

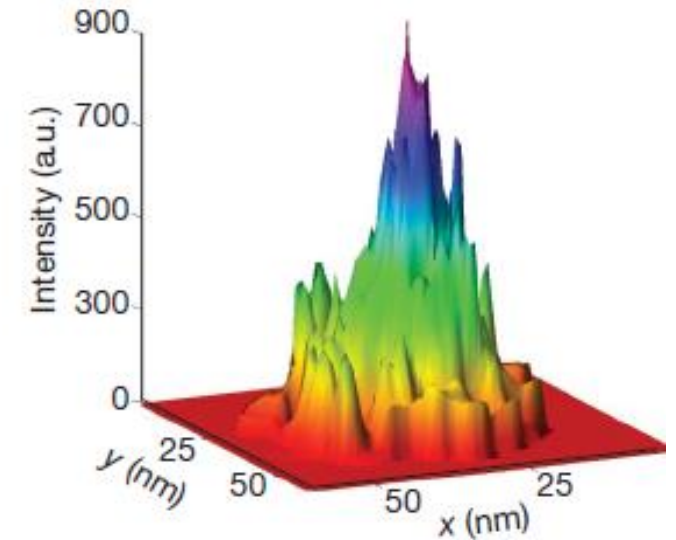
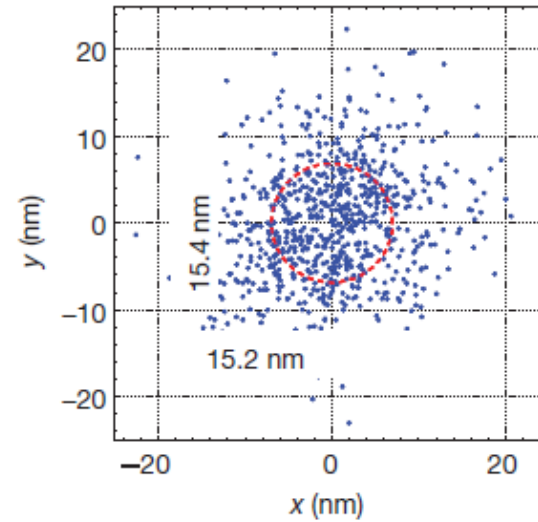
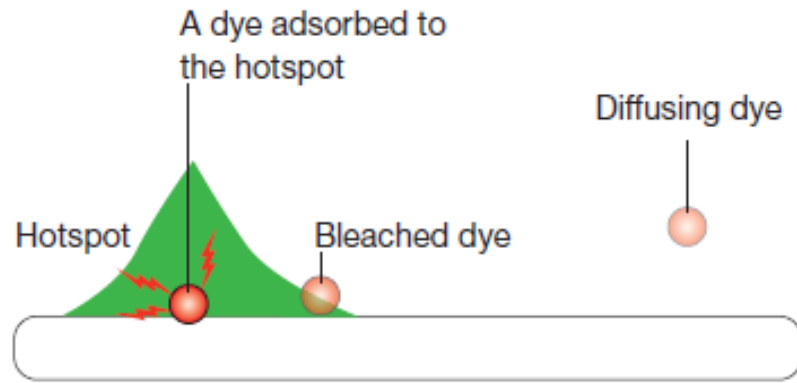


Various SMLM : photochromic fluorophores



Various SMLM : random adsorption PAINT

ON-OFF by adsorption

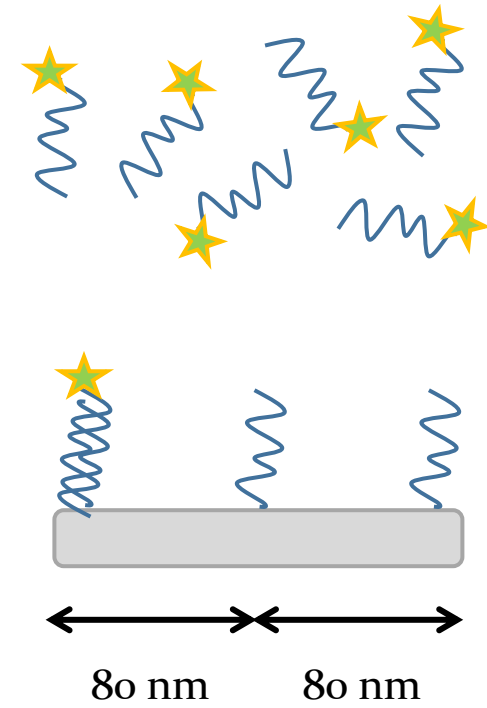
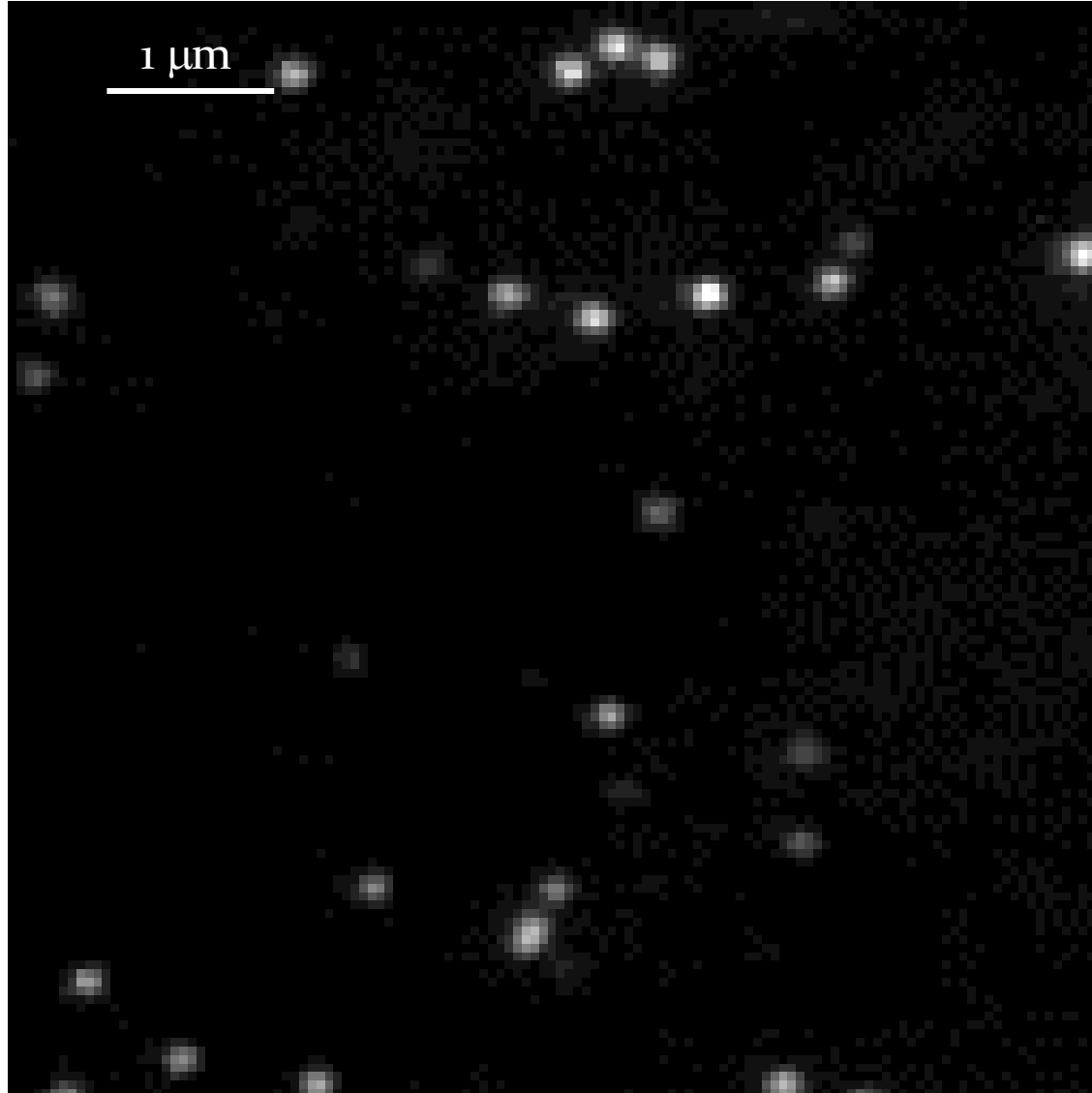


Xiang Zhang Lab - Cang, H. et al. *Nature* 469 (2011) 385-388

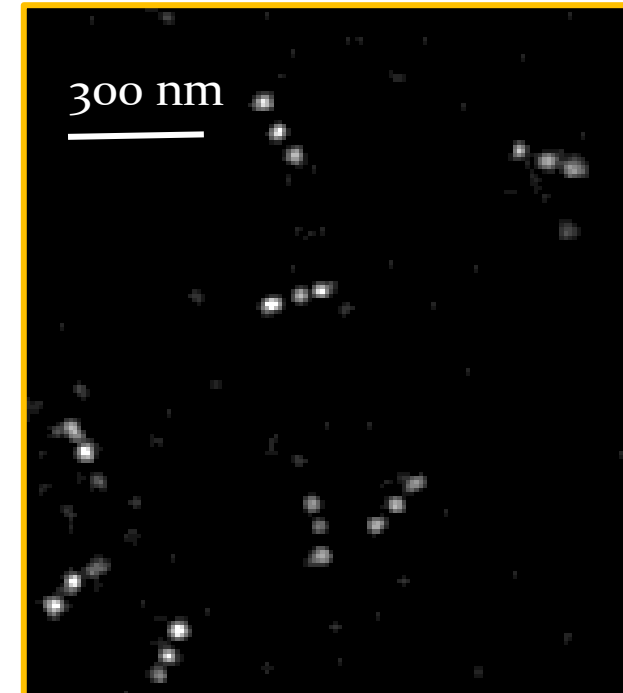
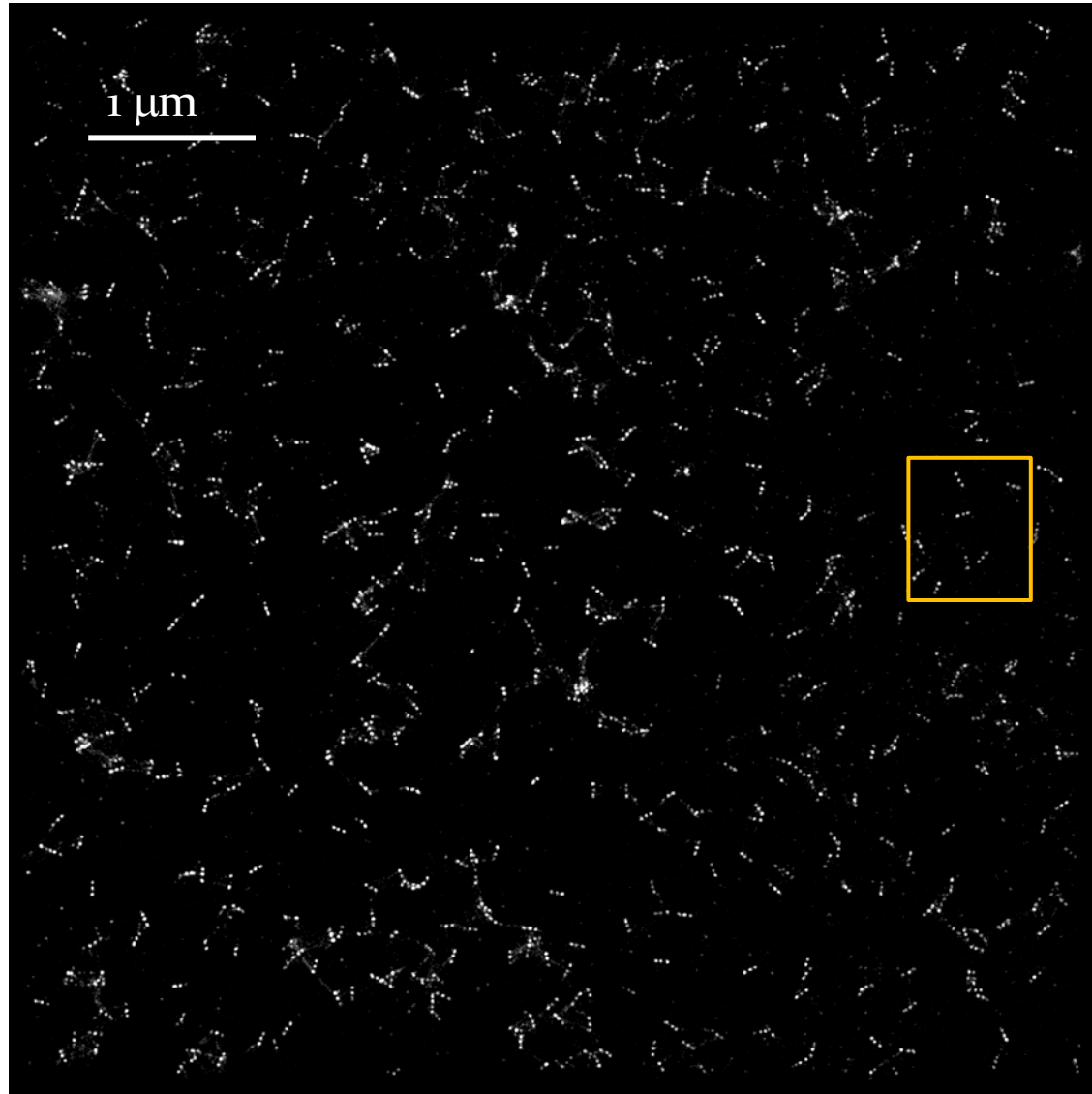
PAINT: Points Accumulation for Imaging in Nanoscale Topography

A. Sharonov & R. M. Hochstrasser *PNAS* 103 (2006) 18911-18916

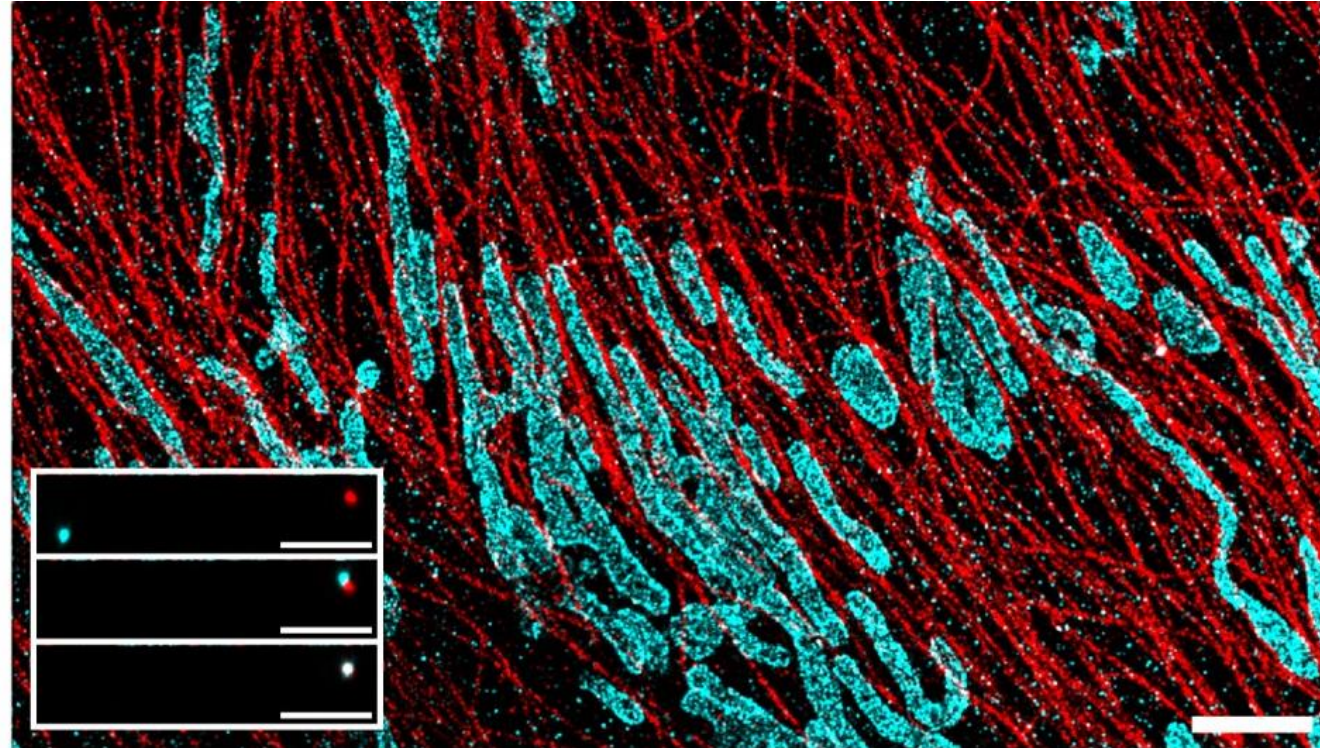
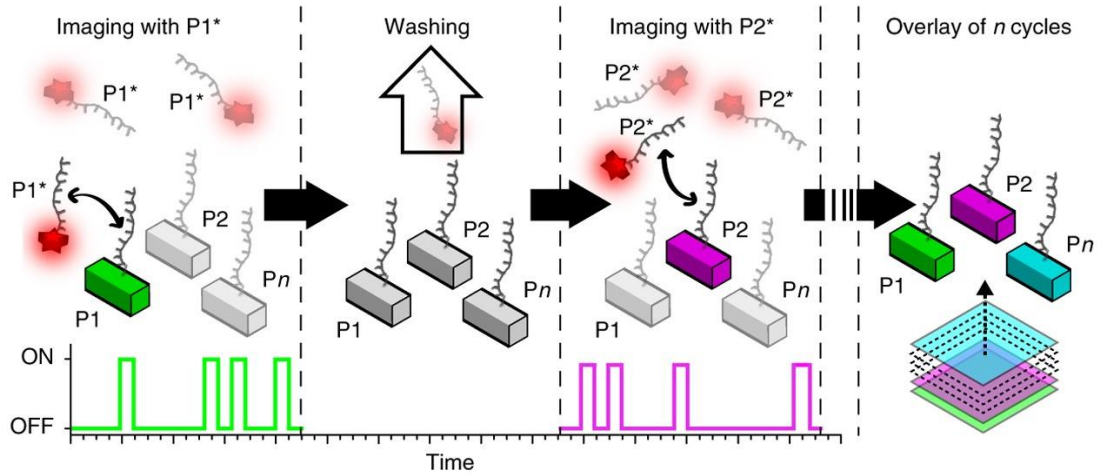
Various SMLM : DNA-PAINT



Various SMLM : DNA-PAINT

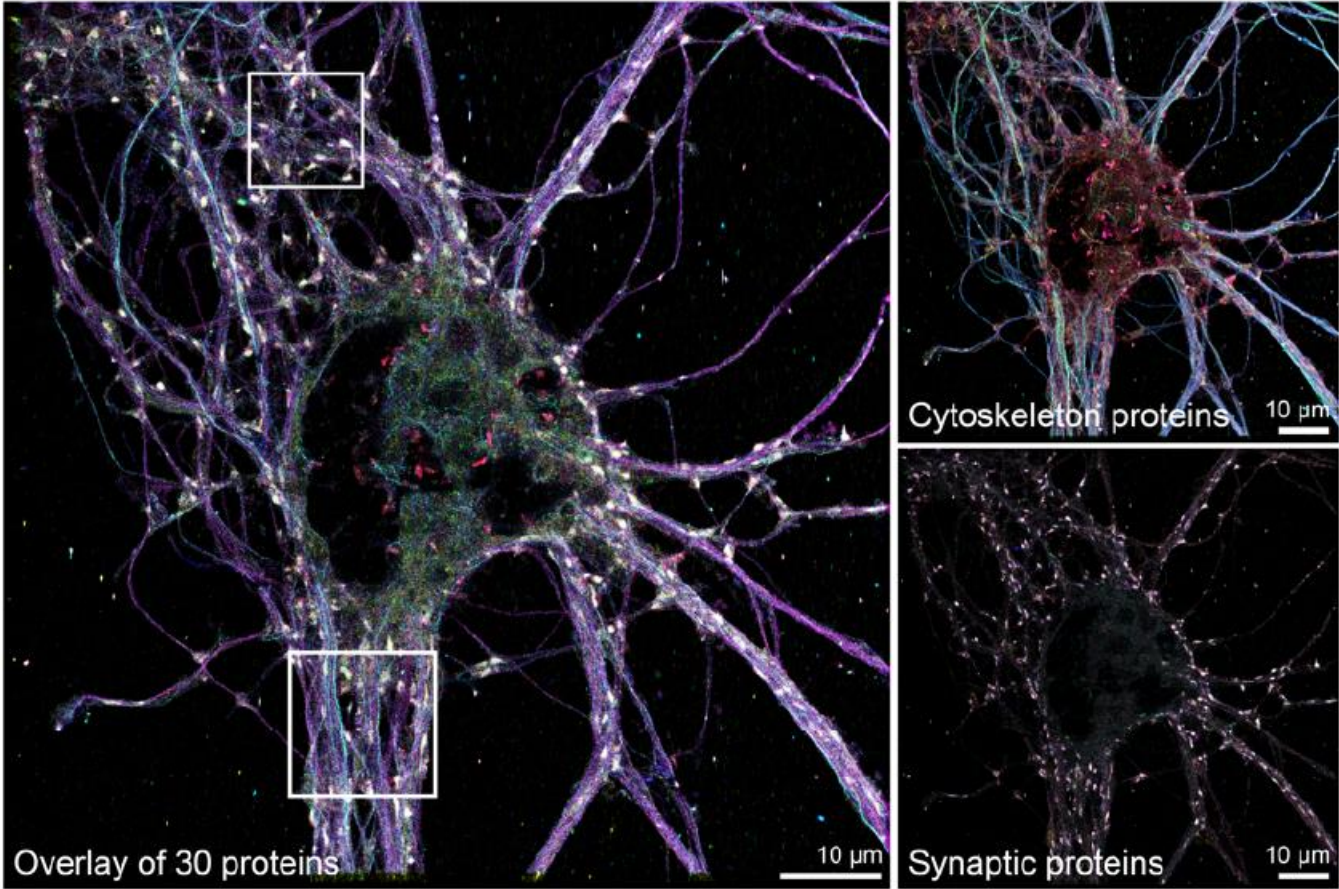


Multicolor DNA-PAINT by sequential multiplexing

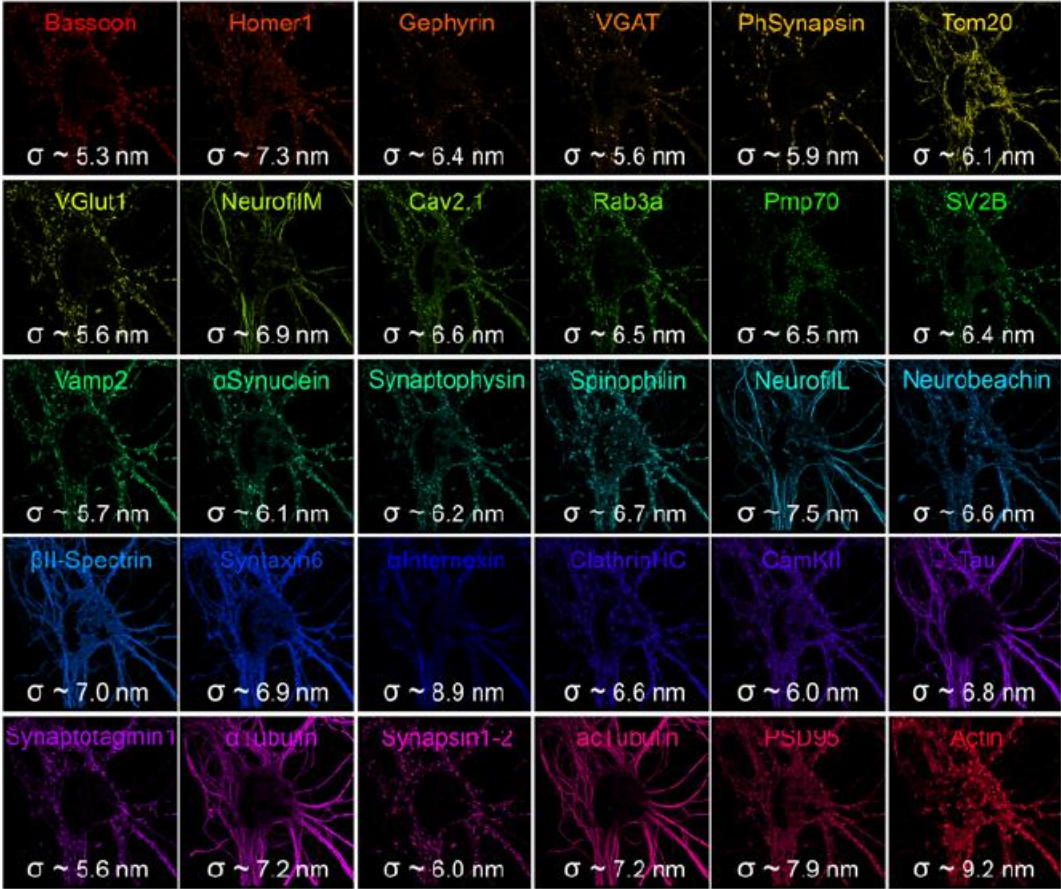


Multicolor DNA-PAINT by sequential multiplexing

30-plex neuron atlas at single-protein resolution



Individual protein targets



Coordinate-targeted super-resolution microscopy

Coordinate-targeted super-resolution microscopy

FLUORESCENT LABELS



Coordinate-targeted super-resolution microscopy

FLUORESCENT LABELS



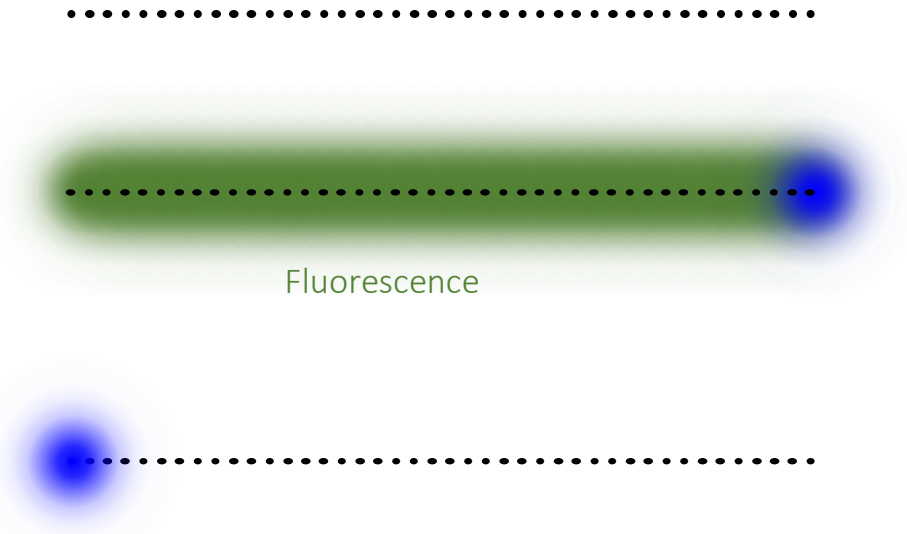
ON
(excitation)



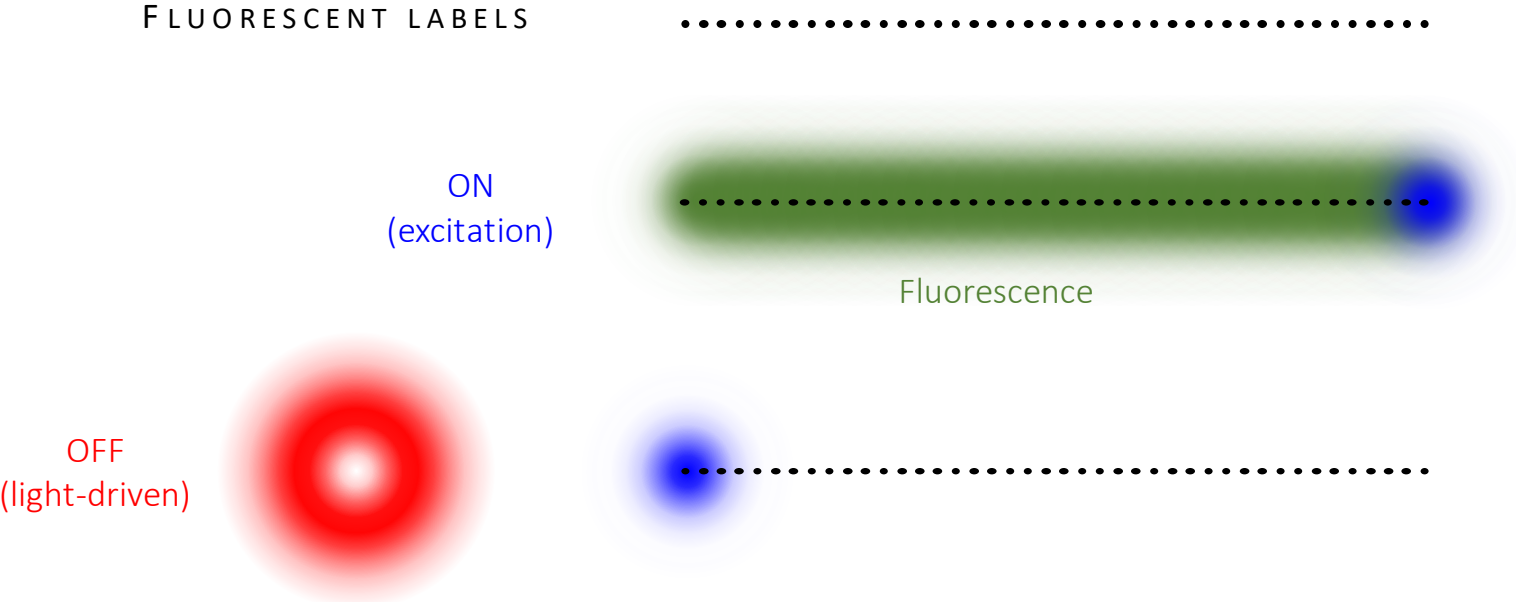
Coordinate-targeted super-resolution microscopy

FLUORESCENT LABELS

ON
(excitation)



Coordinate-targeted super-resolution microscopy



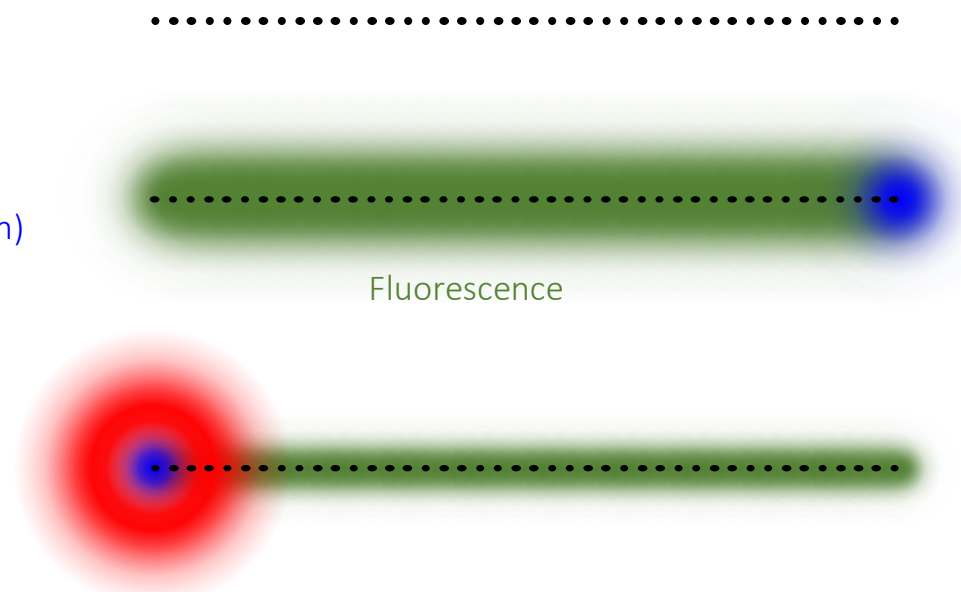
Coordinate-targeted super-resolution microscopy

FLUORESCENT LABELS

ON
(excitation)

Fluorescence

OFF
(light-driven)



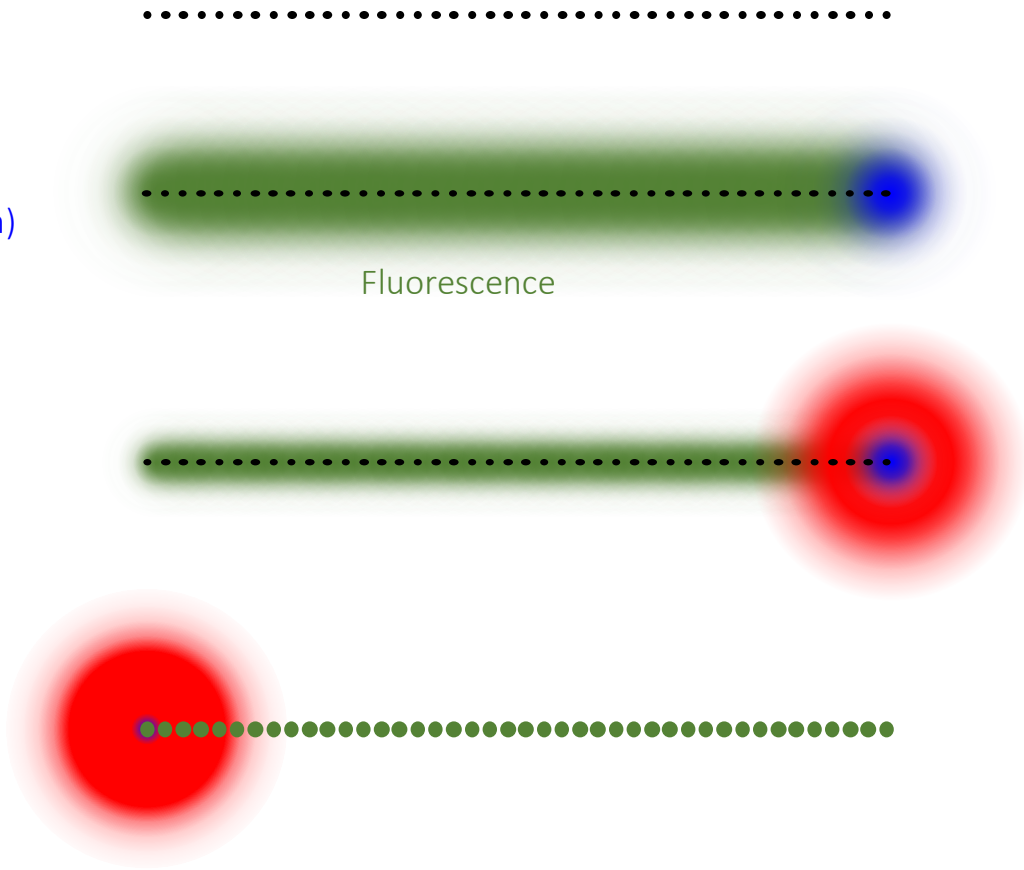
Coordinate-targeted super-resolution microscopy

FLUORESCENT LABELS

ON
(excitation)

Fluorescence

OFF
(light-driven)



Coordinate-targeted super-resolution microscopy

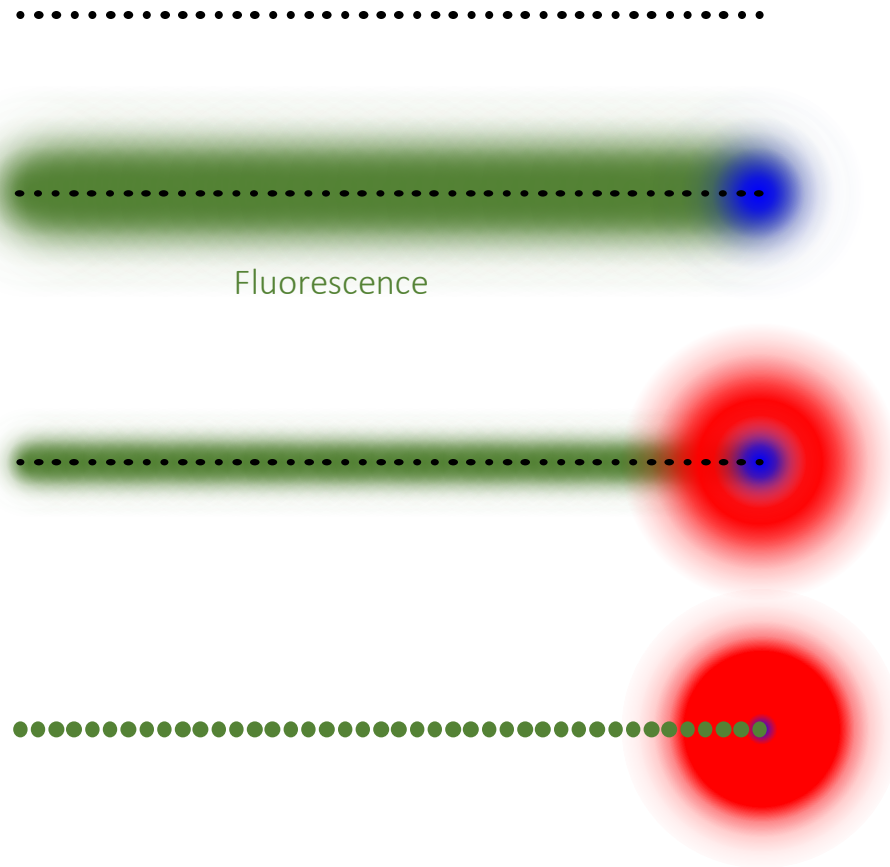
FLUORESCENT LABELS

ON
(excitation)

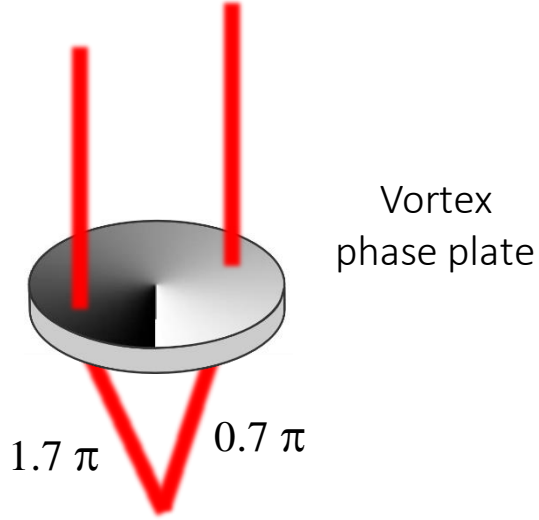
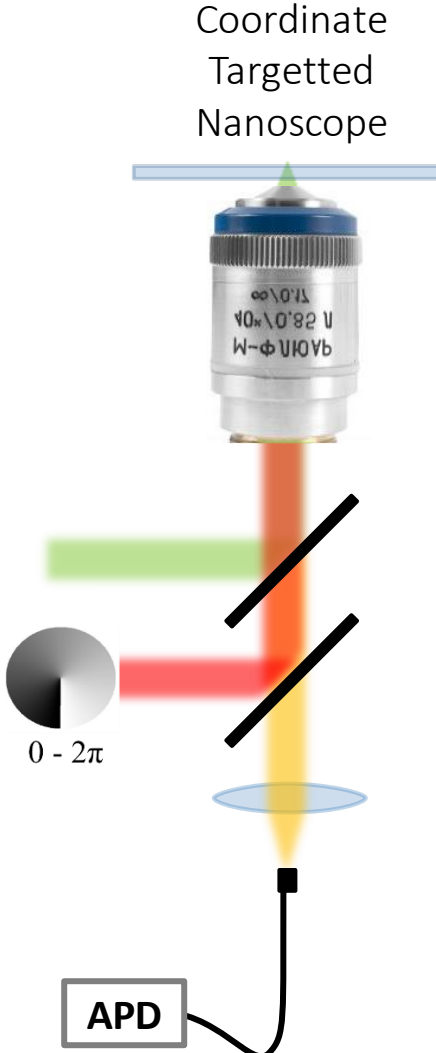
Fluorescence

OFF
(light-driven)

- STIMULATED EMISSION (STED)
- GROUND-STATE DEPLETION (GSDIM)
- PHOTOCROMISM
- ...

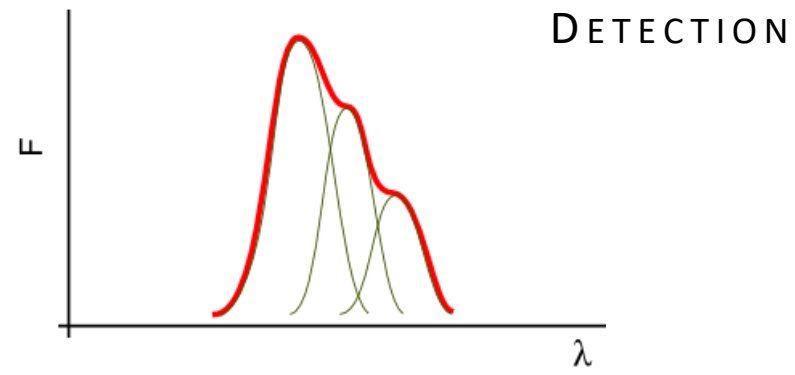
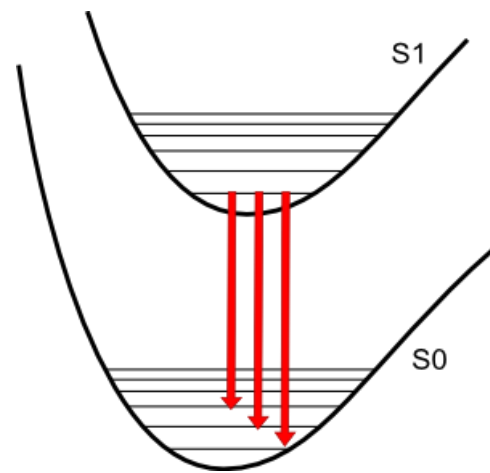
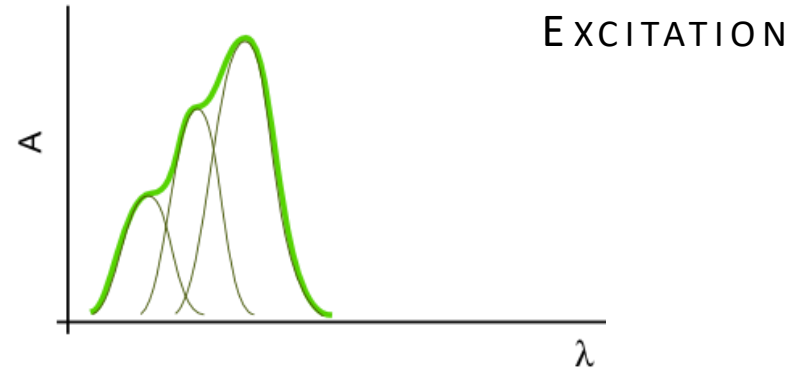
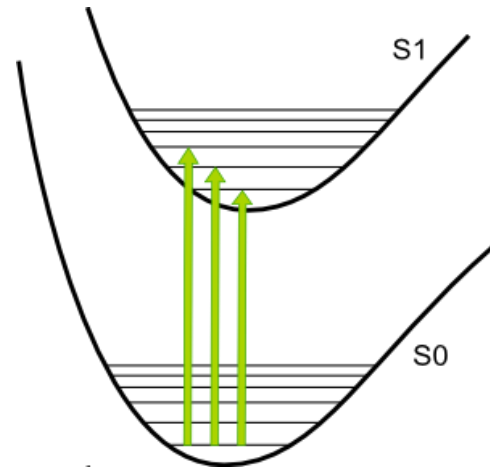


Coordinate-targeted super-resolution microscopy

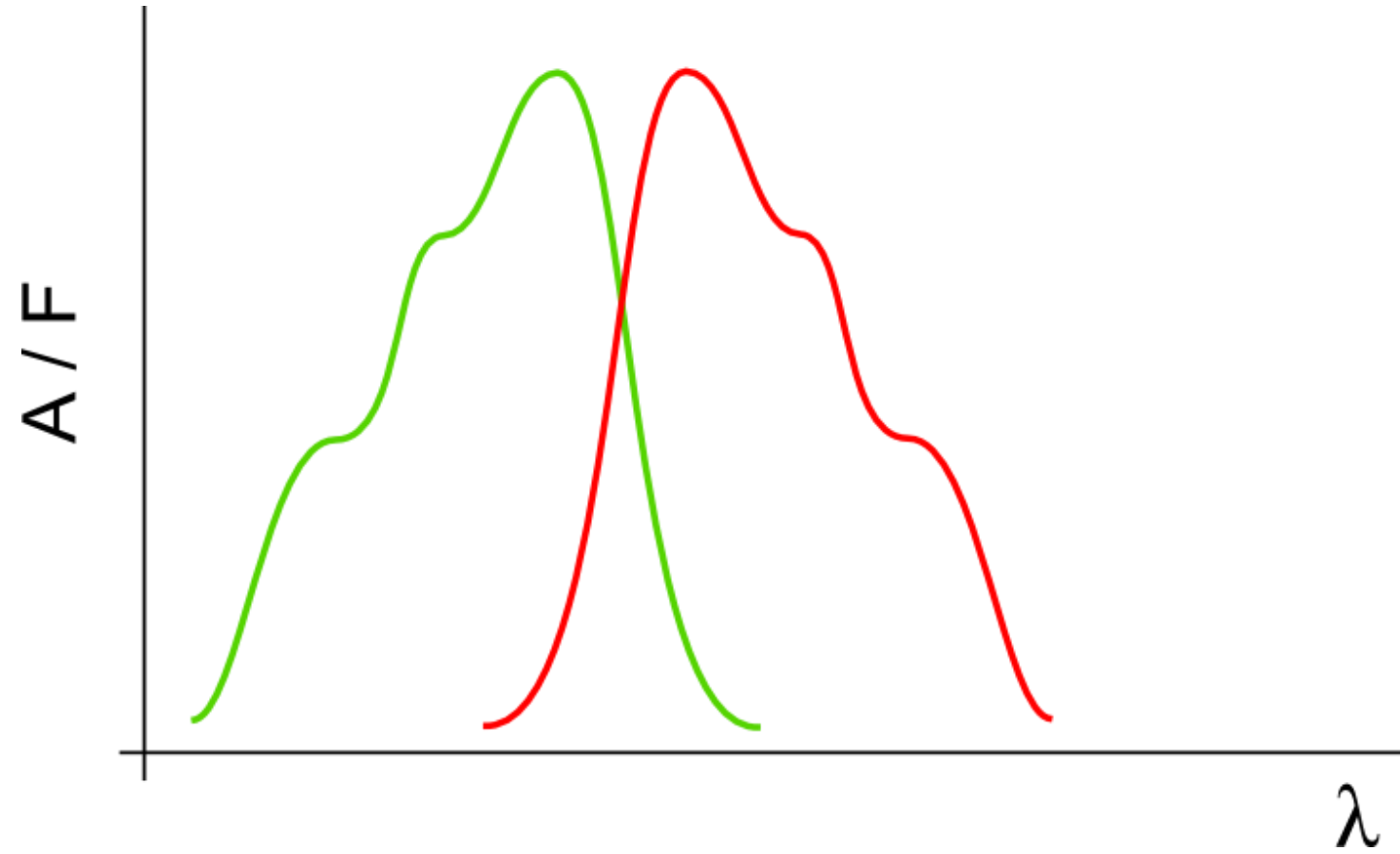


scanning, pulse sync, high-power, positional and focus stability, ...

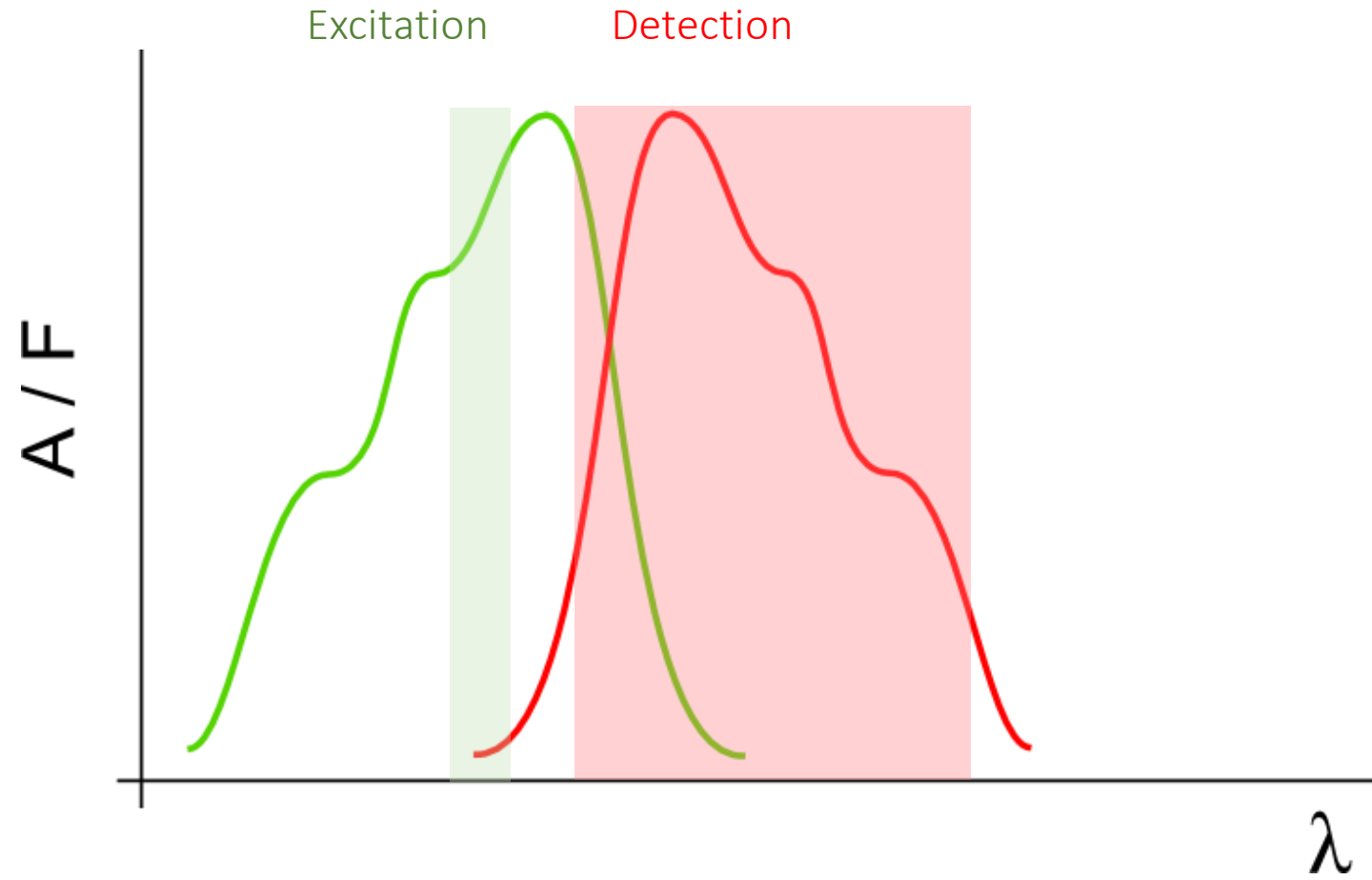
STED nanoscopy



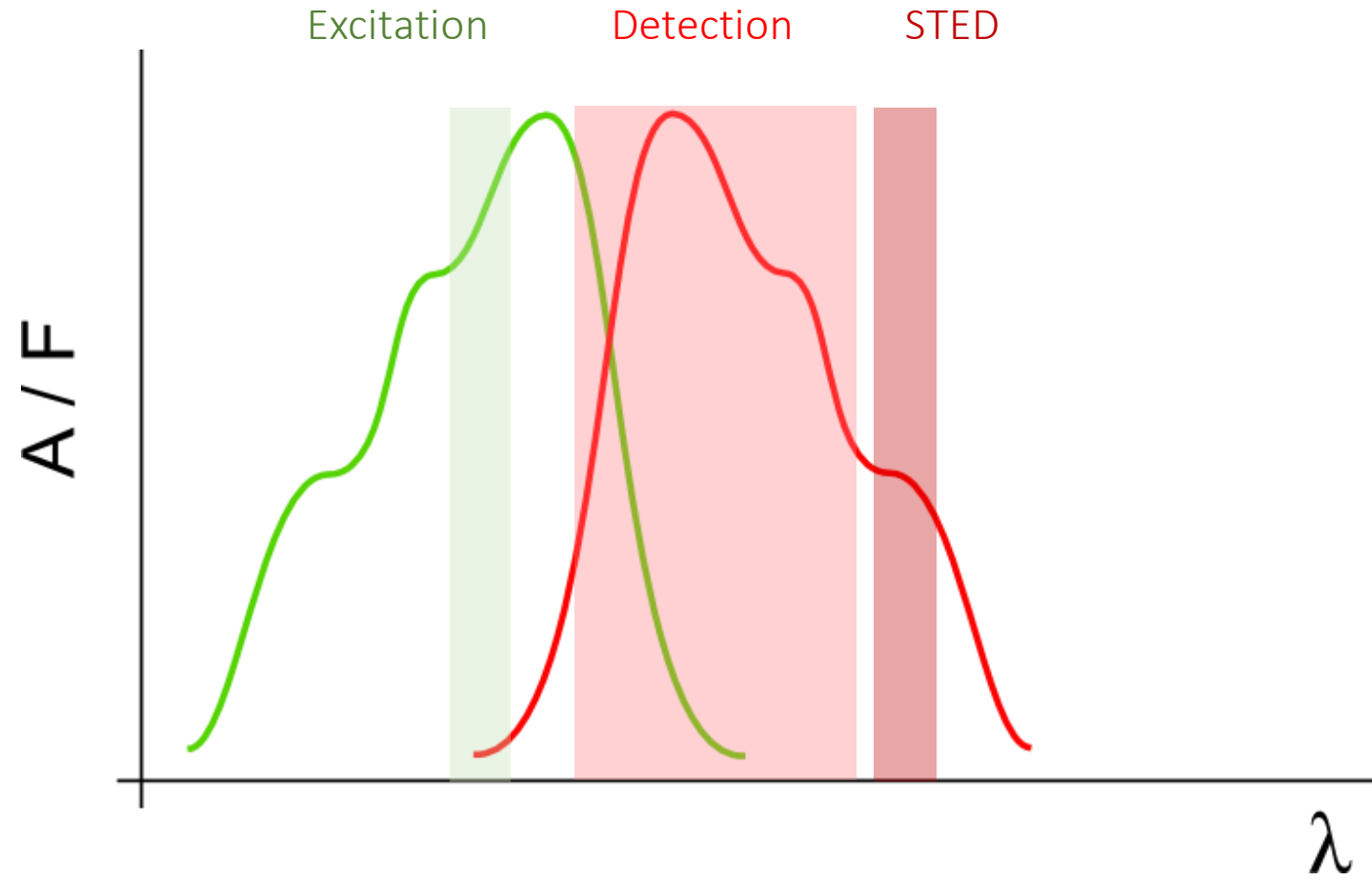
STED nanoscopy



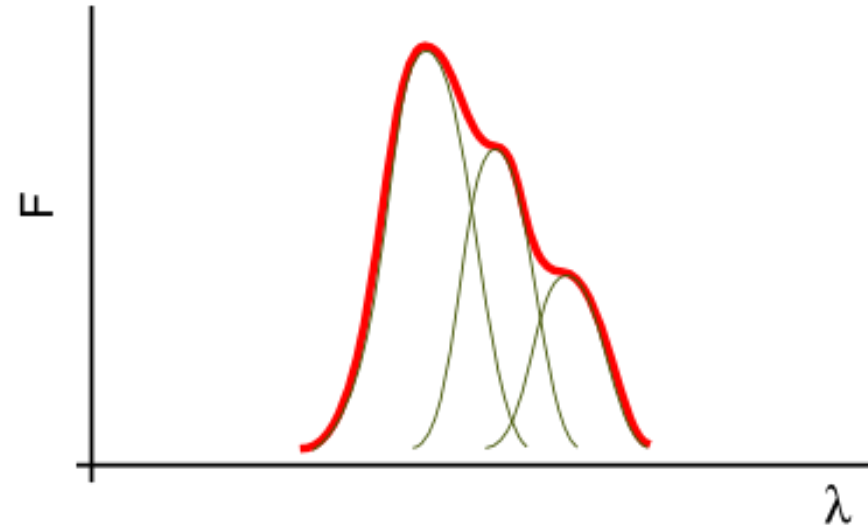
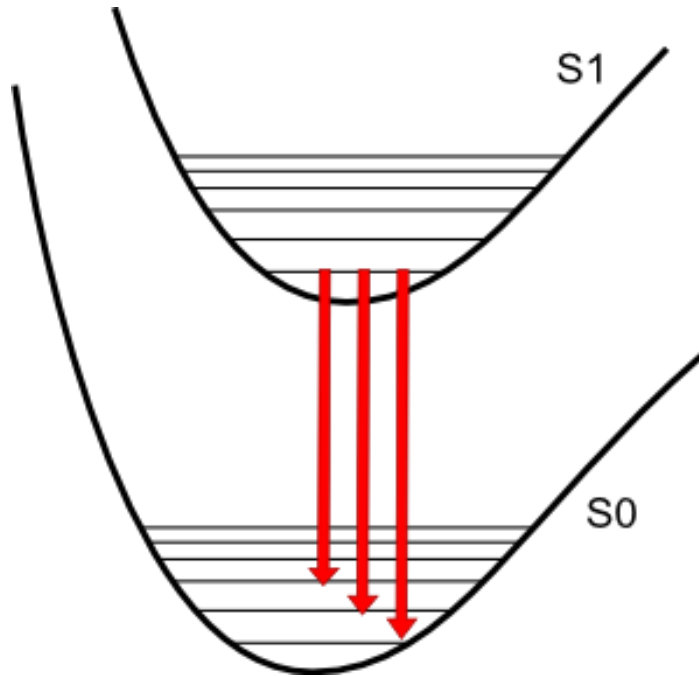
STED nanoscopy



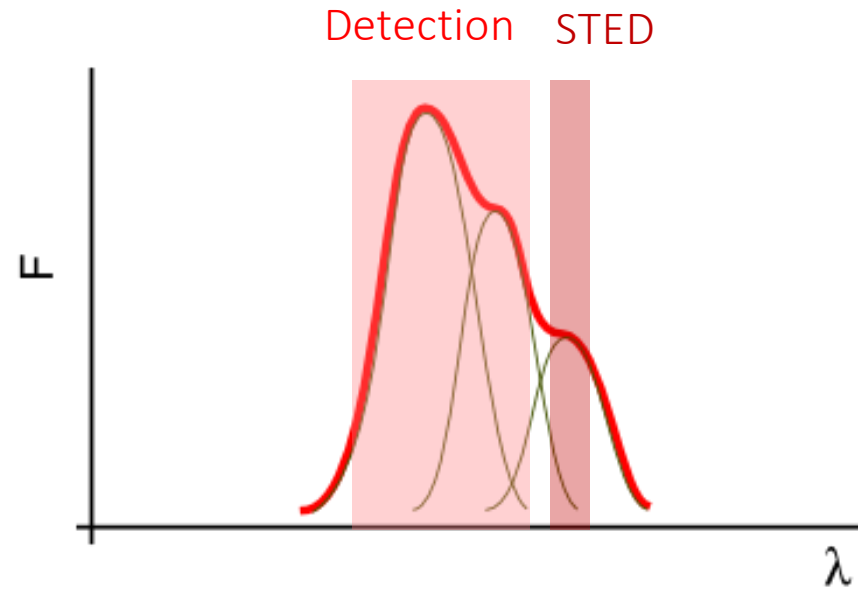
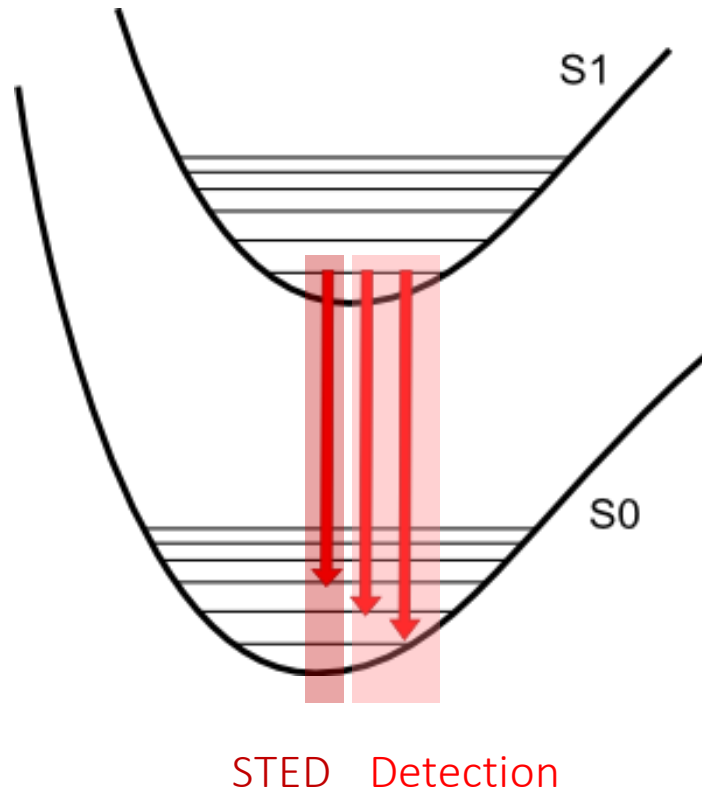
STED nanoscopy



STED nanoscopy

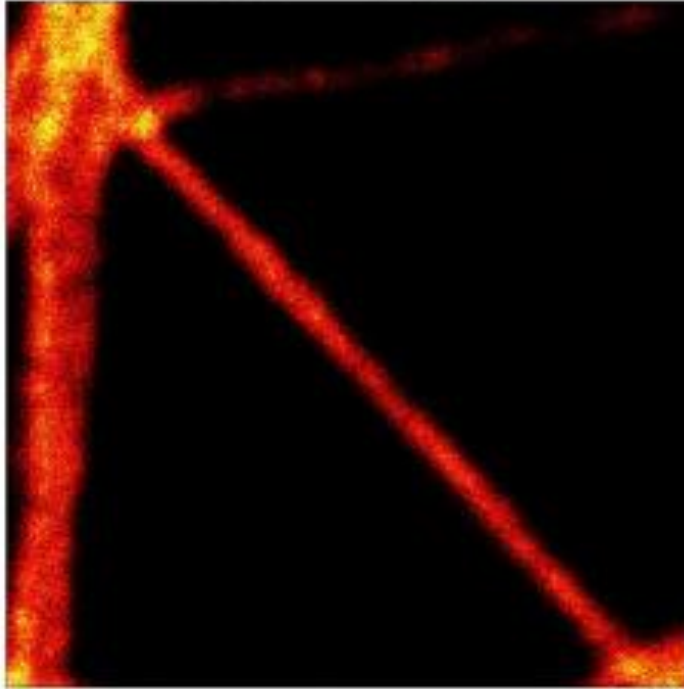


STED nanoscopy

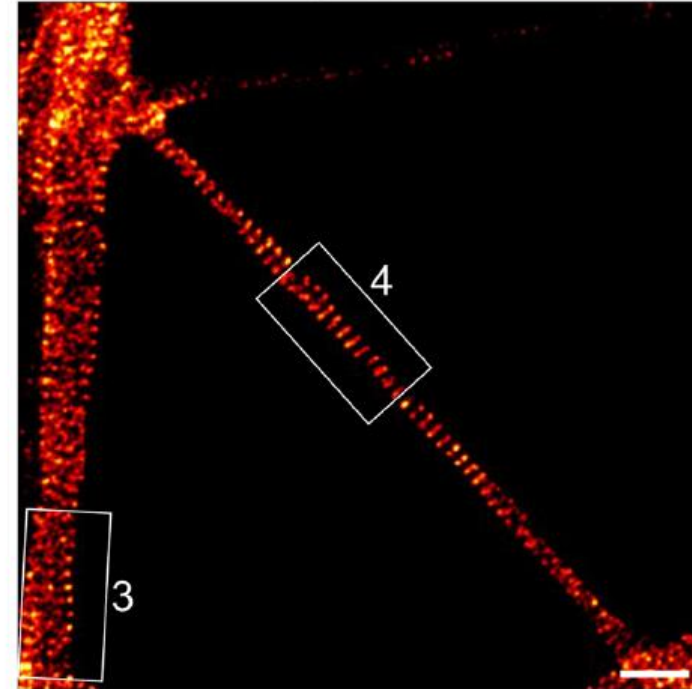


STED nanoscopy

CONFOCAL

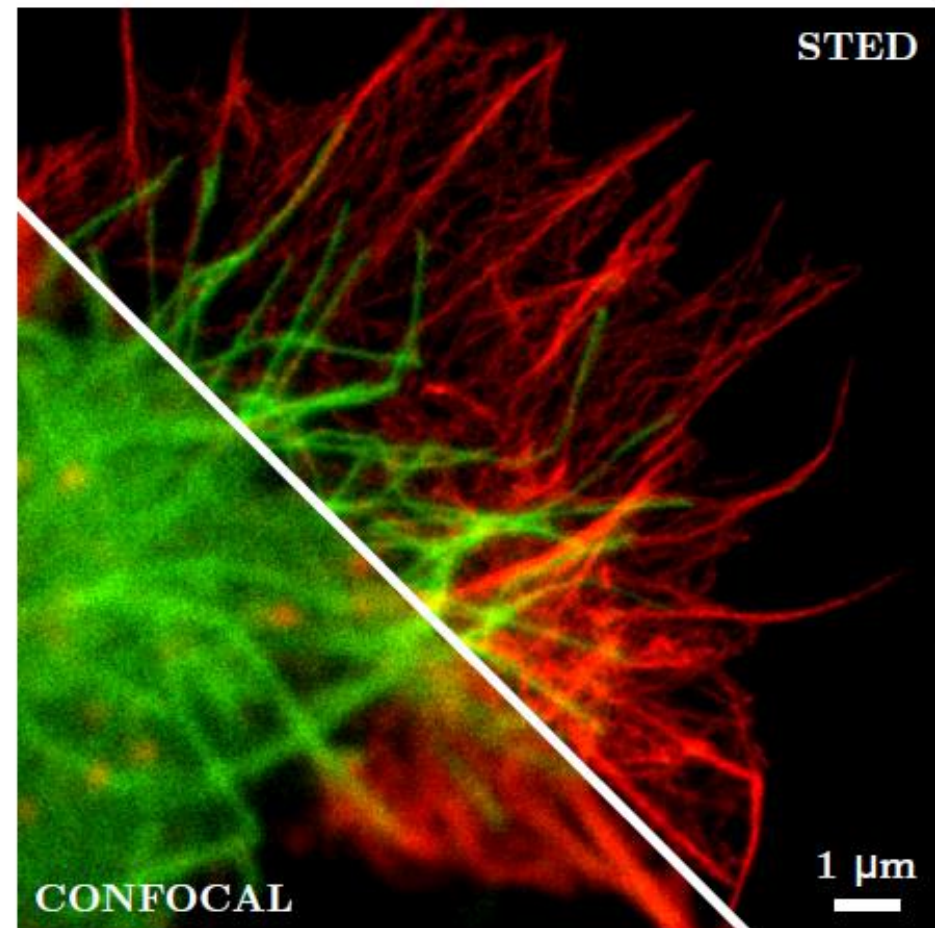
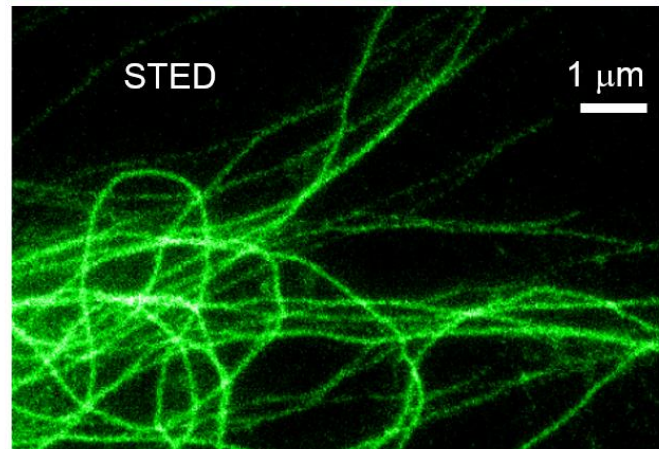
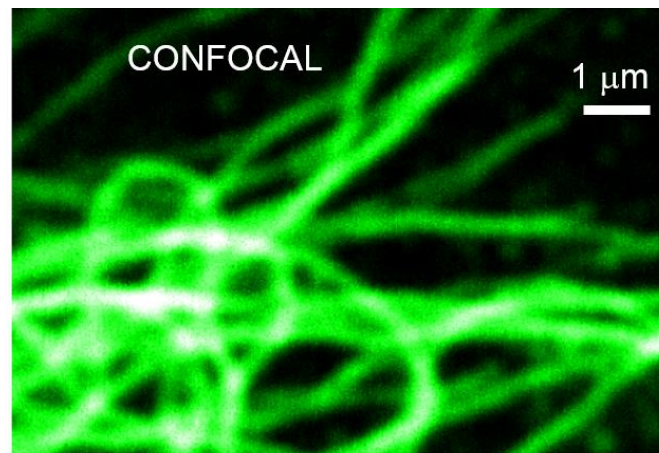
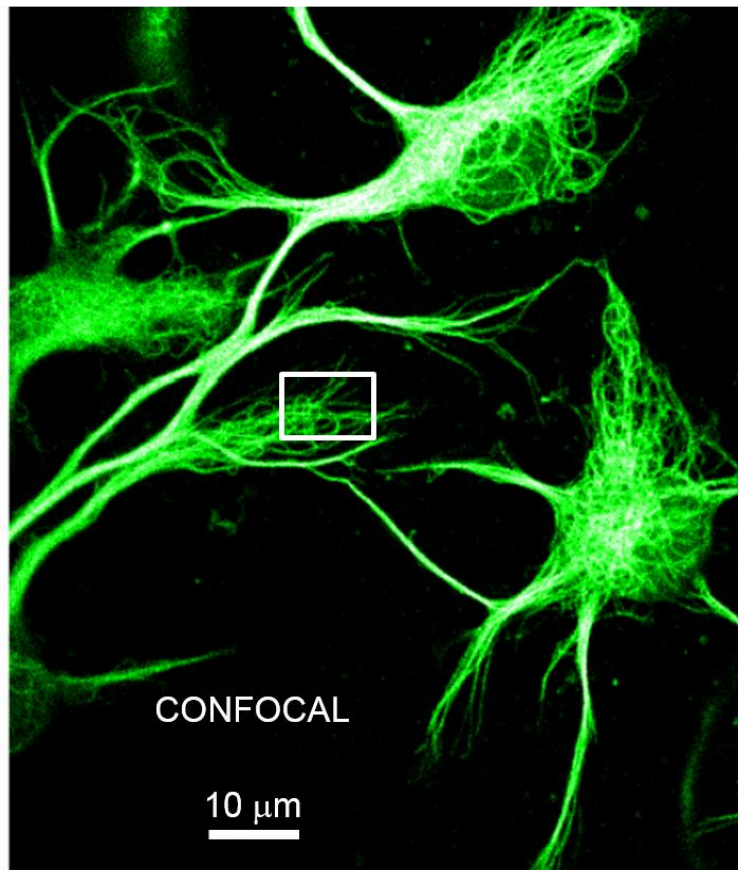


STED



Membrane associated periodic skeleton (MPS) of neurons

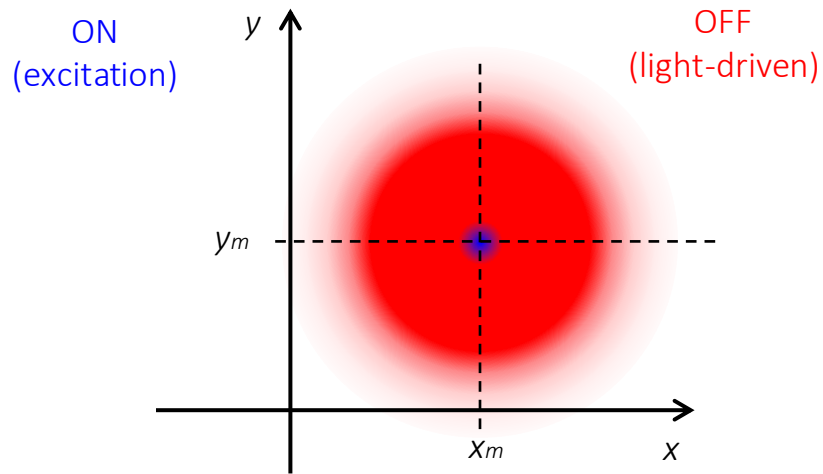
STED nanoscopy



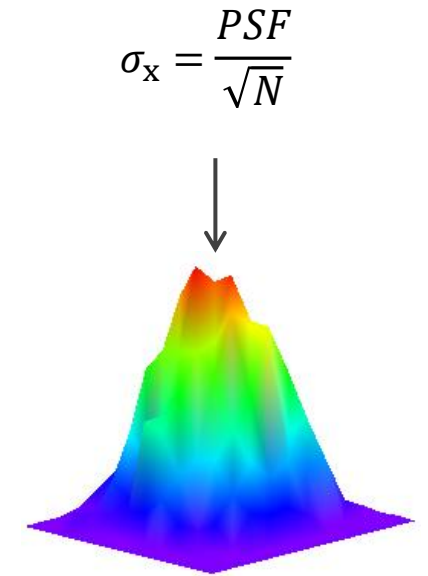
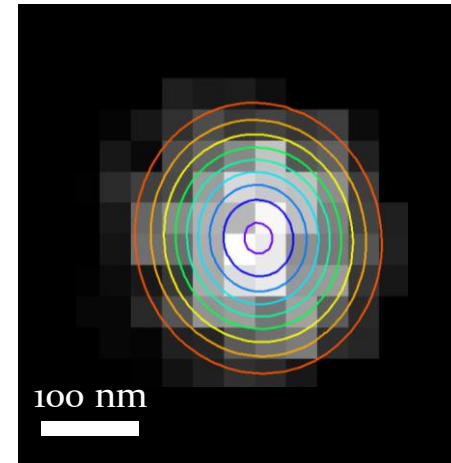
Fluorescence Nanoscopy 2nd Generation

Fluorescence nanoscopy resolution limits

Position information is injected by the light pattern



Position information obtained from the emission (image)

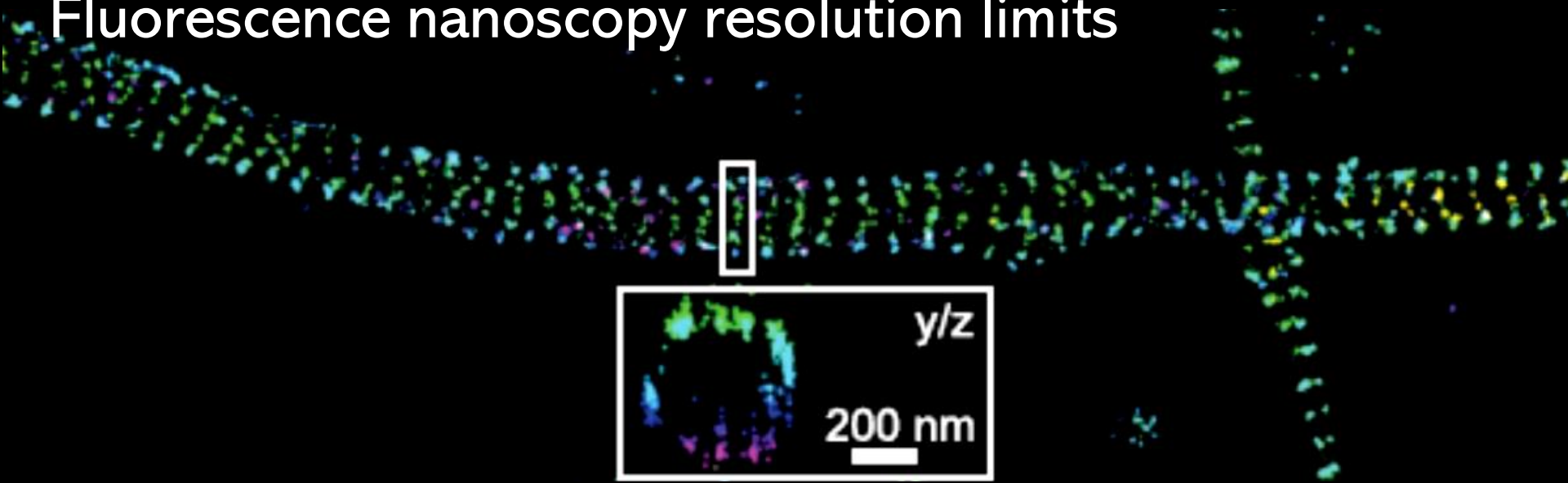


Ideally (no background, perfect off-switching) one photon locates the emitter

In practice, high spatial resolution requires N

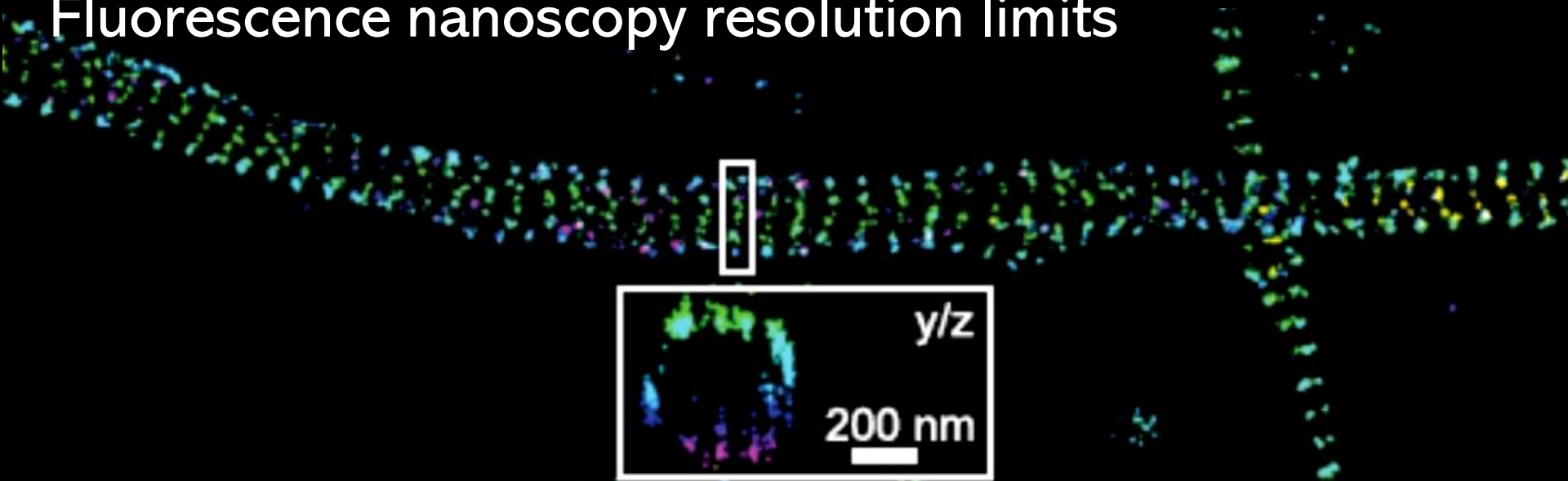
High localization precision require shigh N

Fluorescence nanoscopy resolution limits

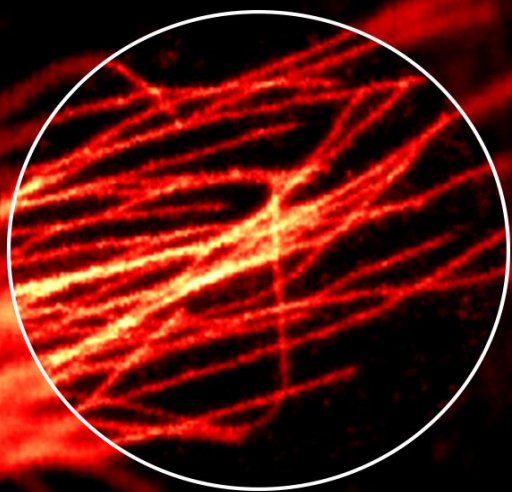


$$\sigma_x = \sigma_y = 15 - 60 \text{ nm}$$
$$\sigma_z = 30 - 120 \text{ nm}$$

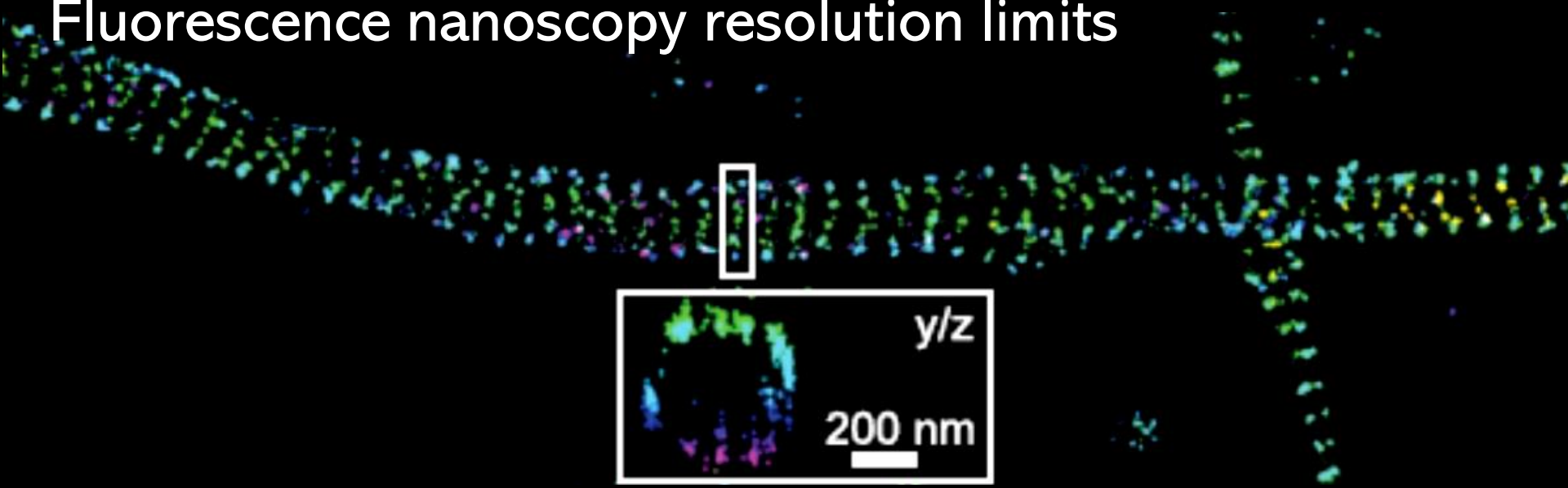
Fluorescence nanoscopy resolution limits



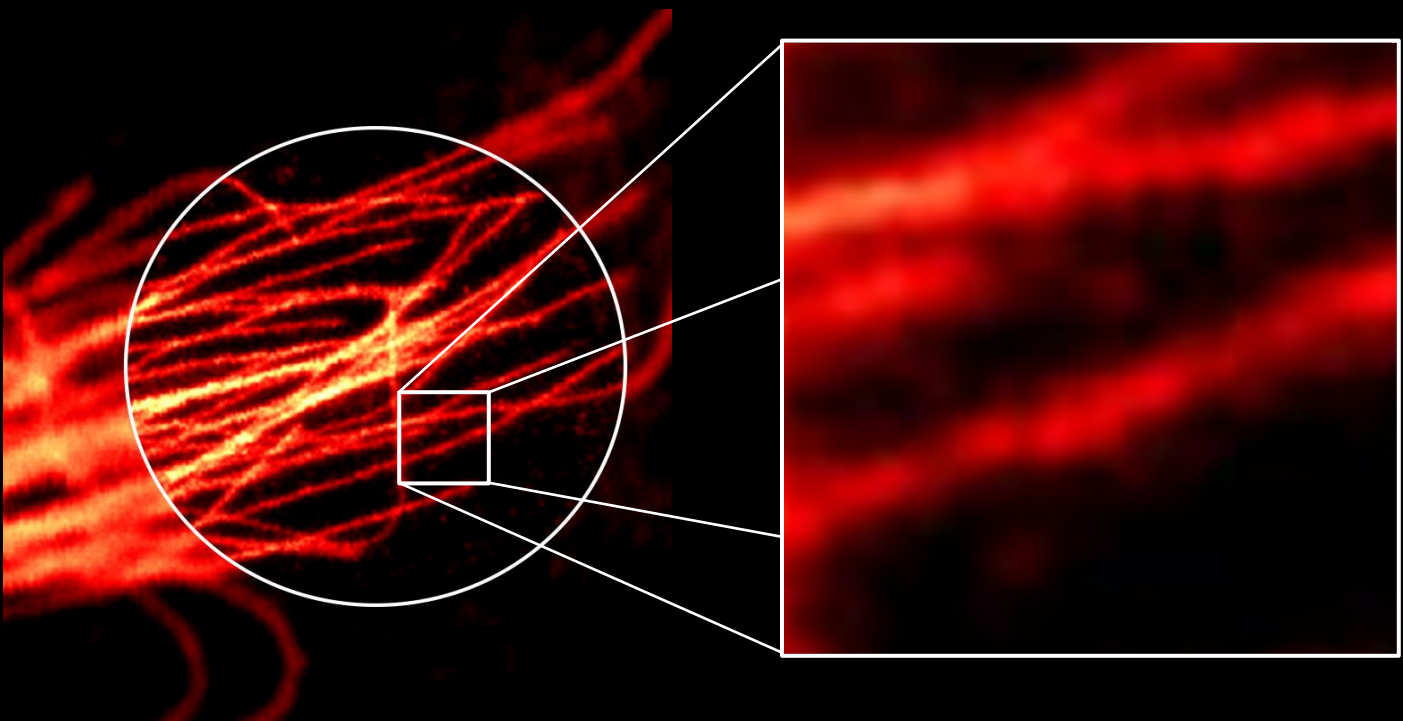
$$\sigma_x = \sigma_y = 15 - 60 \text{ nm}$$
$$\sigma_z = 30 - 120 \text{ nm}$$



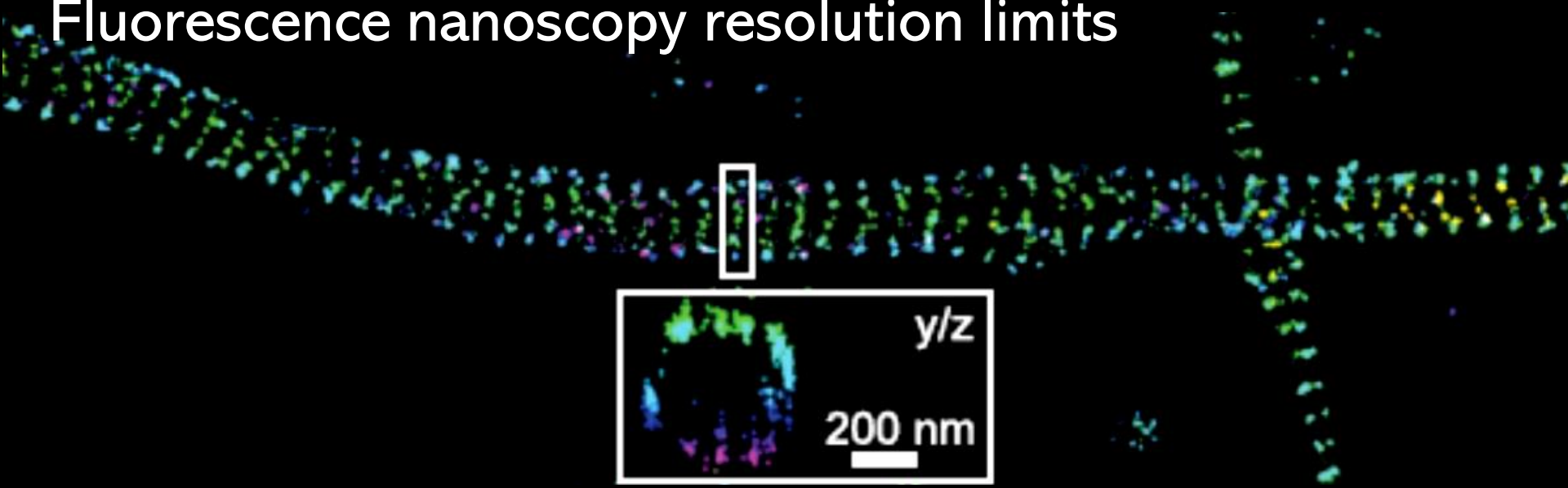
Fluorescence nanoscopy resolution limits



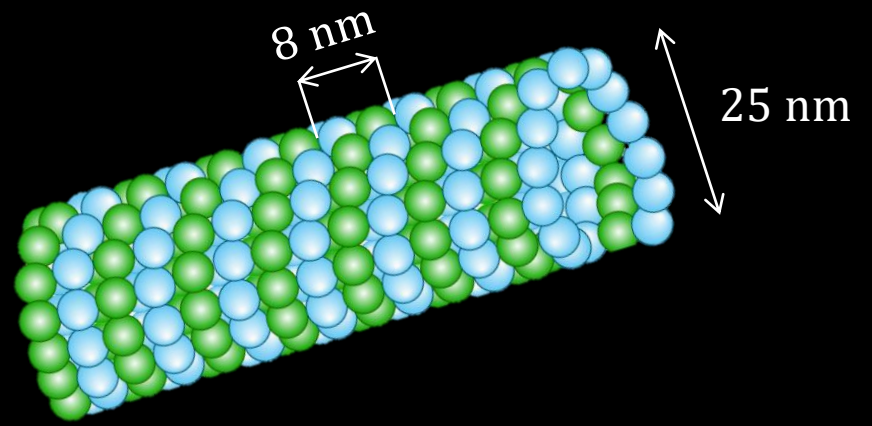
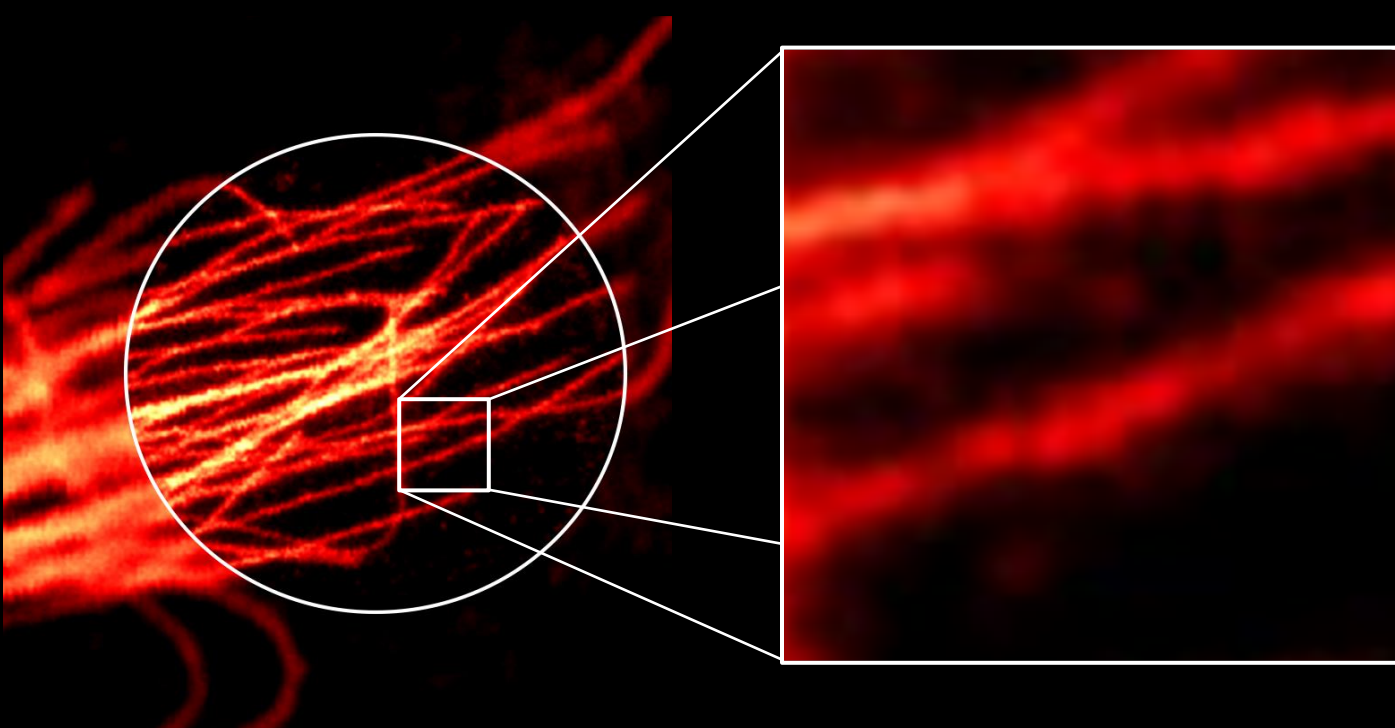
$$\sigma_x = \sigma_y = 15 - 60 \text{ nm}$$
$$\sigma_z = 30 - 120 \text{ nm}$$



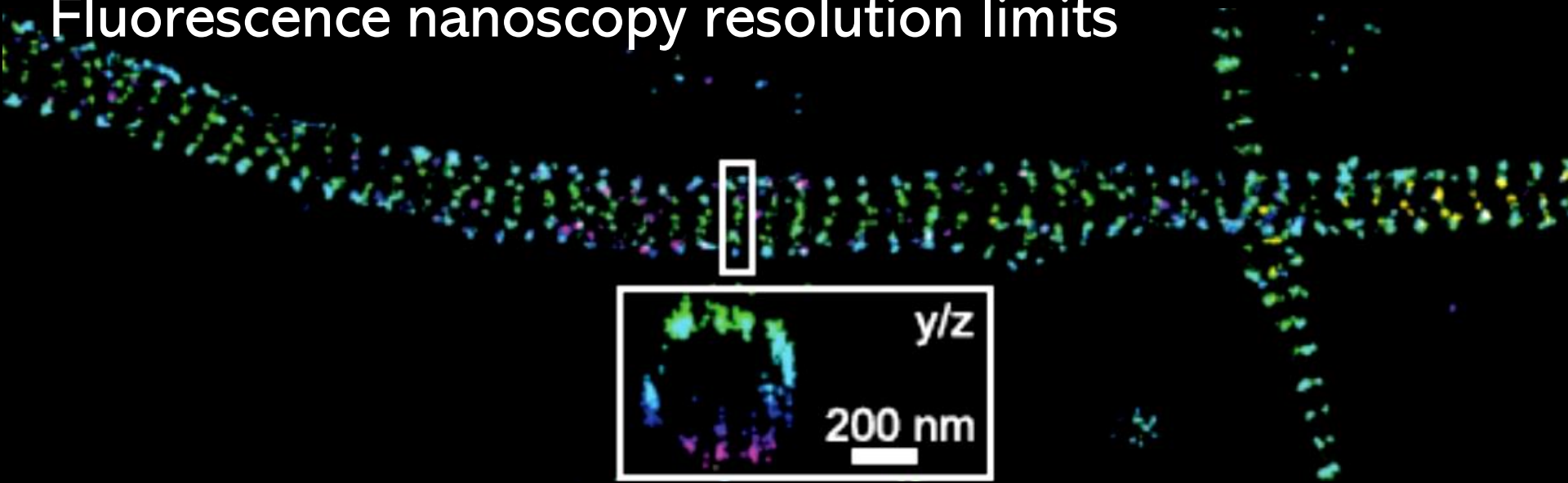
Fluorescence nanoscopy resolution limits



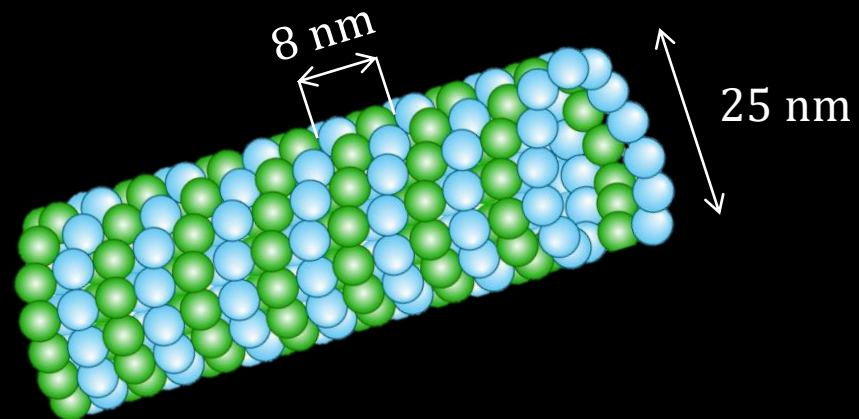
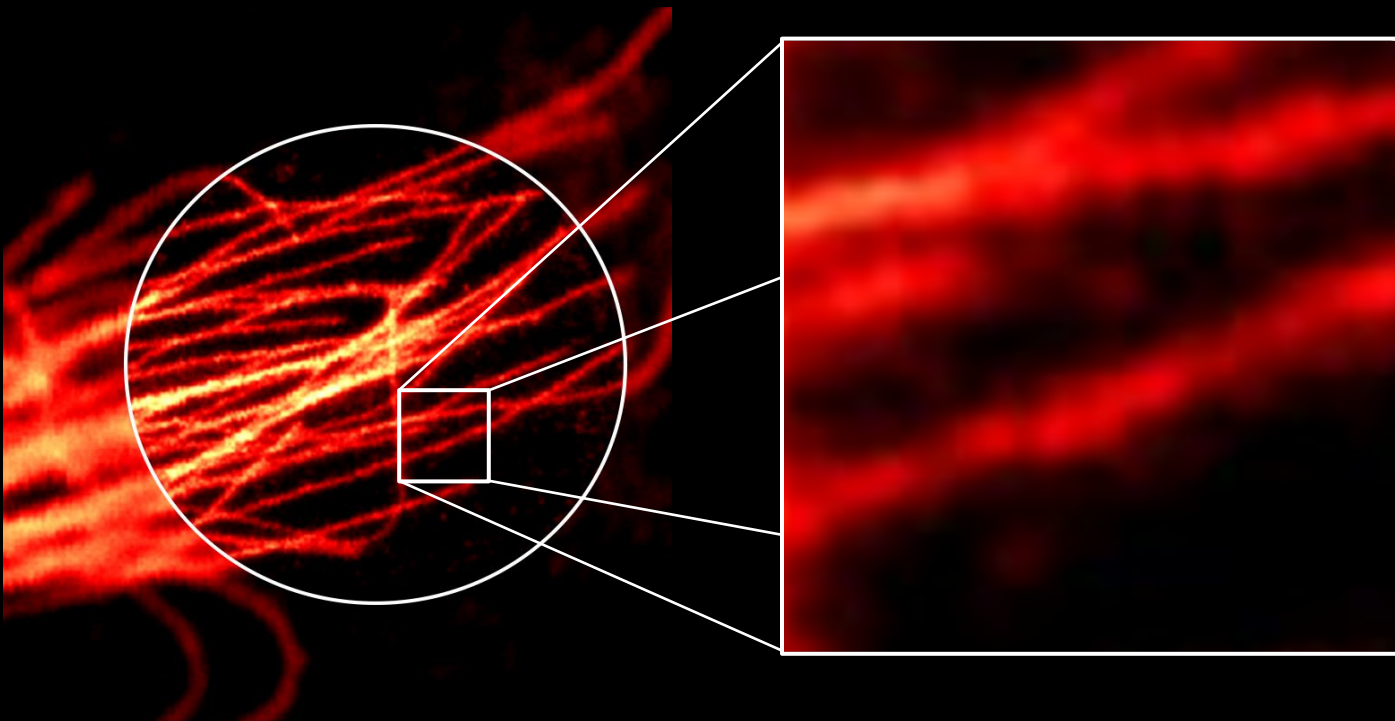
$$\sigma_x = \sigma_y = 15 - 60 \text{ nm}$$
$$\sigma_z = 30 - 120 \text{ nm}$$



Fluorescence nanoscopy resolution limits

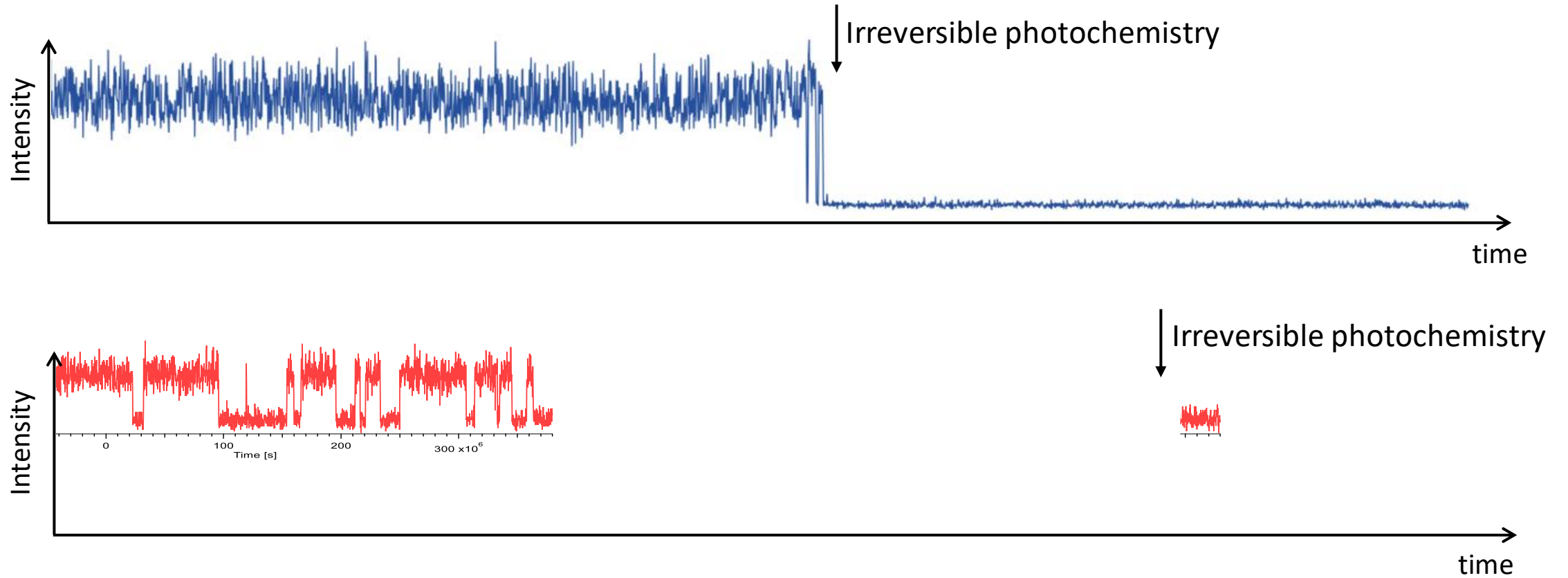


$$\sigma_x = \sigma_y = 15 - 60 \text{ nm}$$
$$\sigma_z = 30 - 120 \text{ nm}$$



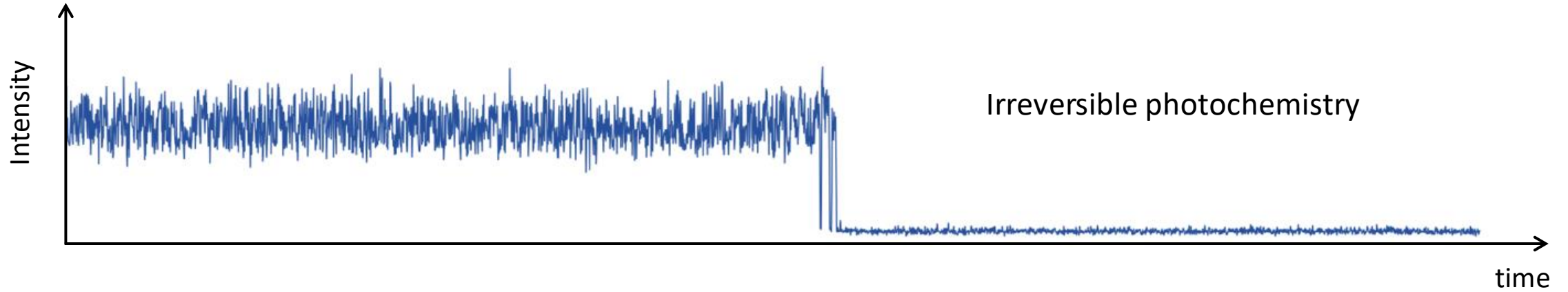
We need an extra push into the sub-10 nm regime

Fluorescence nanoscopy resolution limits: photobleaching



- Imaging resolution limited to a few tens of nm
- Single-molecule tracking limited time/length

Nanoscopy resolution limit: photon budget



Solutions

Get more photons:

DNA-PAINT

Stabilizing buffers

Self-healing dyes

Get more information:

SML-SSI (MINFLUX, RASTMIN)

SIMPLER

STED-FRET

Nanoscopy with sub-10 nm resolution

SML-SSI Single-Molecule Localization with Sequential Structured Illumination

Balzarotti et al. *Science* 355 (2017) 606-612

Masullo et al. *Nano Letters* 21 (2021) 840-846

Masullo et al. *Biophysical Reports* 2 (2022) 100036

Masullo et al. *Light: Science & Applications* 11 (2022) 70

Masullo et al. *Light: Science & Applications* 11 (2022) 199

Zdańkowski et al. *ACS Photonics* 9 (2022) 3777–3785

SIMPLER Supercritical Illumination Microscopy Photometric z-Localization w/ Enhanced Resolution

Szalai et al. *Nature Communications* 12 (2021) 517

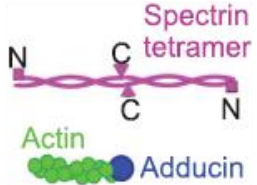
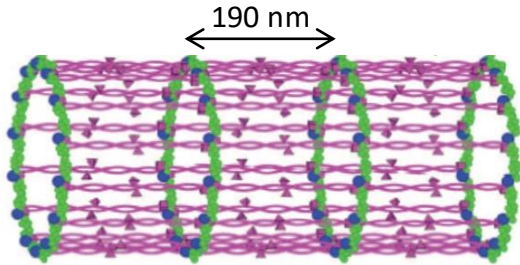
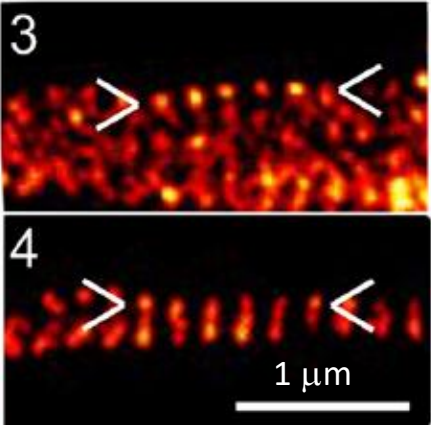
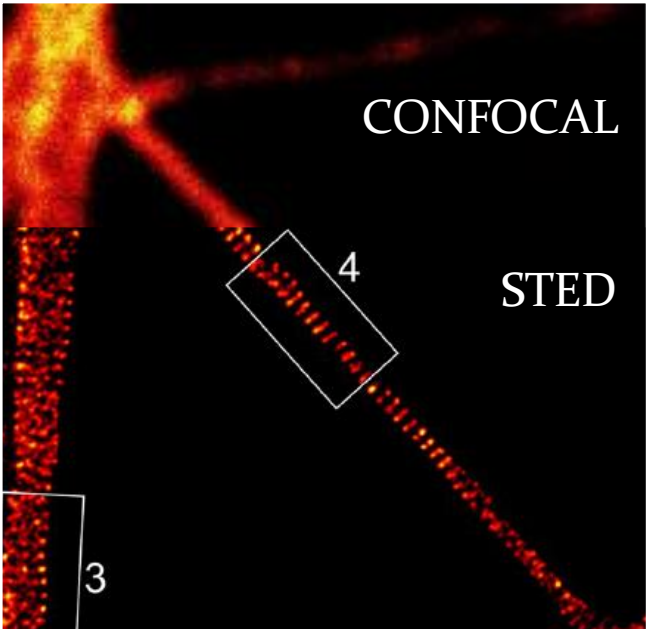
STED-FRET Super-resolved energy transfer imaging

Szalai et al. *Nano Letters* 21 (2021) 2296–2303

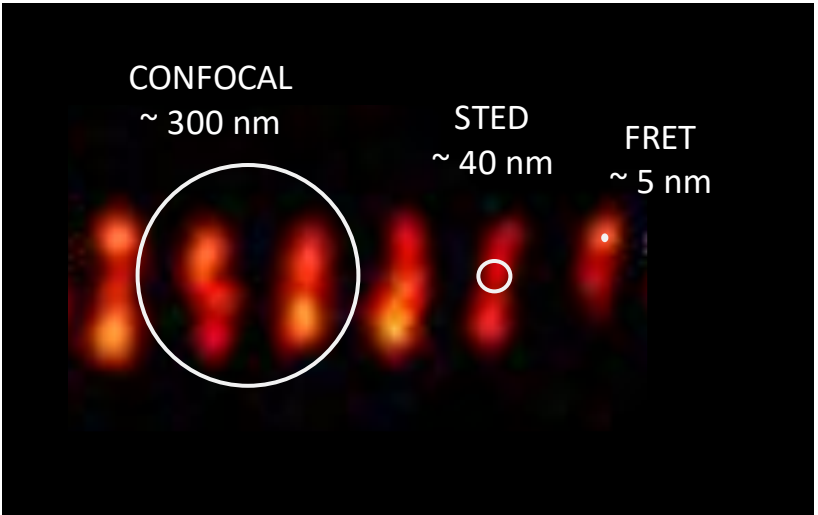
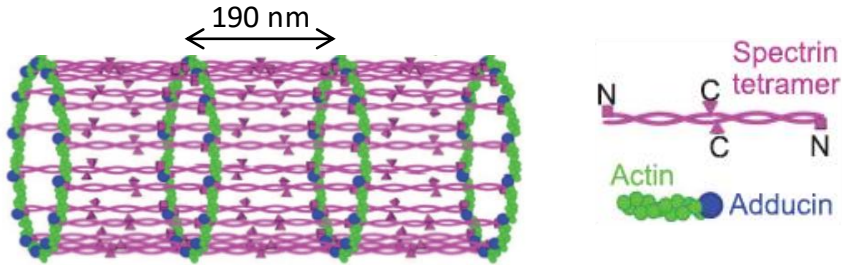
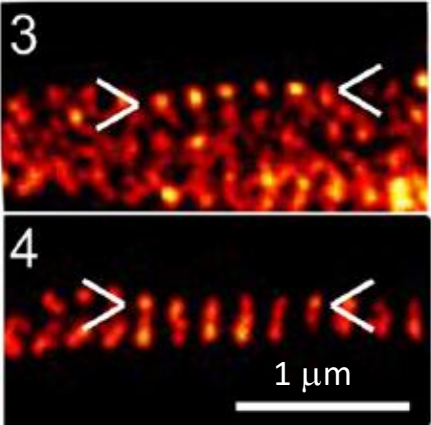
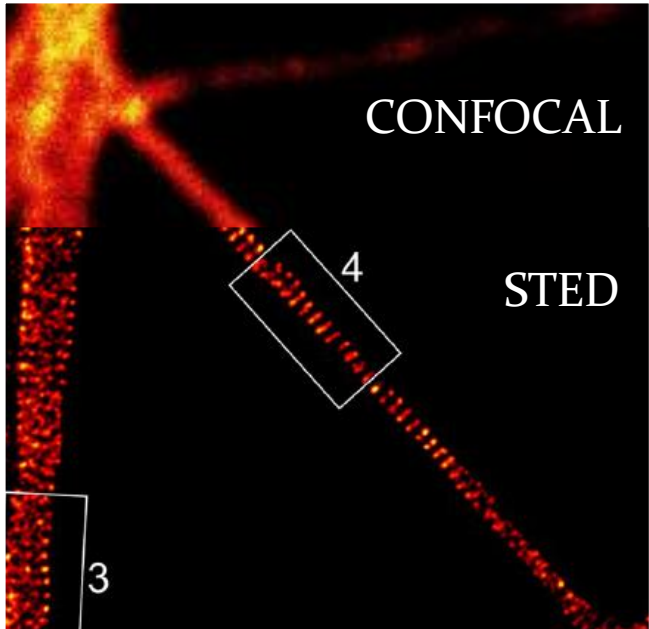
Szalai et al. *Nanoscale* 13 (2021) 18421-18433

REVIEW: Masullo, et al. "Fluorescence nanoscopy at the sub-10 nm scale" *Biophysical Reviews* 13 (2022) 1101-1112

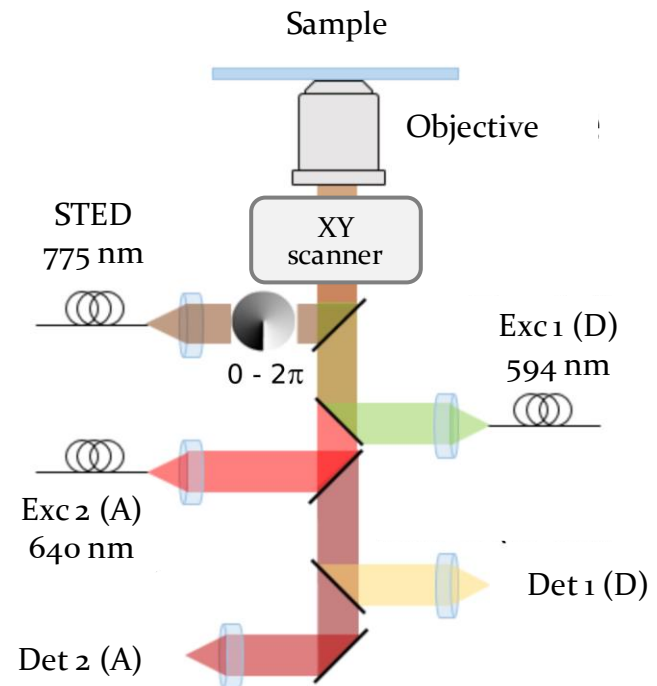
STED-FRET



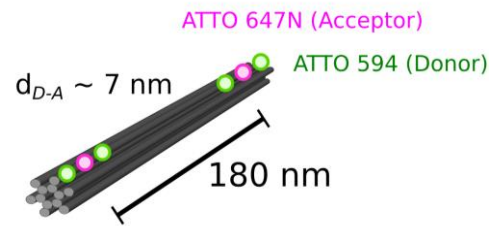
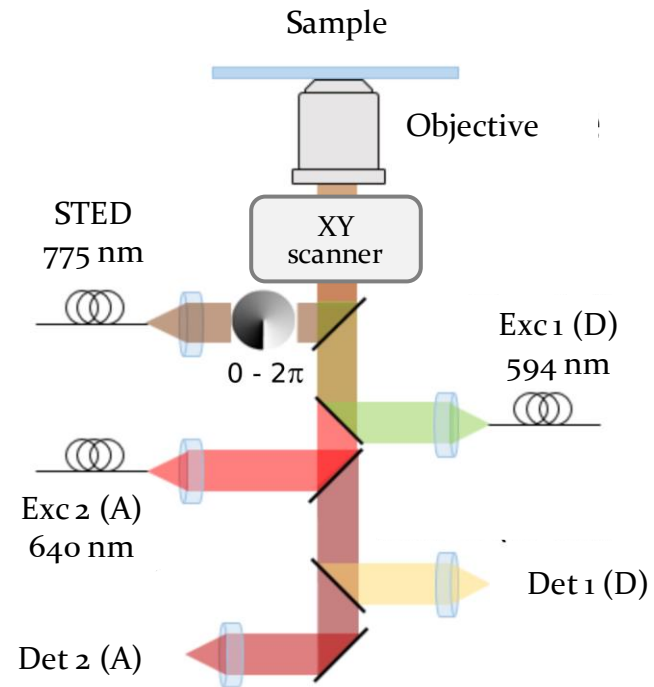
STED-FRET



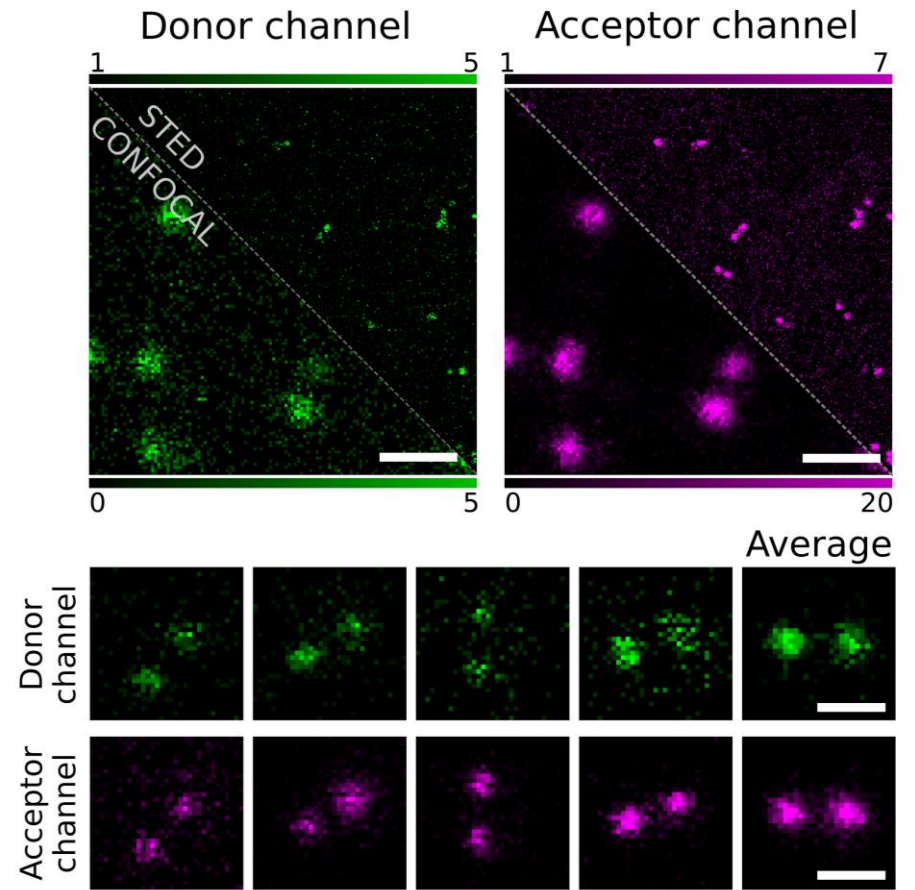
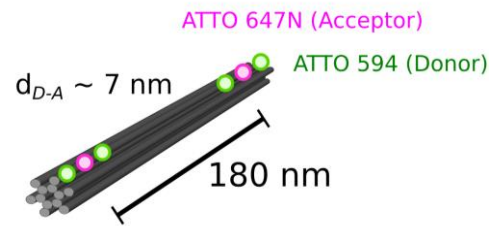
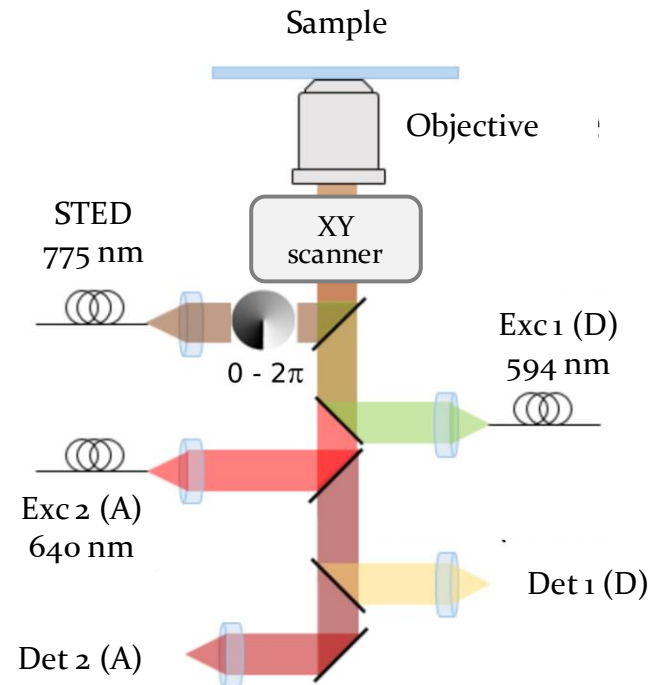
STED-FRET



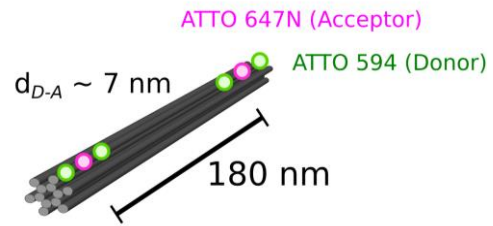
STED-FRET



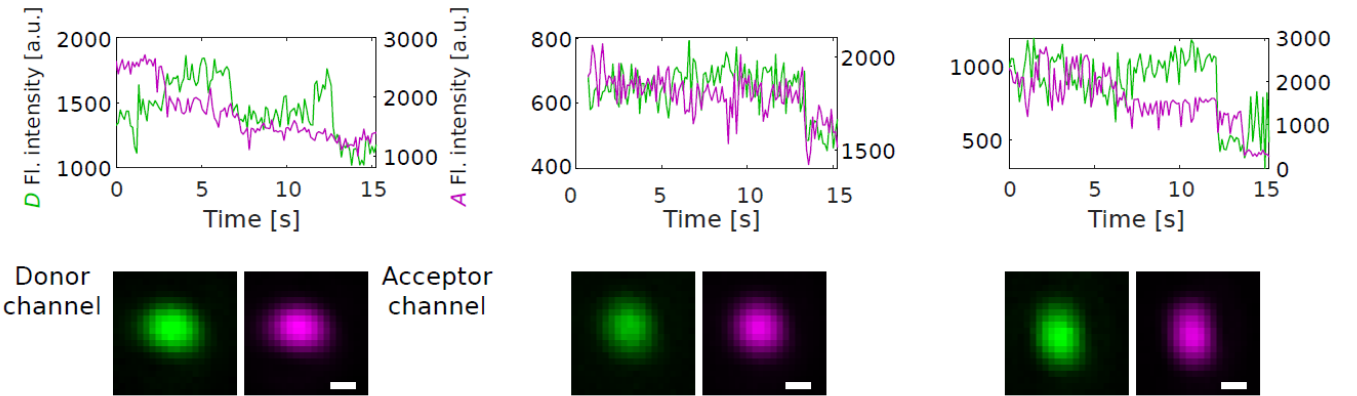
STED-FRET



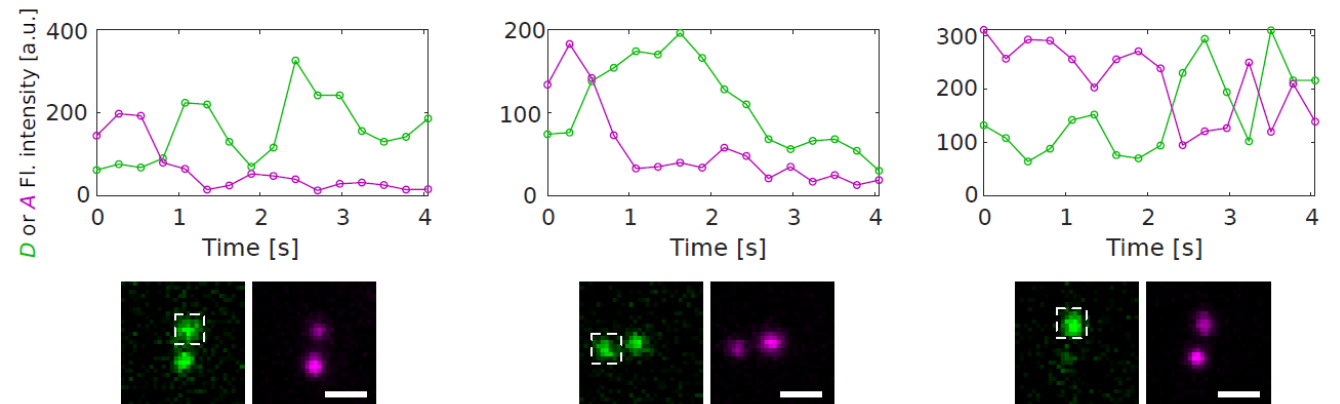
STED-FRET



Confocal FRET



STED-FRET



STED-FRET

$$\frac{k_t}{k_{f,D}} = \left[\frac{\Gamma_0}{r_{DA}} \right]^6 = \frac{1}{\phi_A} \frac{d_D^D}{d_A^A} \mathbf{F} \quad \Gamma_0^6 = \frac{J\kappa^2}{n^4}$$

$$\mathbf{F} = \frac{F_A^D}{F_D^D} - \frac{I^D \sigma_A^D f_{bl} F_A^A}{I^A \sigma_A^A F_D^D} - \frac{d_D^A}{d_D^D}$$

F_D^D and F_A^D of singly-labeled samples $\Rightarrow \frac{d_D^A}{d_D^D} \Rightarrow$

Work-flow:

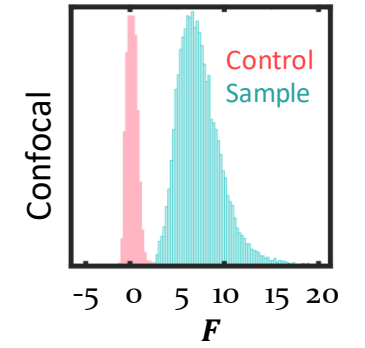
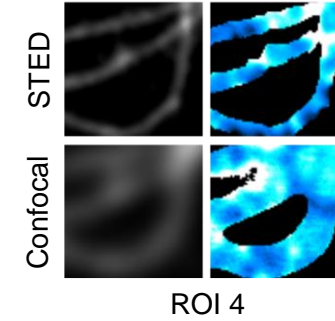
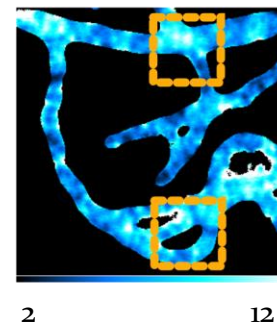
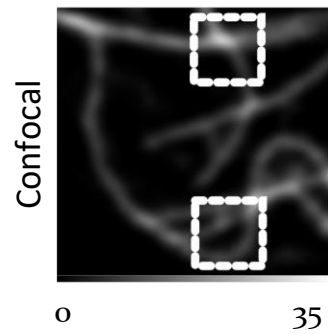
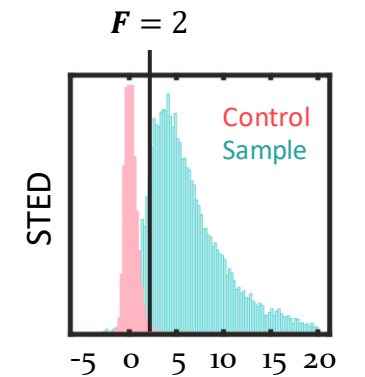
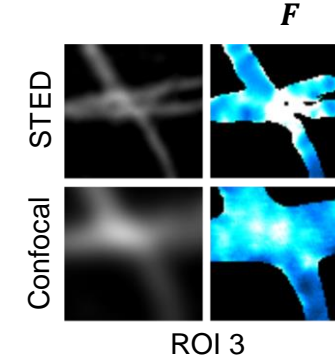
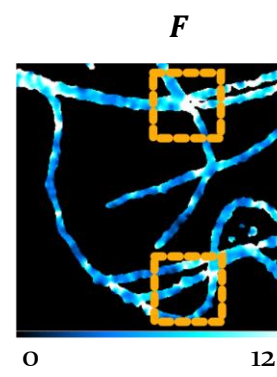
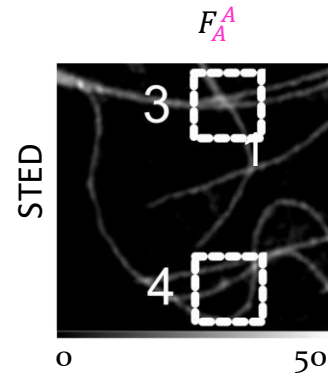
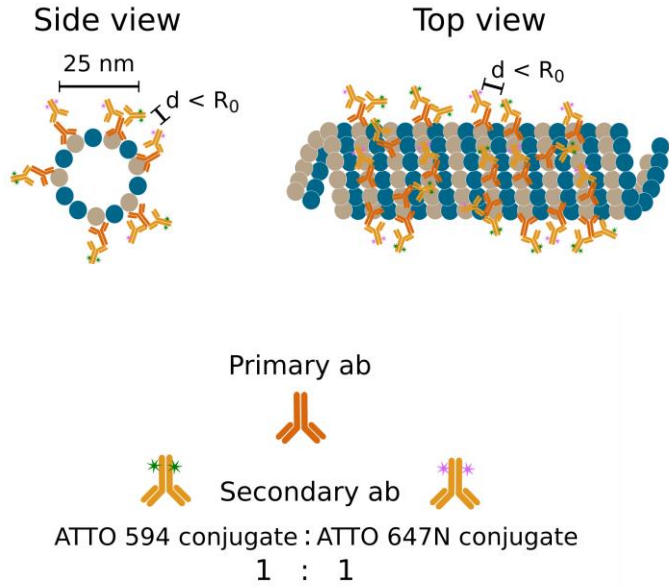
1. Confocal F_A^A
2. STED F_A^A
3. Confocal F_A^A
4. STED $F_D^D + F_A^D$
5. Denoising (optional)
6. Background subtraction
7. Masking
8. f_{bl} from 1. and 3.
9. \mathbf{F} image $\propto k_t$

T. Jovin, E. Jares-Erijman, et al.

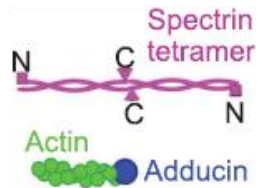
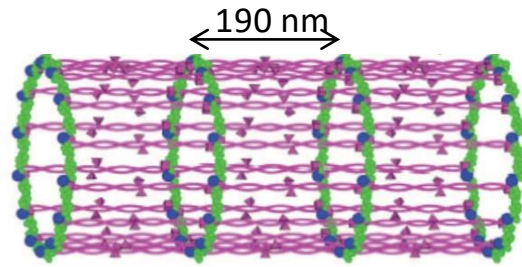
- Chapter 12 in "FRET and FLIM Imaging"
- **ChemPhysChem** 2011, 12 (3), 563–566.

STED-FRET

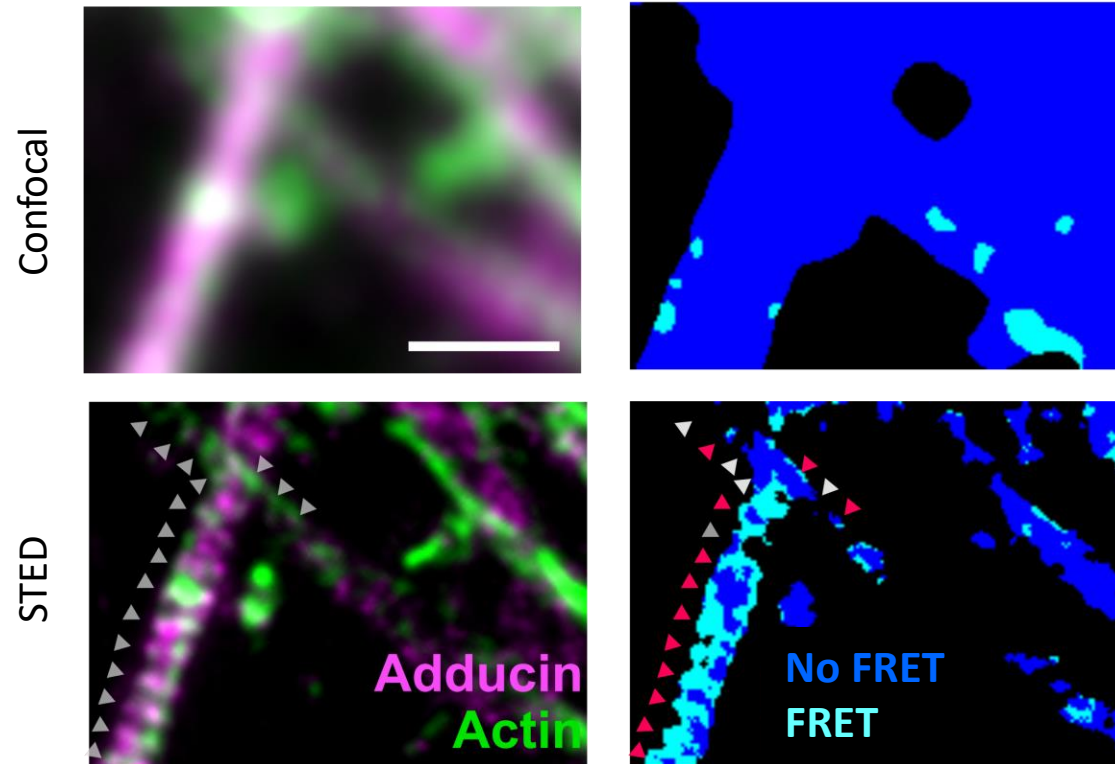
CONTROL to account for local variations of D and A concentrations



Super-resolving biomolecular interactions with STED-FRET



Example neuron



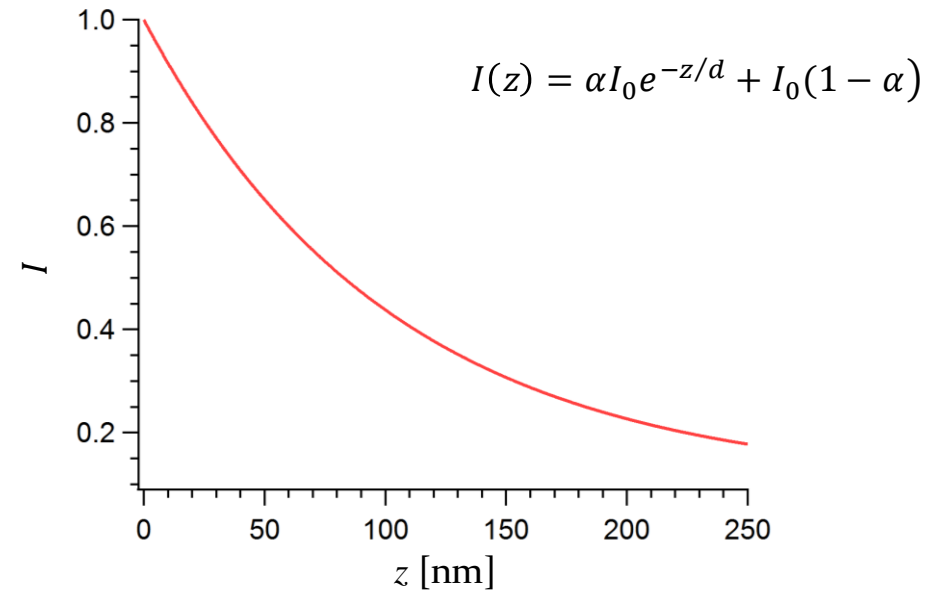
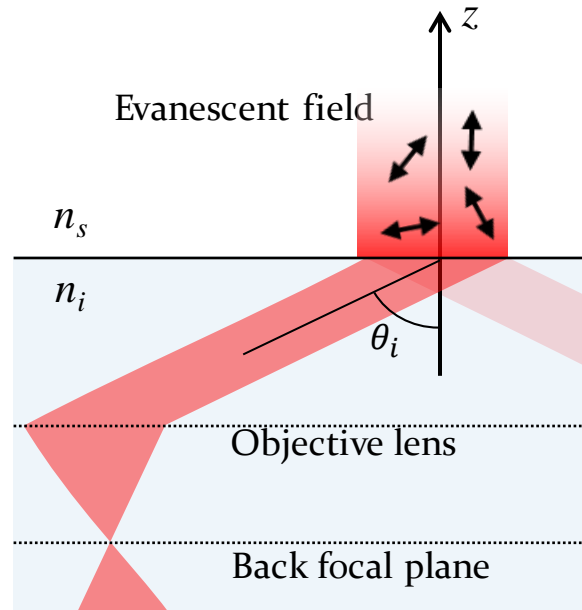
SIMPLER 3D TIRF Nanoscopy

Supercritical Illumination Microscopy Photometric z-Localization w/ Enhanced Resolution

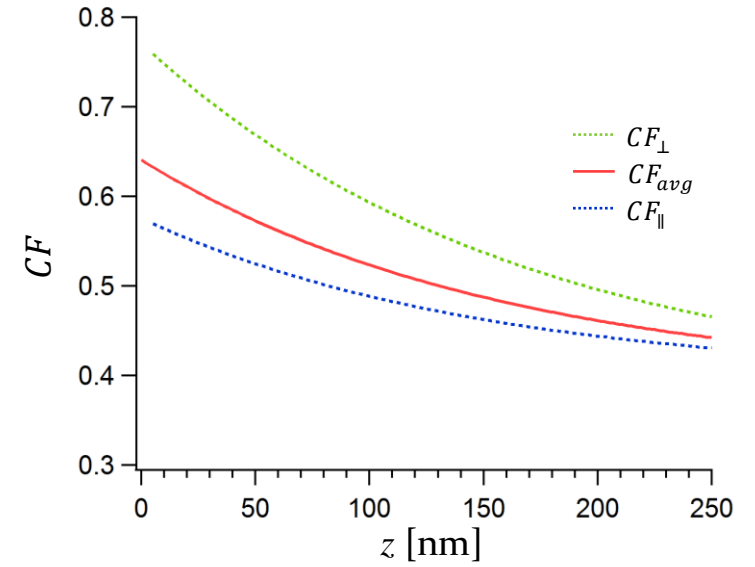
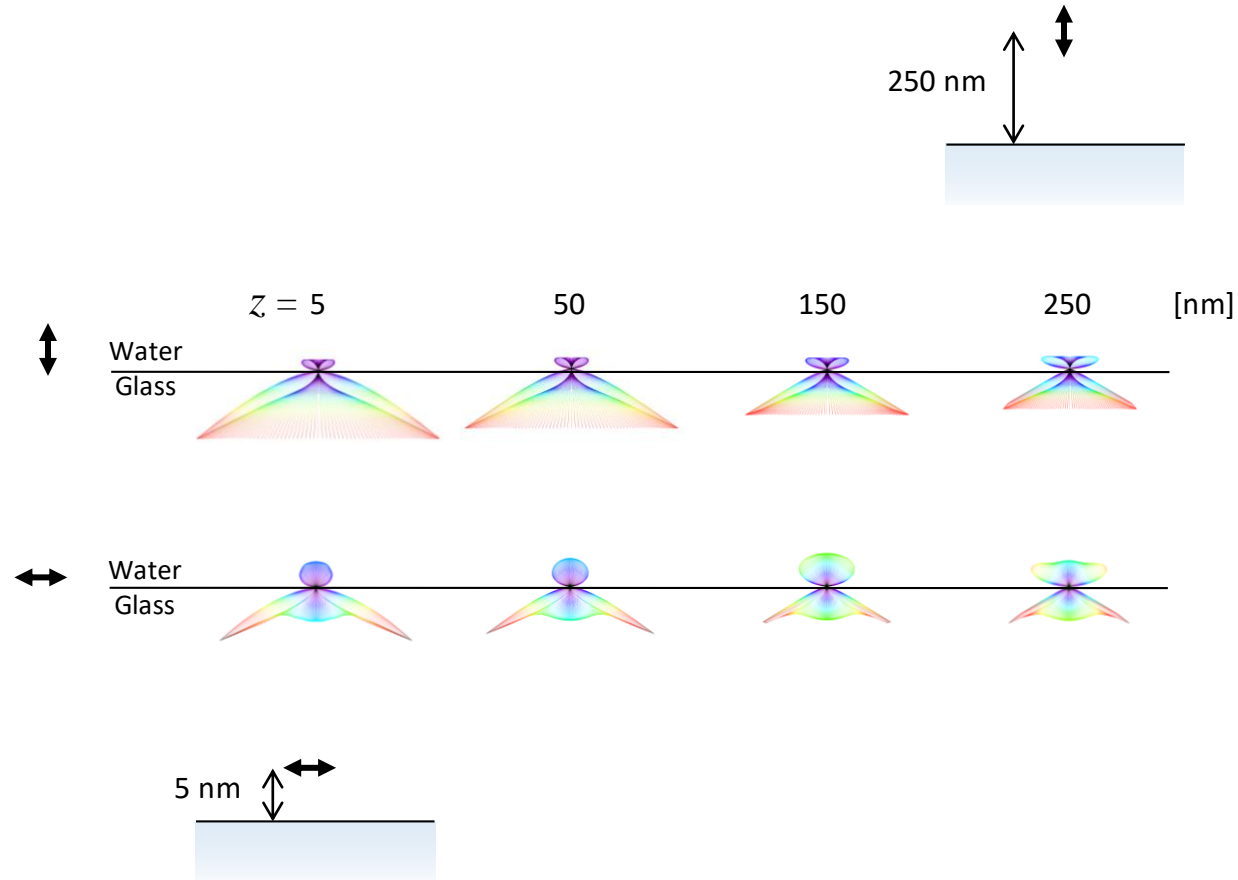
*Szalai et al. **Nature Communications** 12 (2021) 517*

<https://stefani-lab.ar/>

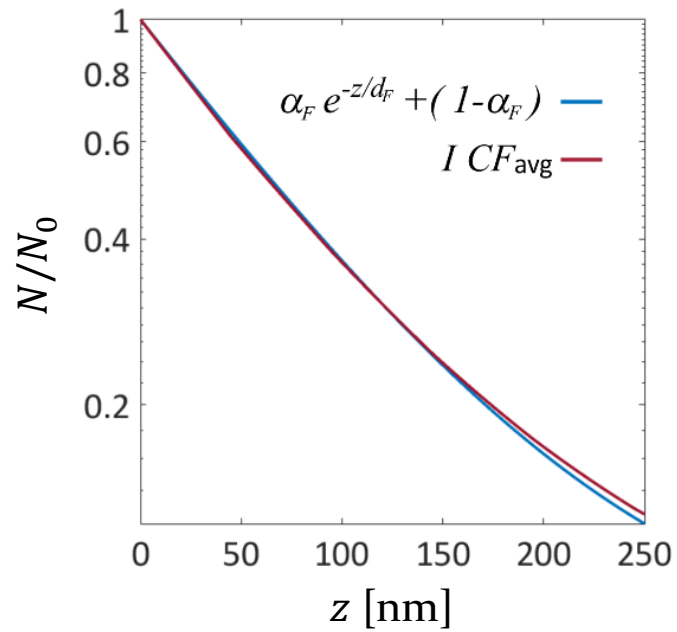
SIMPLER 3D TIRF nanoscopy



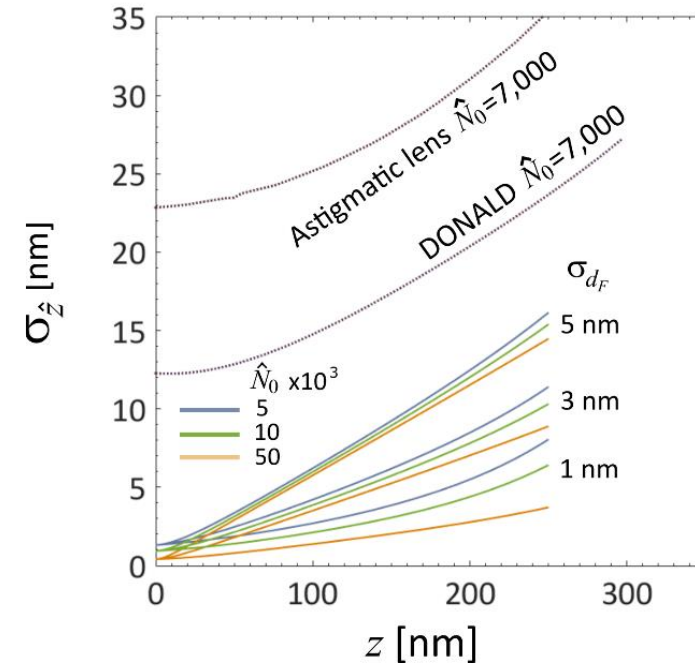
SIMPLER 3D TIRF nanoscopy



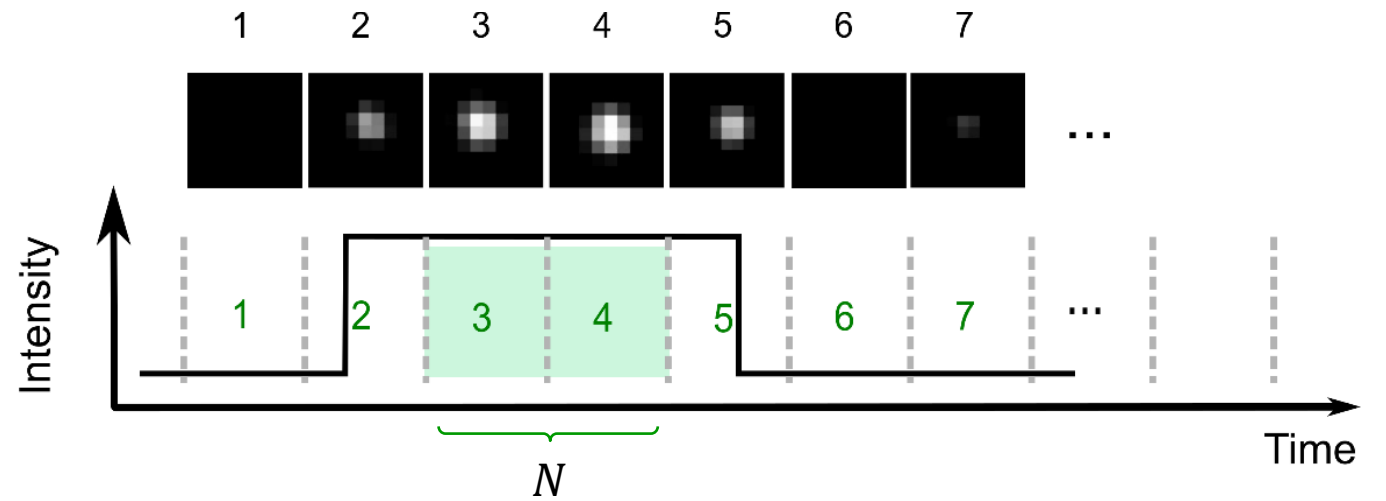
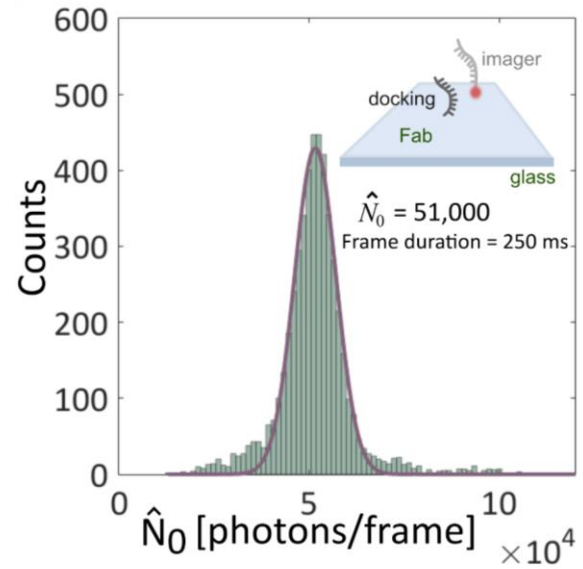
SIMPLER 3D TIRF nanoscopy



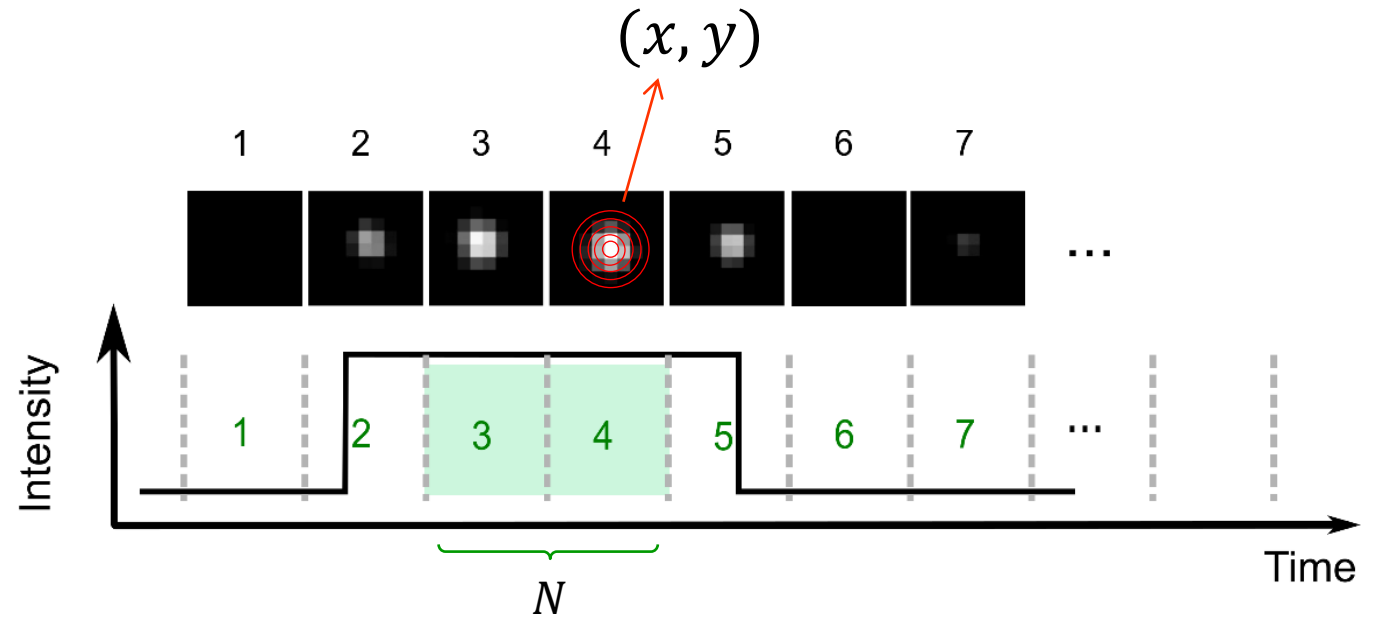
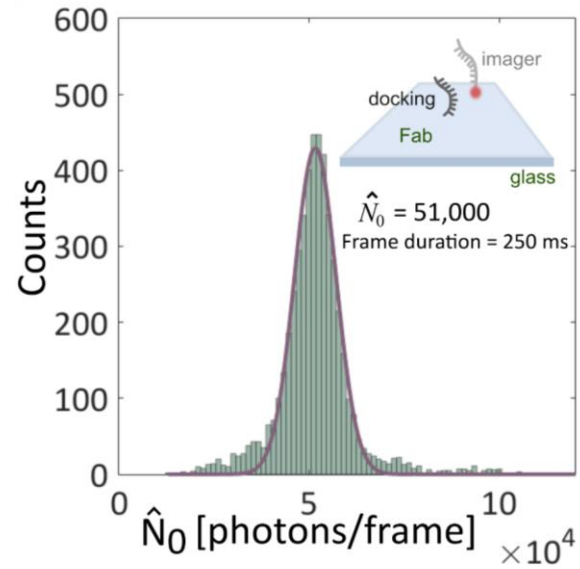
$$\frac{N(z)}{N_0} = \alpha_F e^{-z/d_F} + (1 - \alpha_F)$$



SIMPLER 3D TIRF nanoscopy

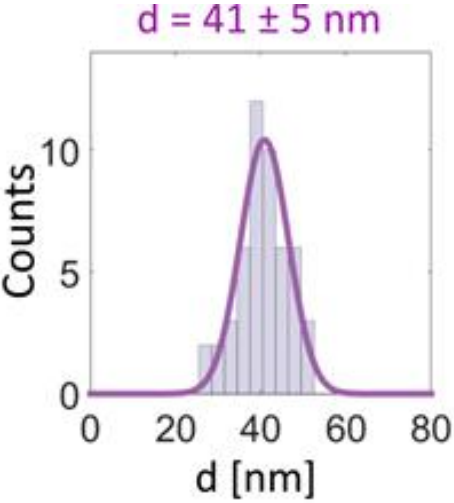
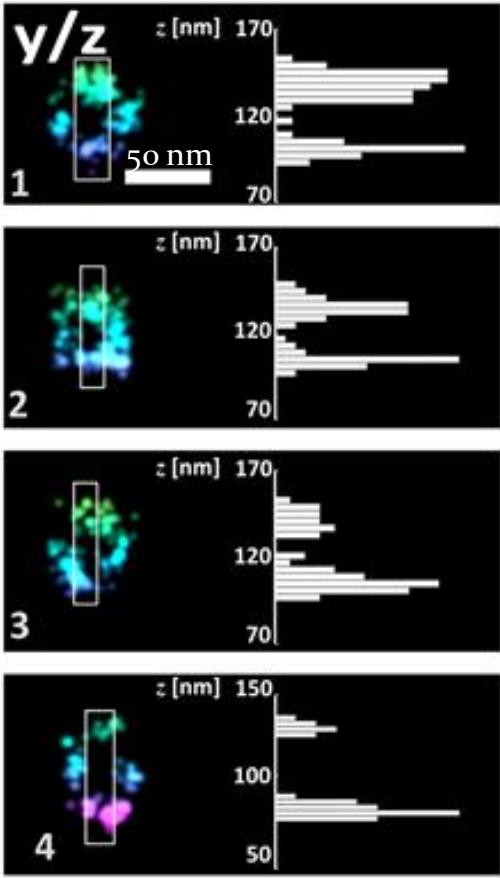
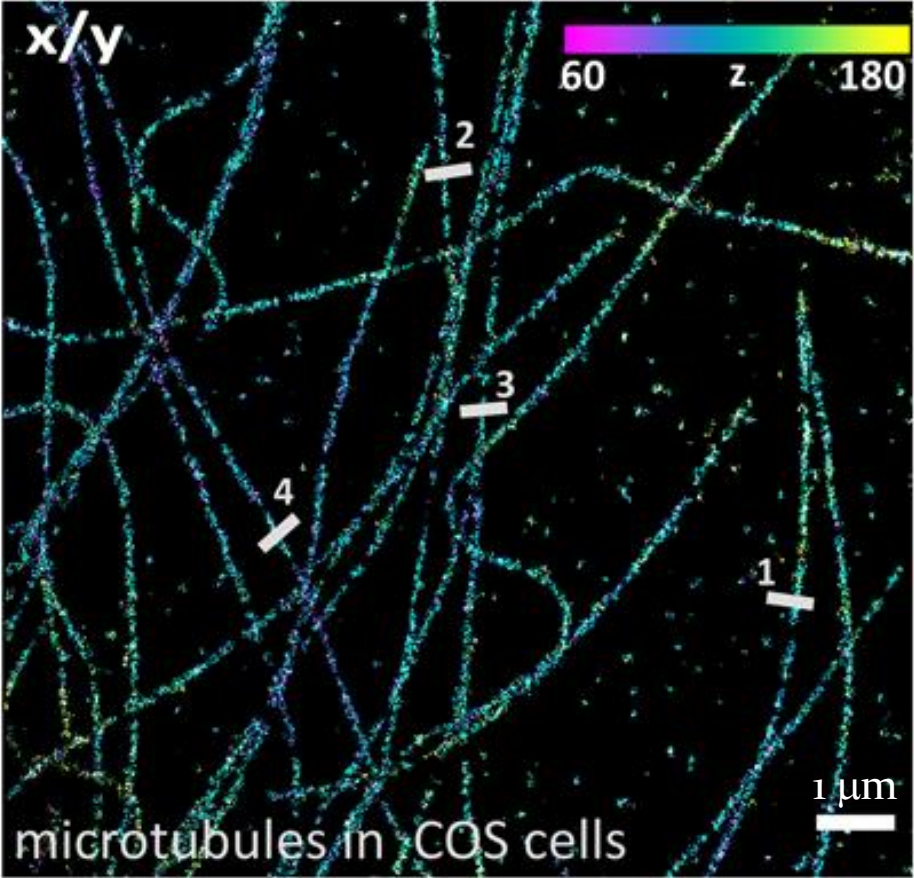


SIMPLER 3D TIRF nanoscopy

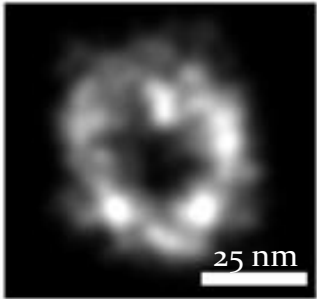


$$\frac{N}{N_0} \rightarrow z$$

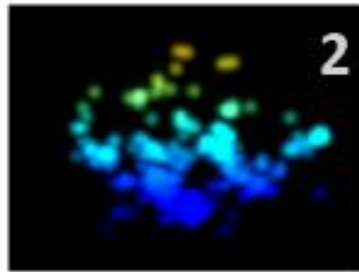
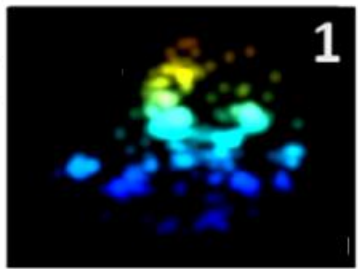
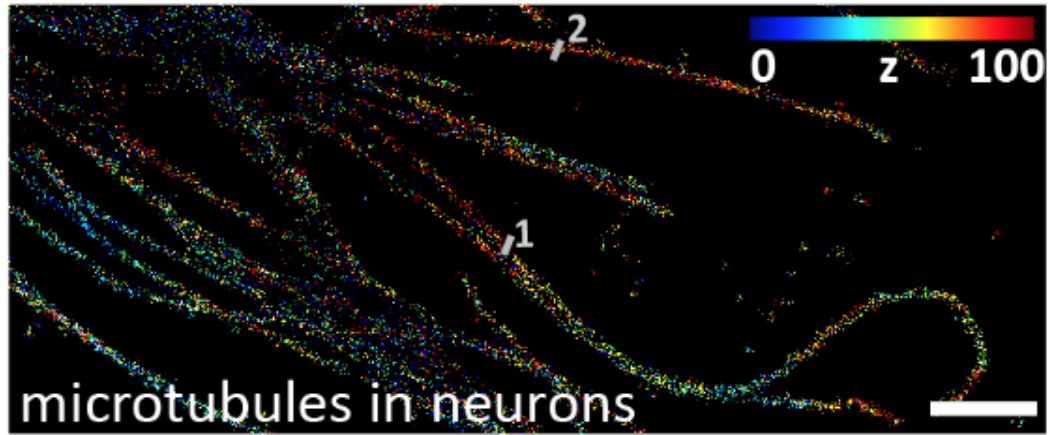
SIMPLER 3D TIRF nanoscopy



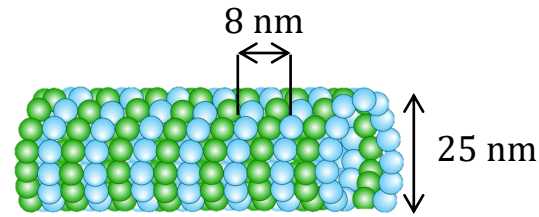
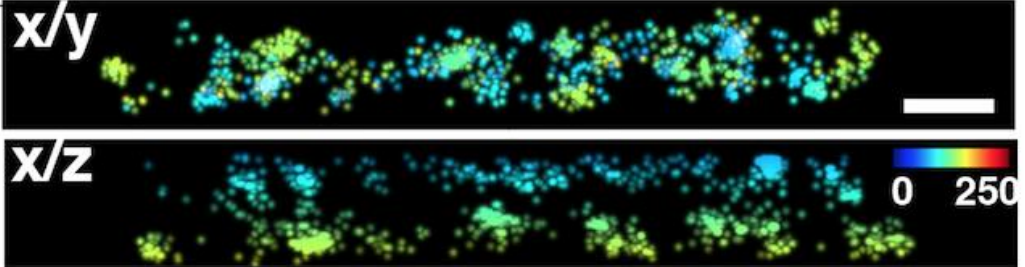
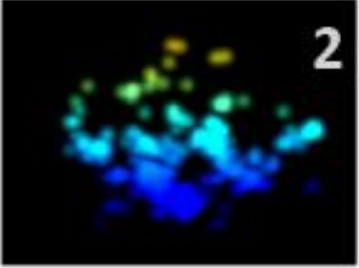
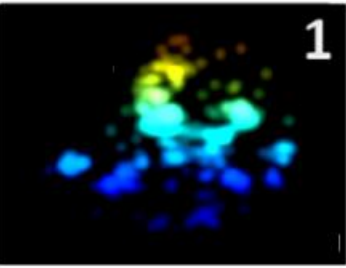
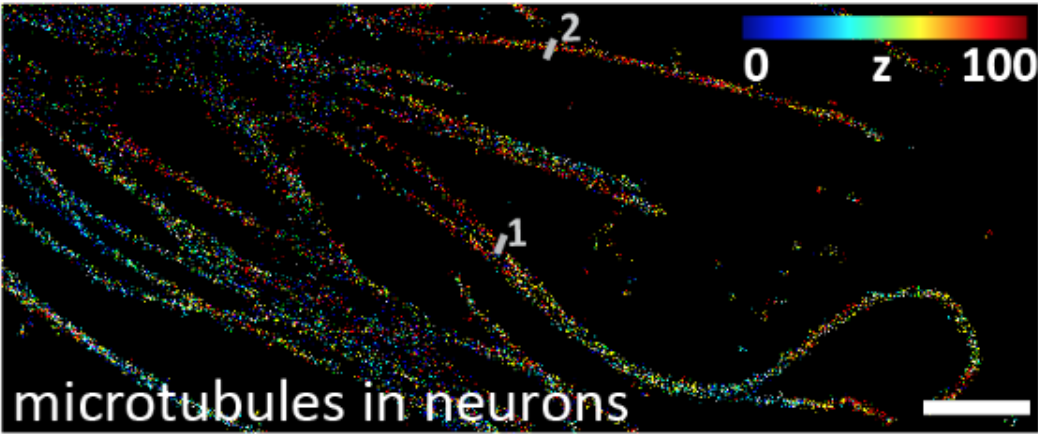
average MT



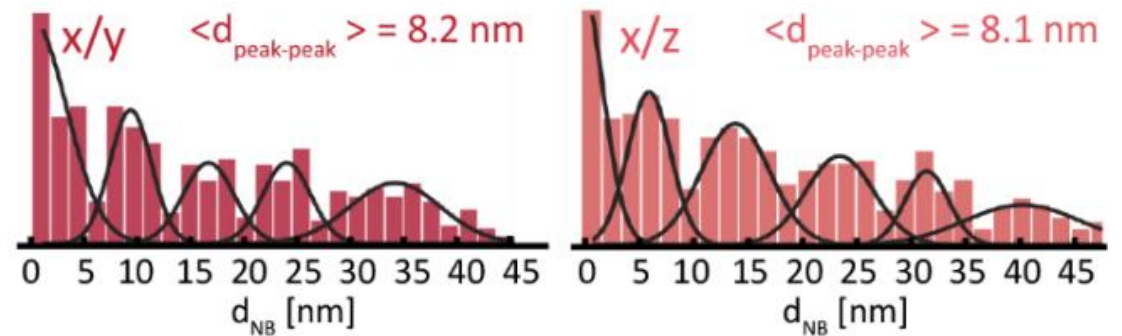
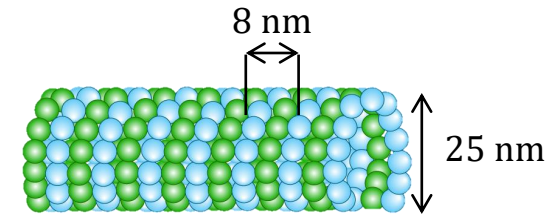
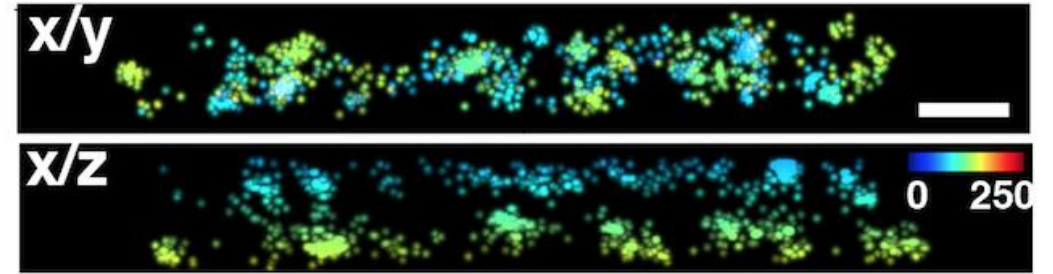
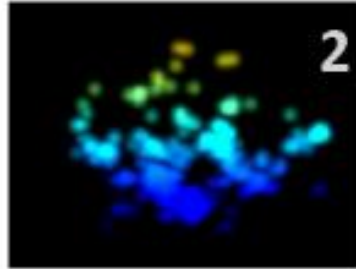
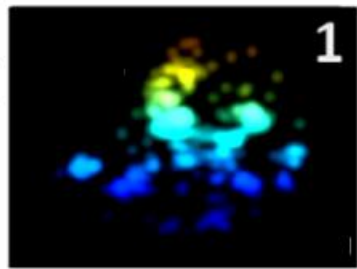
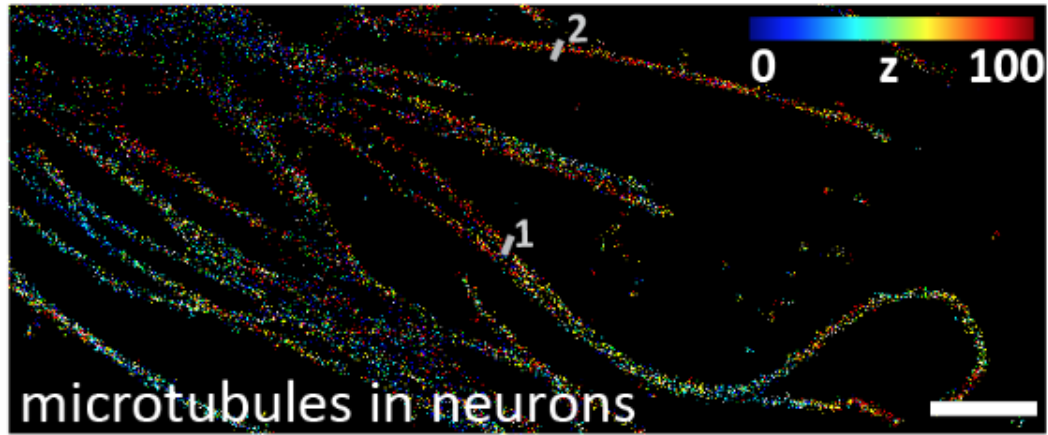
SIMPLER 3D TIRF nanoscopy



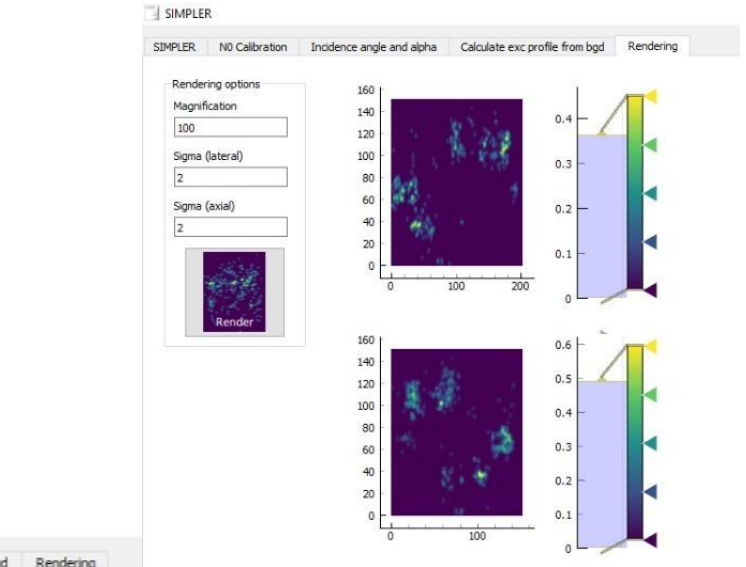
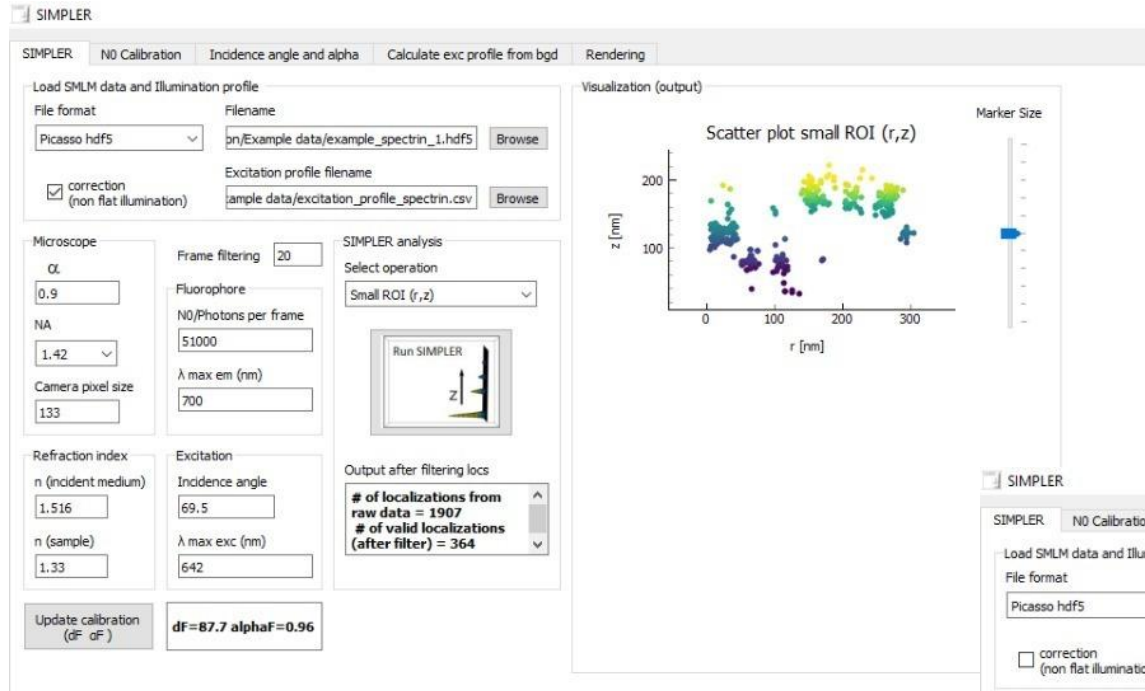
SIMPLER 3D TIRF nanoscopy



SIMPLER 3D TIRF nanoscopy

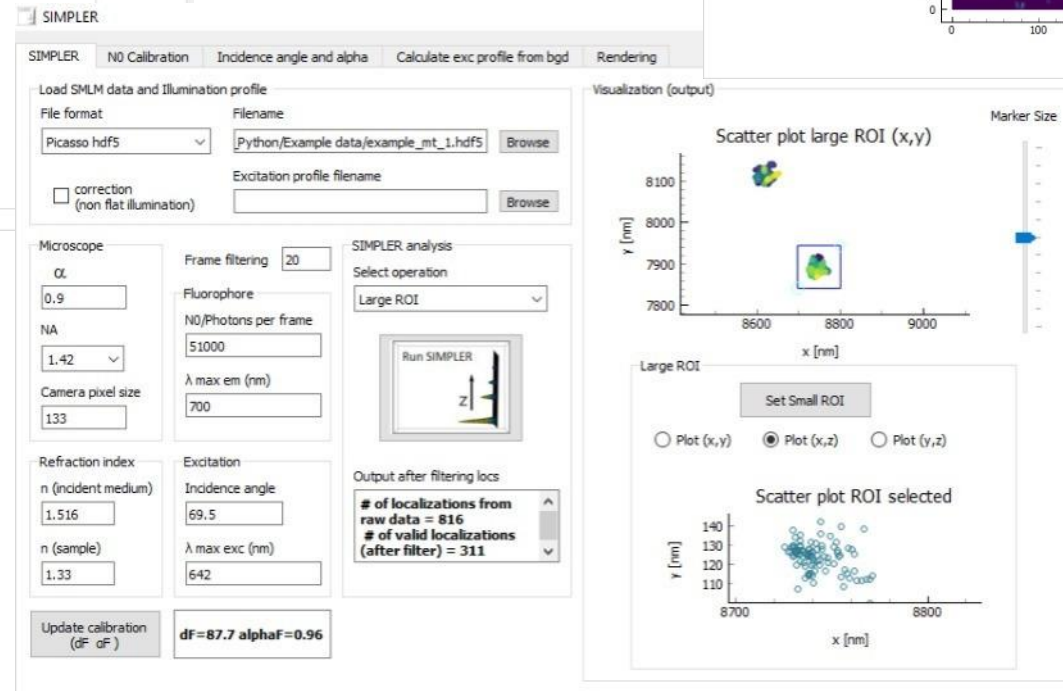


SIMPLER 3D TIRF nanoscopy



Open-source Python software

<https://github.com/Stefani-Lab>



SML-SSI Single-Molecule Localization with Sequential Structured Illumination

Balzarotti et al. *Science* 355 (2017) 606-612

Masullo et al. *Nano Letters* 21 (2021) 840-846

Masullo et al. *Biophysical Reports* 2 (2022) 100036

Stefani, F. D. *Nature Photonics* 17 (2023) 552-553

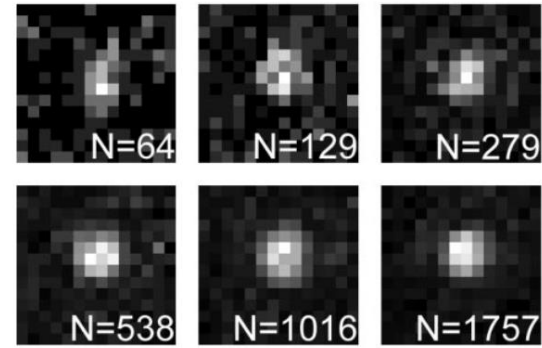
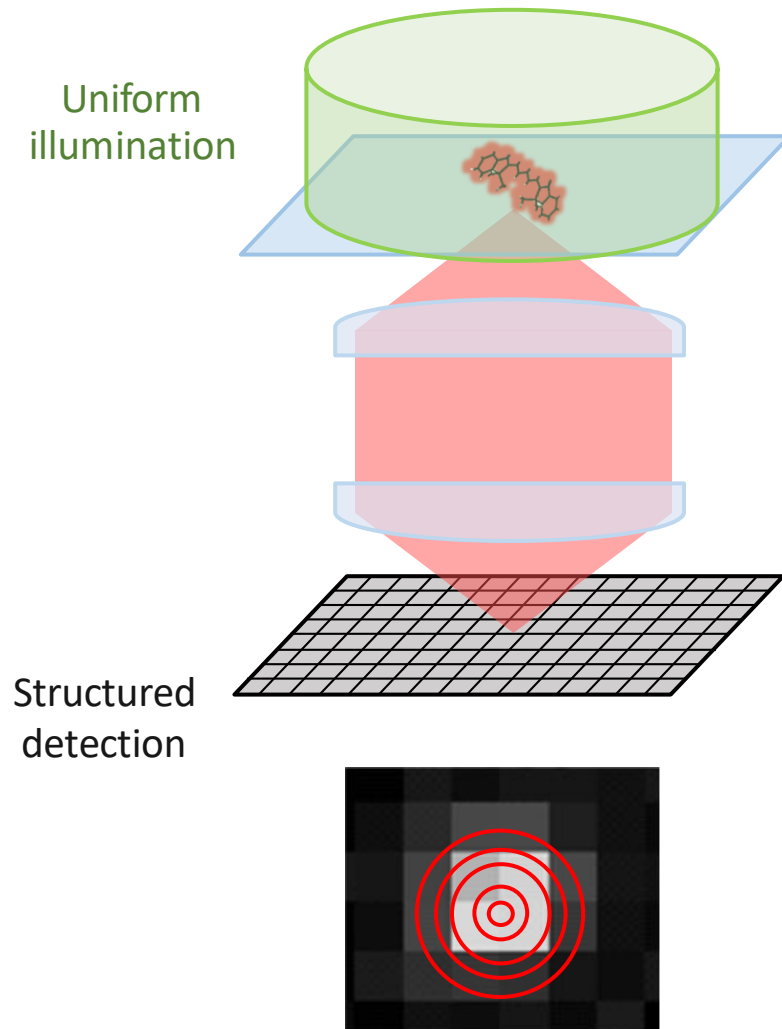
Masullo et al. *Light: Science & Applications* 11 (2022) 70

Masullo et al. *Light: Science & Applications* 11 (2022) 199

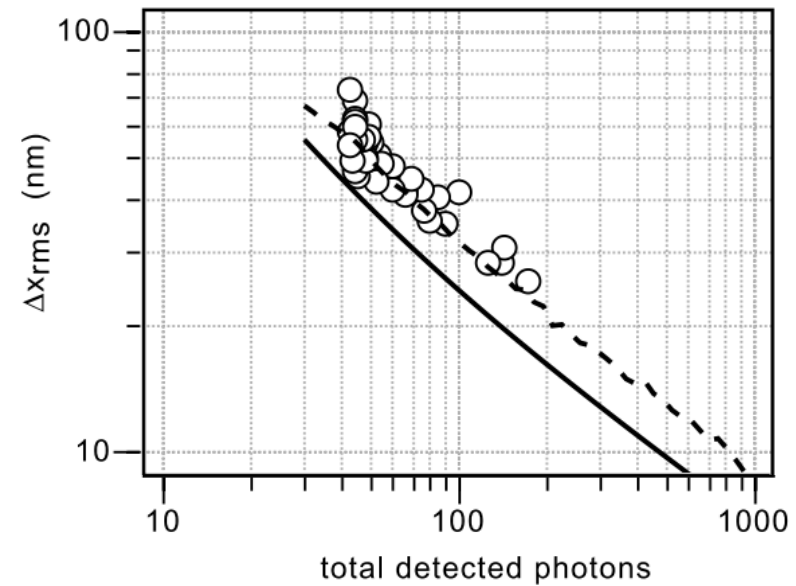
Zdańkowski et al. *ACS Photonics* 9 (2022) 3777–3785

Cole et al. *Nature Photonics* (2024) – published online

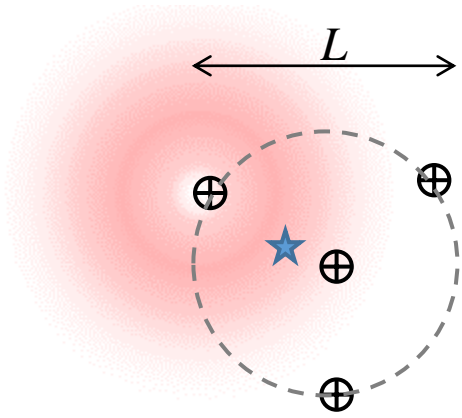
Camera-based single molecule localization



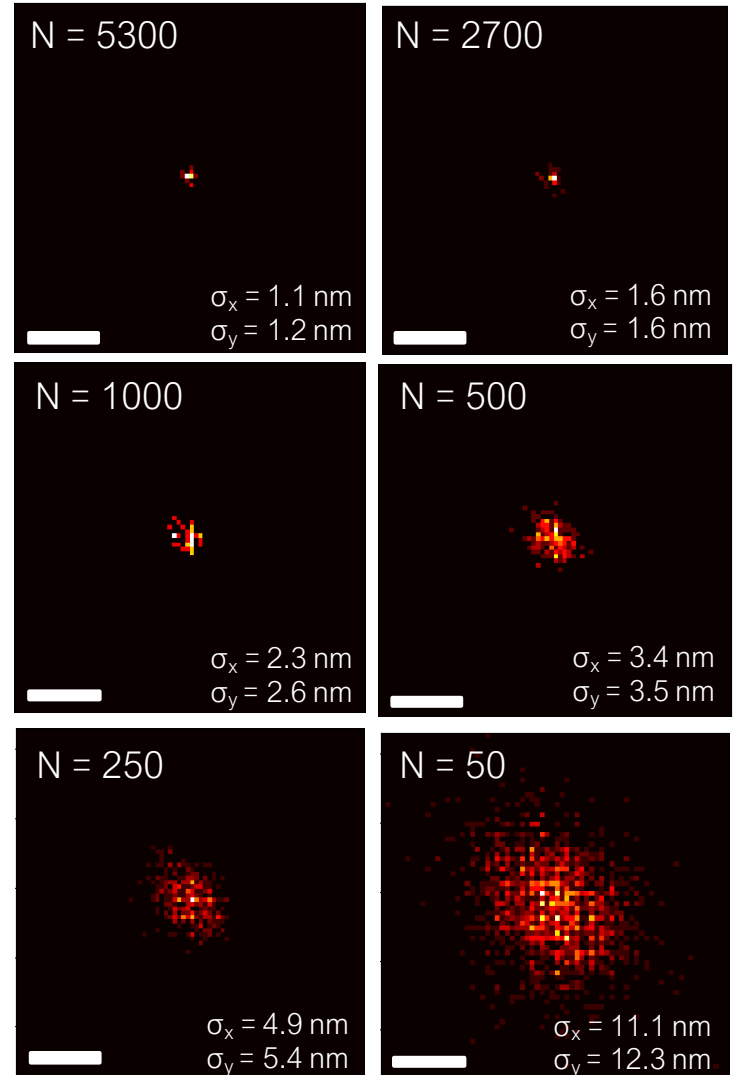
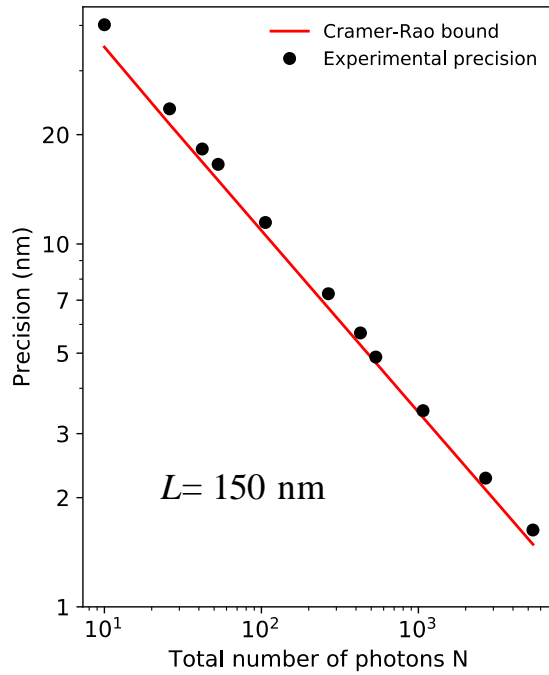
$$\sigma \approx PSF / \sqrt{N}$$



MINFLUX



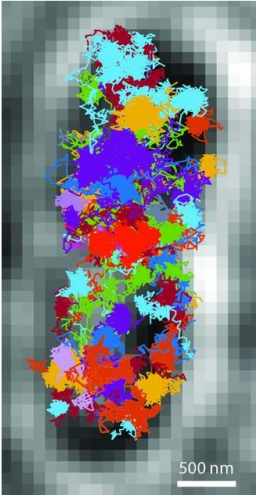
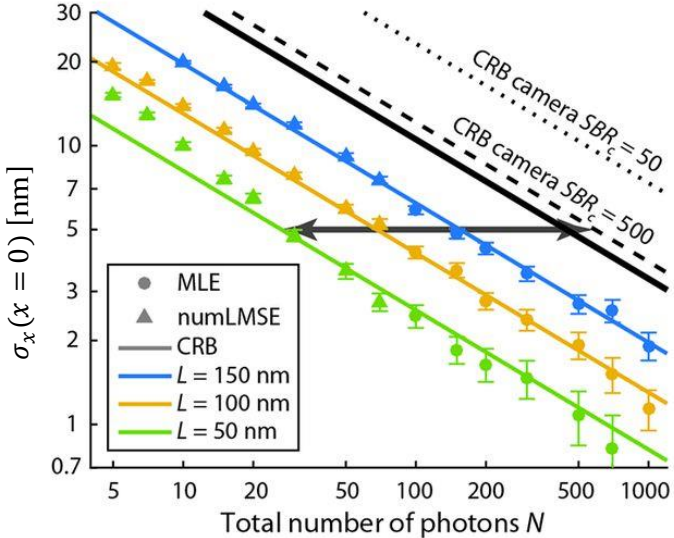
$$\sigma_{CRB} \propto \frac{L}{\sqrt{N}}$$



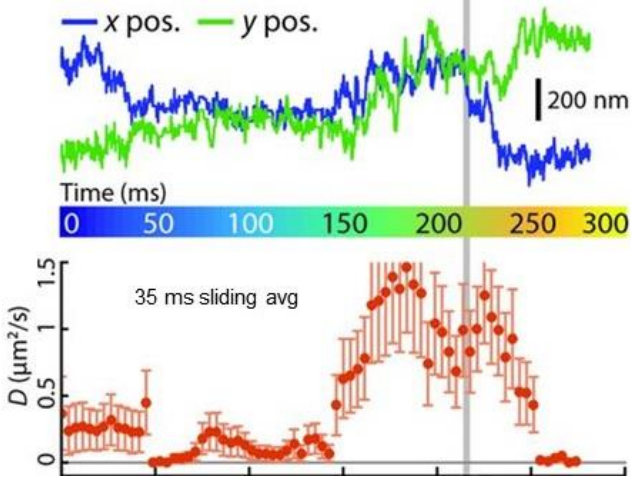
Scale bar: 20 nm

MINFLUX

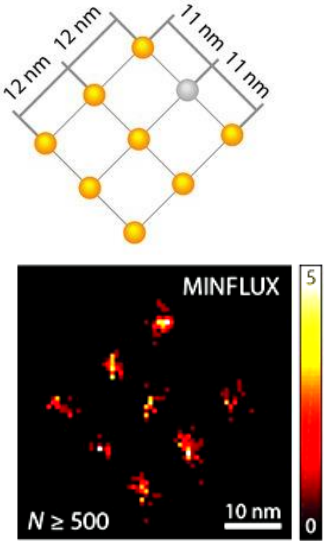
Tunable nanometer resolution



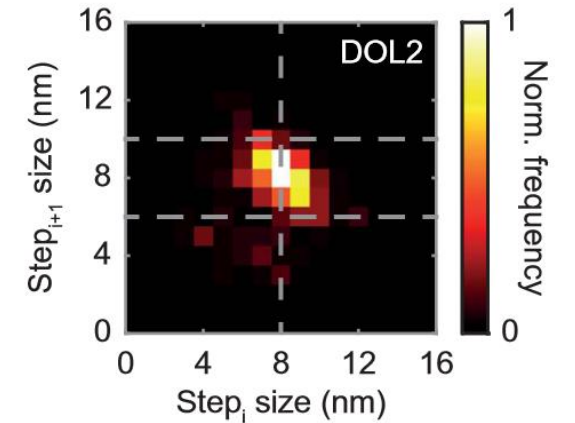
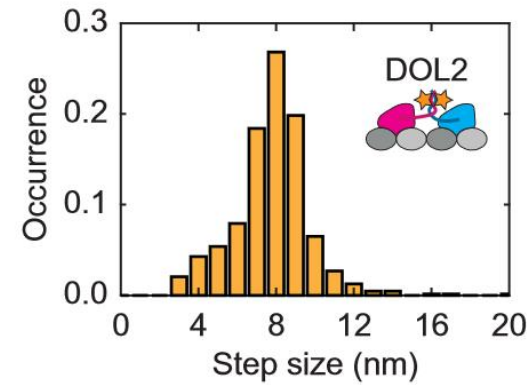
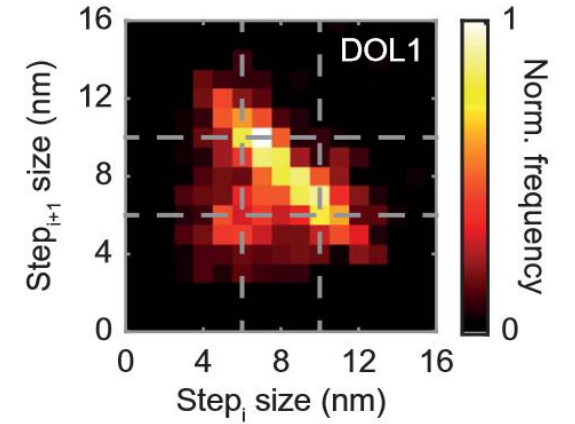
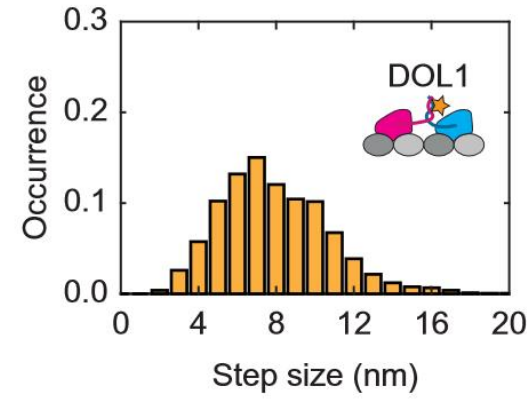
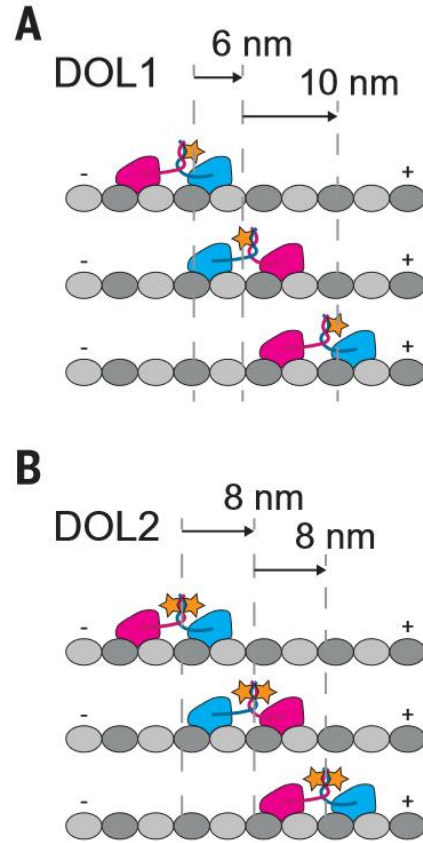
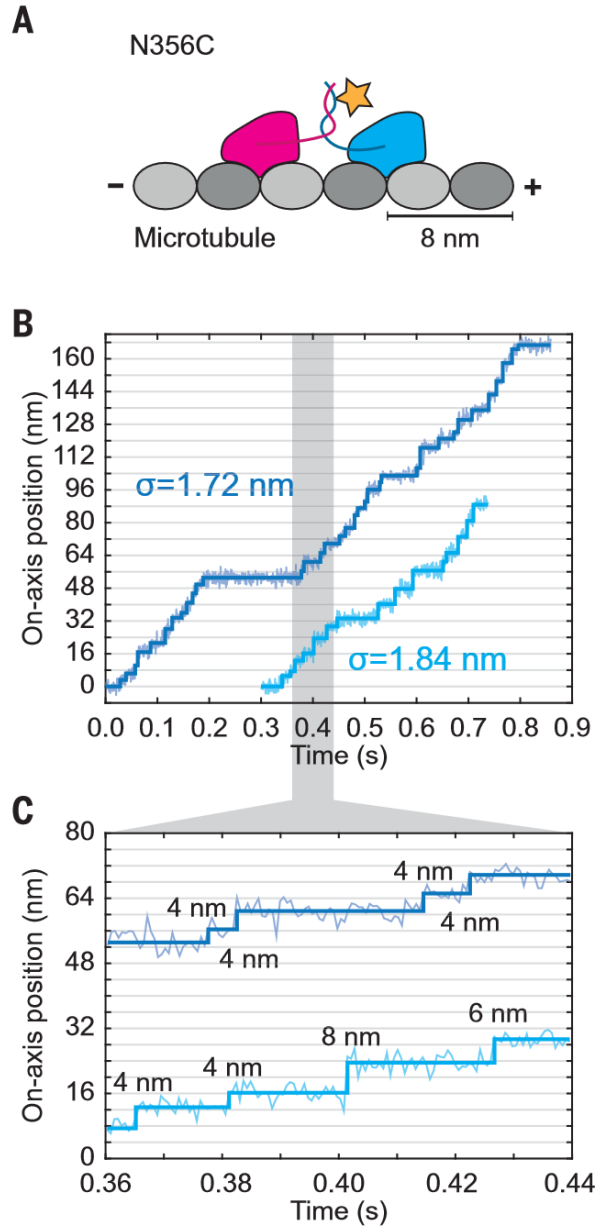
Superfast tracking



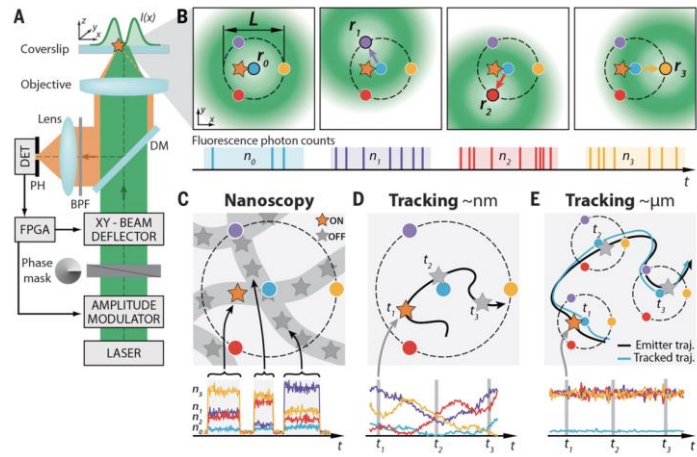
Nanoscopy



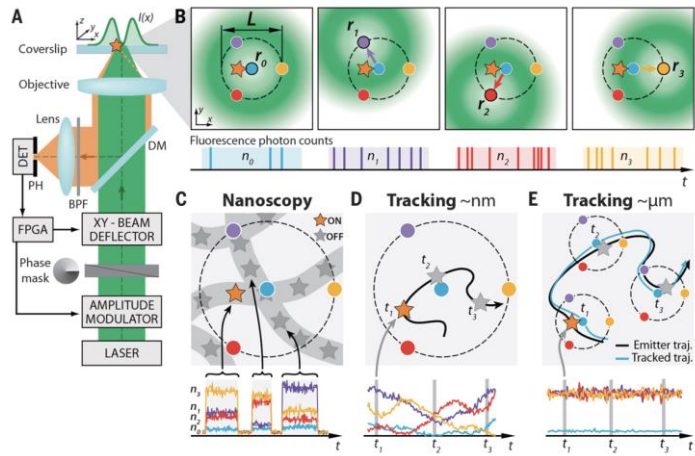
MINFLUX



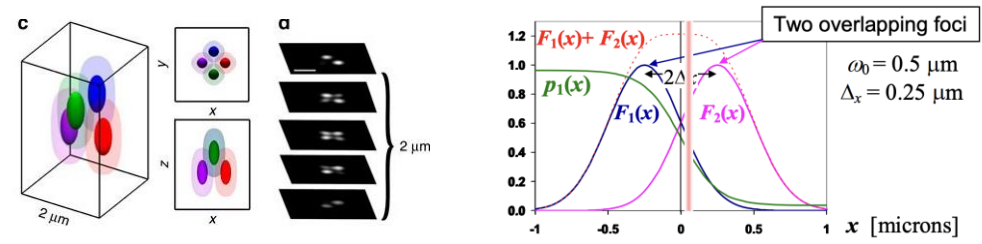
MINFLUX Balzarotti et al, *Science* (2017)



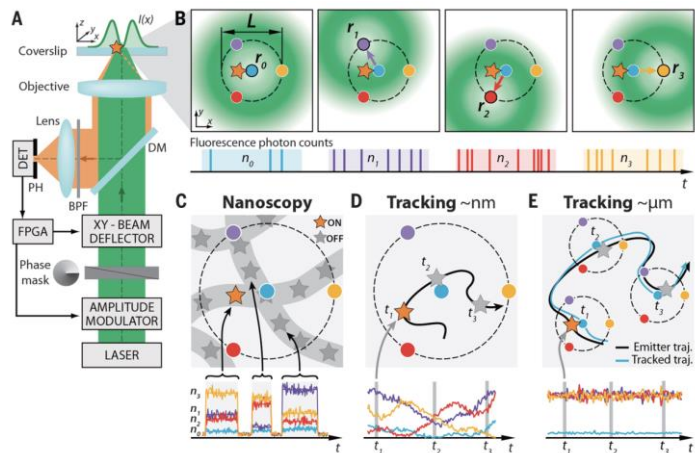
MINFLUX Balzarotti et al, *Science* (2017)



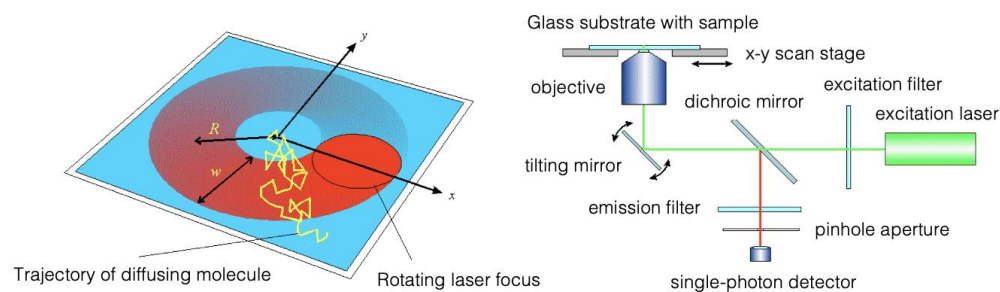
Four-focus particle tracking Perillo et al, *Nat. Comm.* (2015) Davis et al, *Opt. Express* (2014)



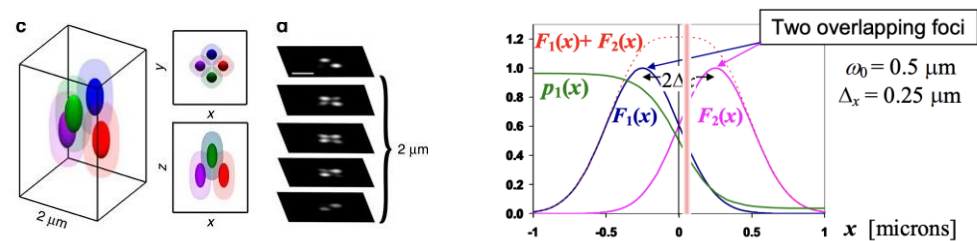
MINIFLUX Balzarotti et al, *Science* (2017)



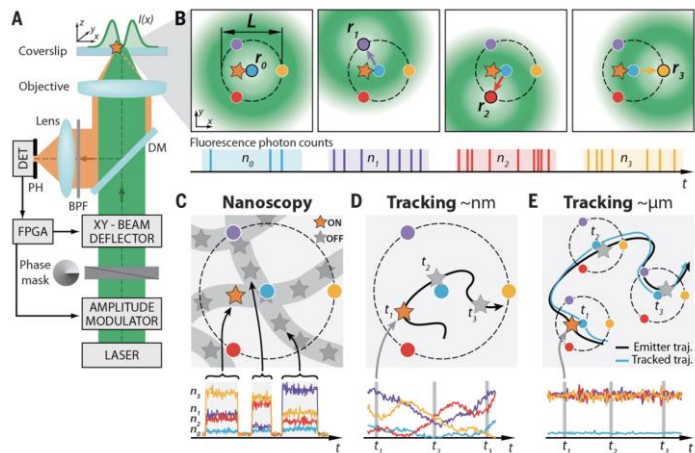
Orbital tracking Enderlein, *Appl. Phys. B Lasers Opt.* (2000) Levi et al, *Biochem. Soc. Trans.* (2003)



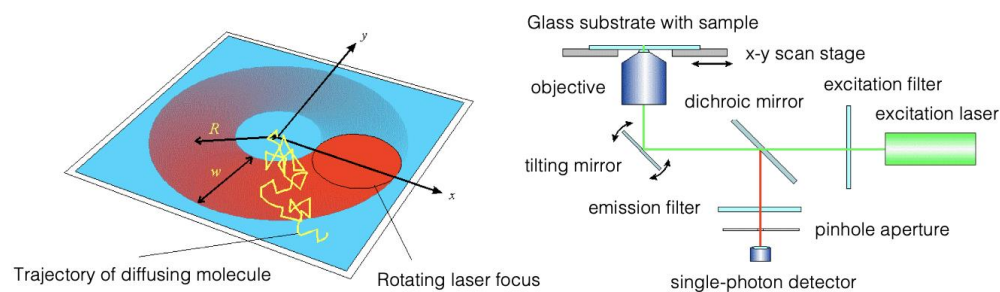
Four-focus particle tracking Perillo et al, *Nat. Comm.* (2015) Davis et al, *Opt. Express* (2014)



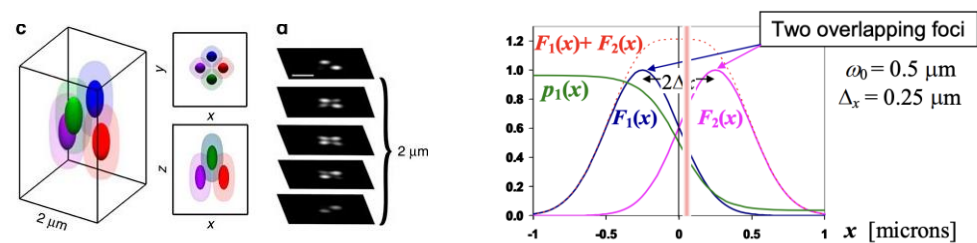
MINIFLUX Balzarotti et al, *Science* (2017)



Orbital tracking Enderlein, *Appl. Phys. B Lasers Opt.* (2000) Levi et al, *Biochem. Soc. Trans.* (2003)

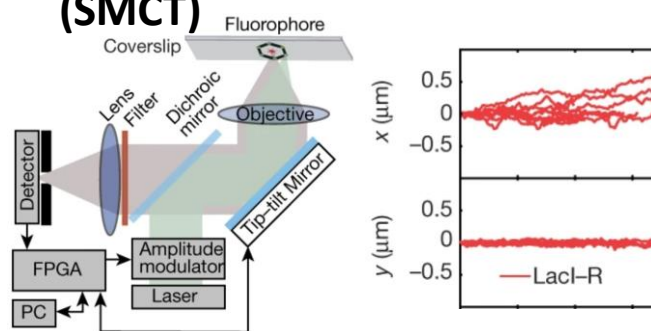


Four-focus particle tracking Perillo et al, *Nat. Comm.* (2015) Davis et al, *Opt. Express* (2014)

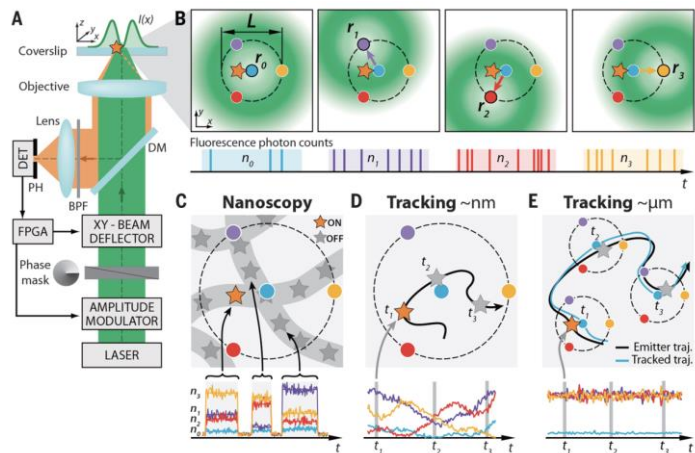


Single-molecule confocal tracking (SMCT)

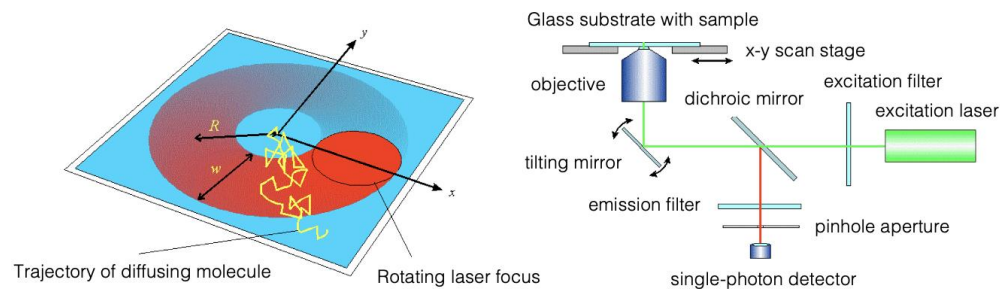
Marklund et al, *Nature* (2020)



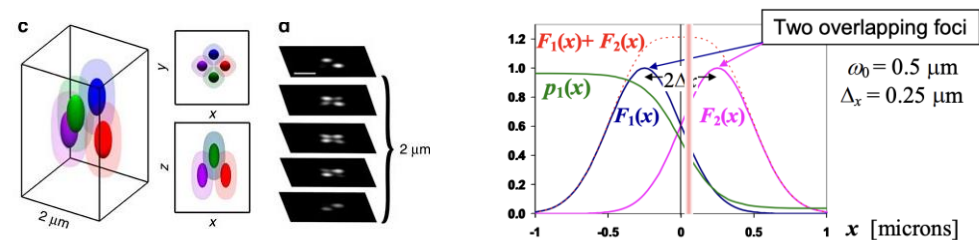
MINFLUX Balzarotti et al, *Science* (2017)



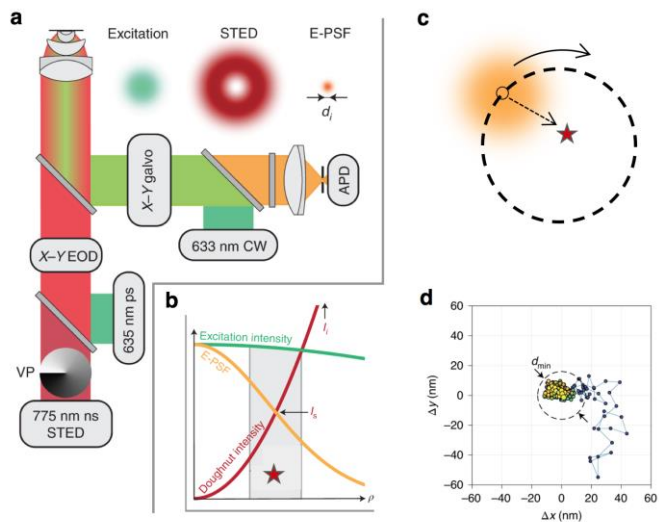
Orbital tracking Enderlein, *Appl. Phys. B Lasers Opt.* (2000) Levi et al, *Biochem. Soc. Trans.* (2003)



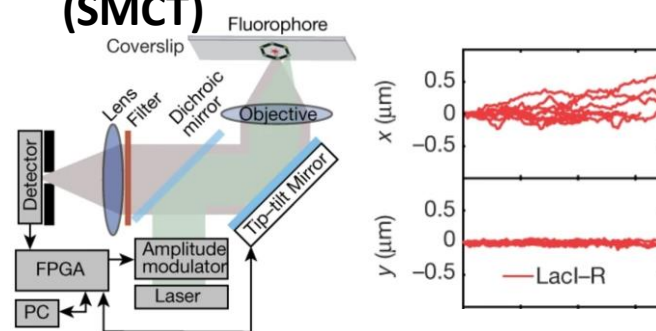
Four-focus particle tracking Perillo et al, *Nat. Comm.* (2015) Davis et al, *Opt. Express* (2014)



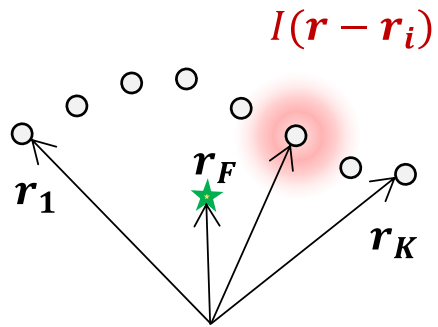
MINSTED Weber et al, *Nature Photonics* (2021)



Single-molecule confocal tracking (SMCT) Marklund et al, *Nature* (2020)

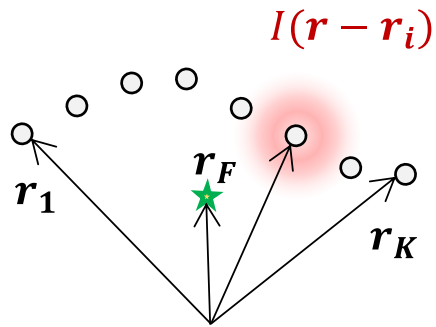


Common framework for SML-SSI



Common framework for SML-SSI

Goal: inferring the position of the emitter \mathbf{r}_F , given:

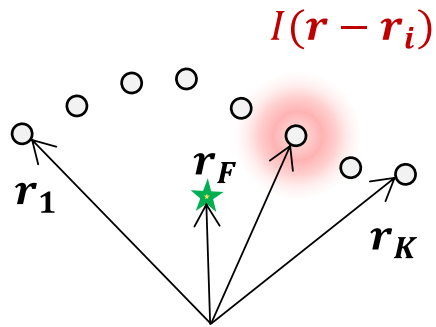


$I(\mathbf{r})$ The structure of the illumination

\mathbf{r}_i The sequence of beam positions

n_i The detected signal in each position

Common framework for SML-SSI



Goal: inferring the position of the emitter \mathbf{r}_F , given:

- $I(\mathbf{r})$ The structure of the illumination
- \mathbf{r}_i The sequence of beam positions
- \mathbf{n}_i The detected signal in each position

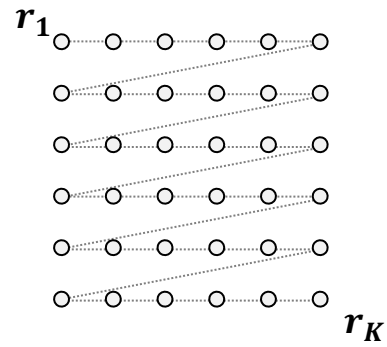


Maximum likelihood estimation

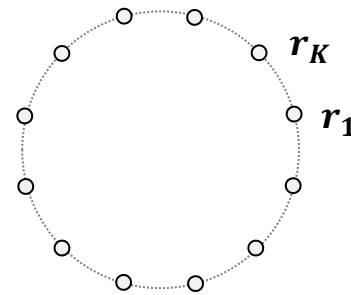
$$\mathcal{L}(\mathbf{r}_E | I(\mathbf{r} - \mathbf{r}_i), \mathbf{n}_i)$$
$$\widehat{\mathbf{r}}_E^{MLE} = \arg \max \mathcal{L}$$

Common framework for SML-SSI

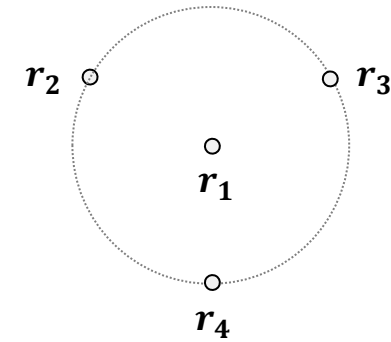
Raster scanning



Orbital scanning

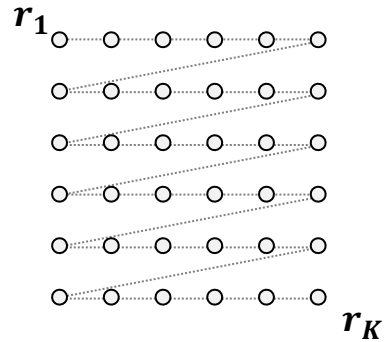


MINFLUX

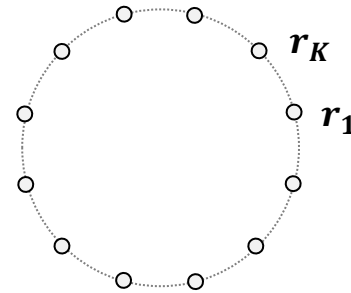


Common framework for SML-SSI

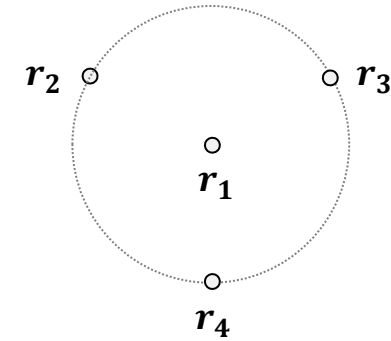
Raster scanning



Orbital scanning



MINFLUX



Intensity MAX

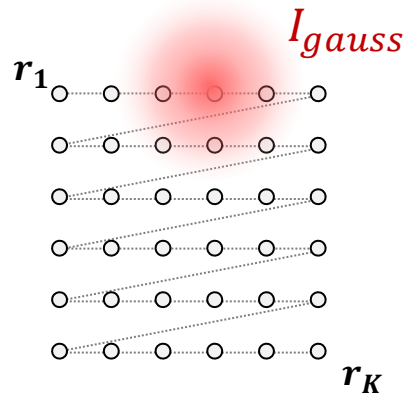


Intensity MIN



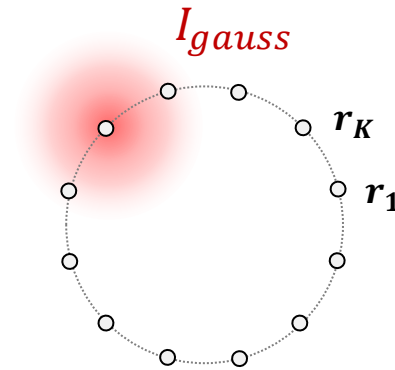
Common description of known methods

Confocal SMLM



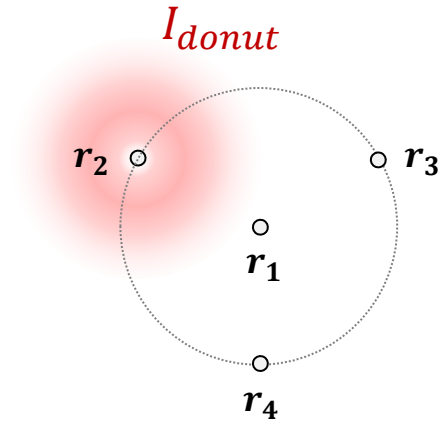
Thiele et al. *ACS Nano* 2020
Zaza et al. *Small Methods* 2023

Orbital tracking / MINSTED



Enderlein *Appl. Phys. B* 2000
Kis-Petikova, et al. *Microsc. Res. Tech.* 2004
Wehnekamp et al. *eLife* 2019
Marklund et al. *Nature* 2020
Weber et al. *Nature Phot.* 2021

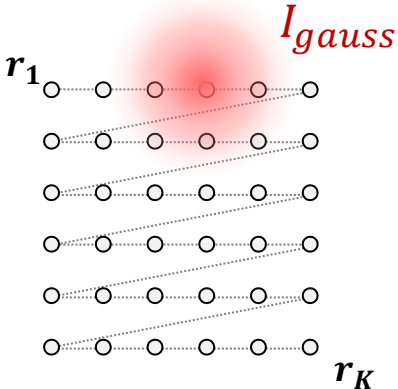
MINFLUX



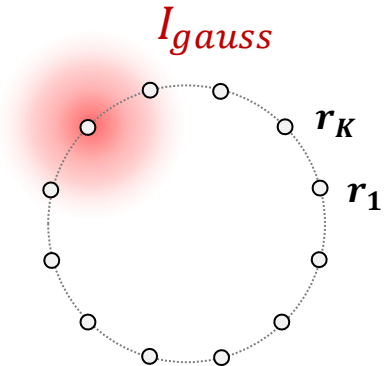
Balzarotti et al. *Science* 2017
Masullo et al. *Nano Lett.* 2021

Common description of known and **new methods**

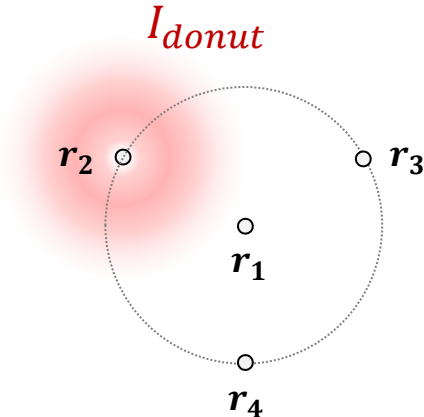
RASTMAX



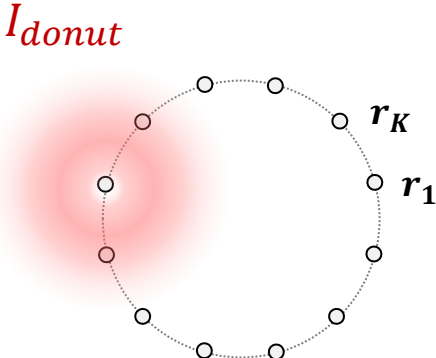
OT



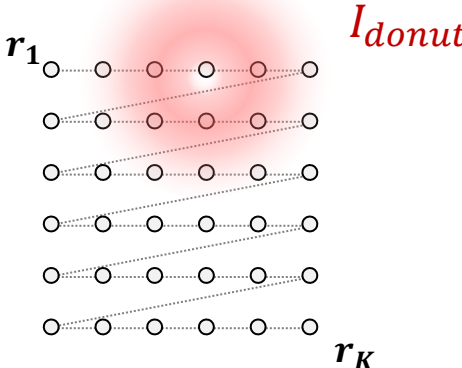
MINFLUX



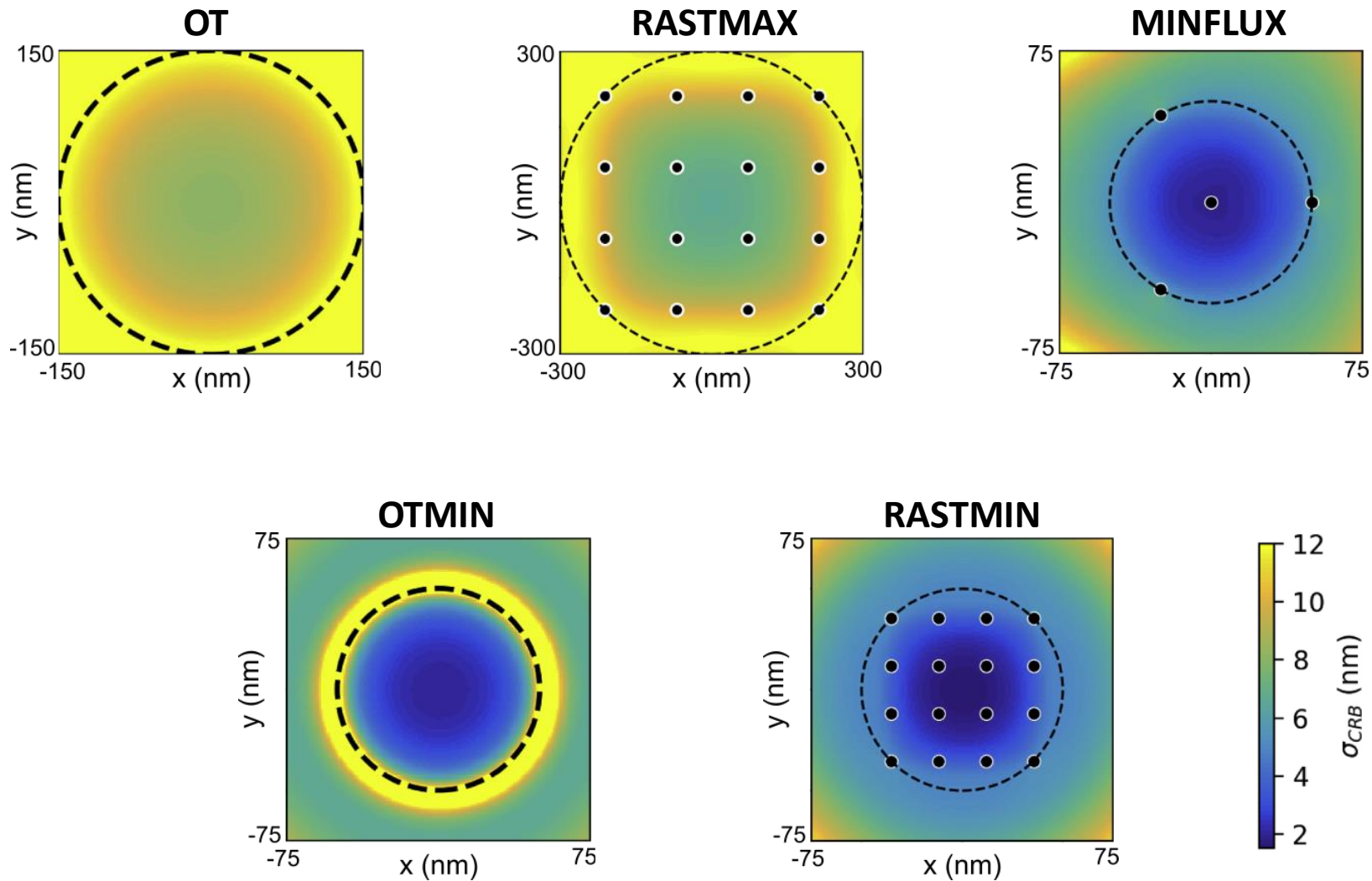
OTMIN



RASTMIN

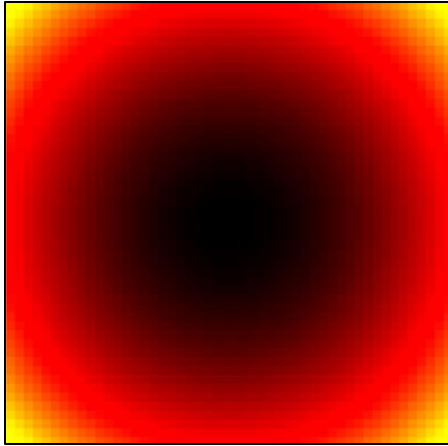


Fair benchmarking of known and **new methods**



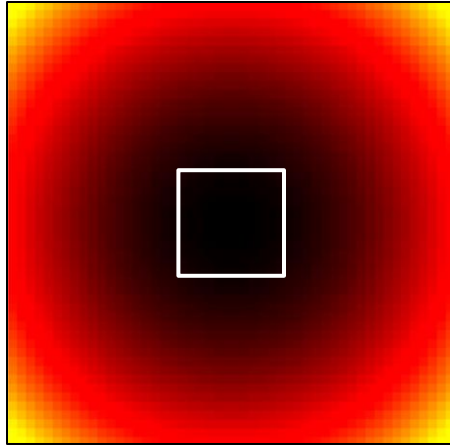
Indefinite zooming-in with zeroes

Parabolic minimum

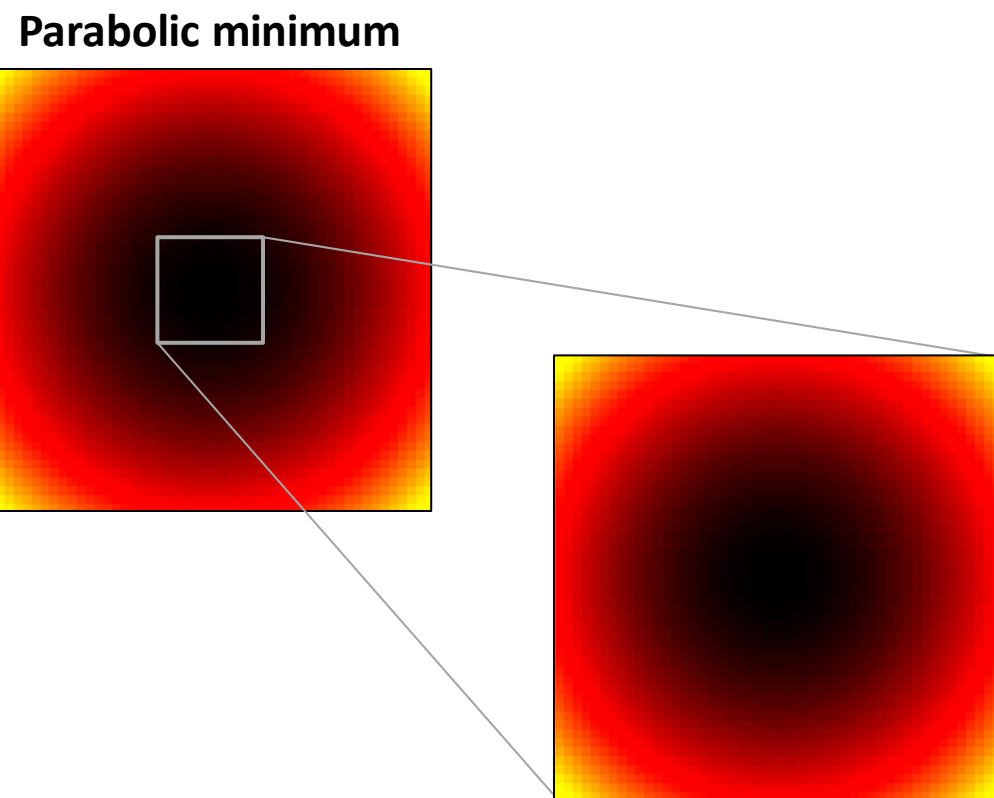


Indefinite zooming-in with zeroes

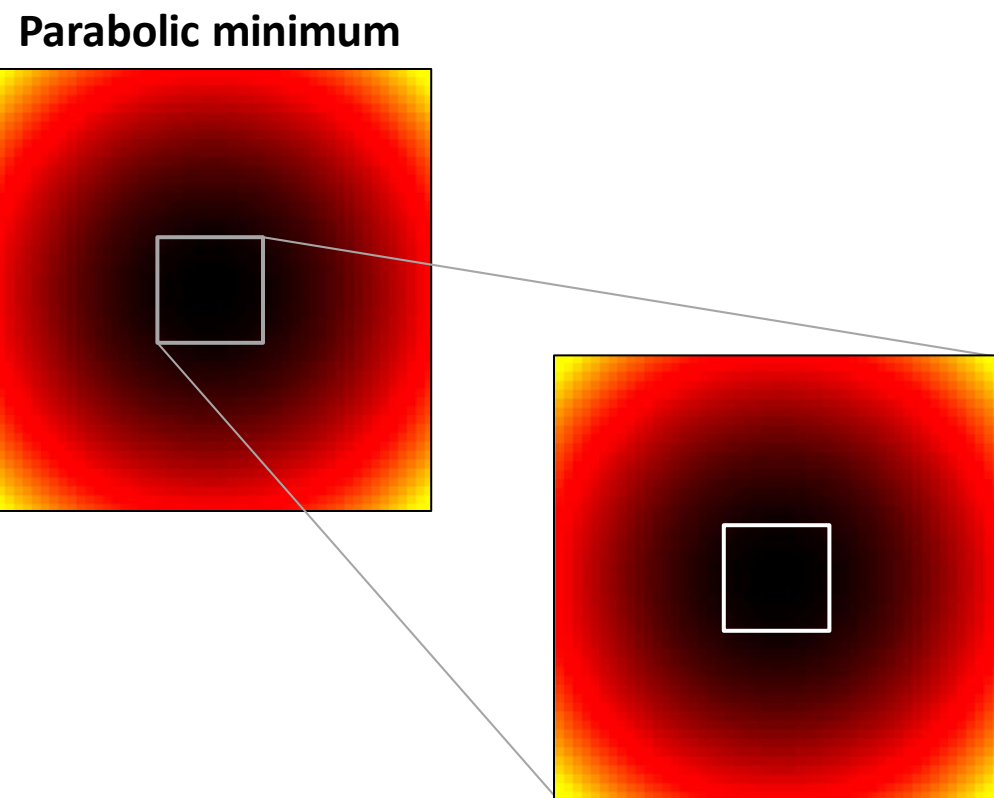
Parabolic minimum



Indefinite zooming-in with zeroes

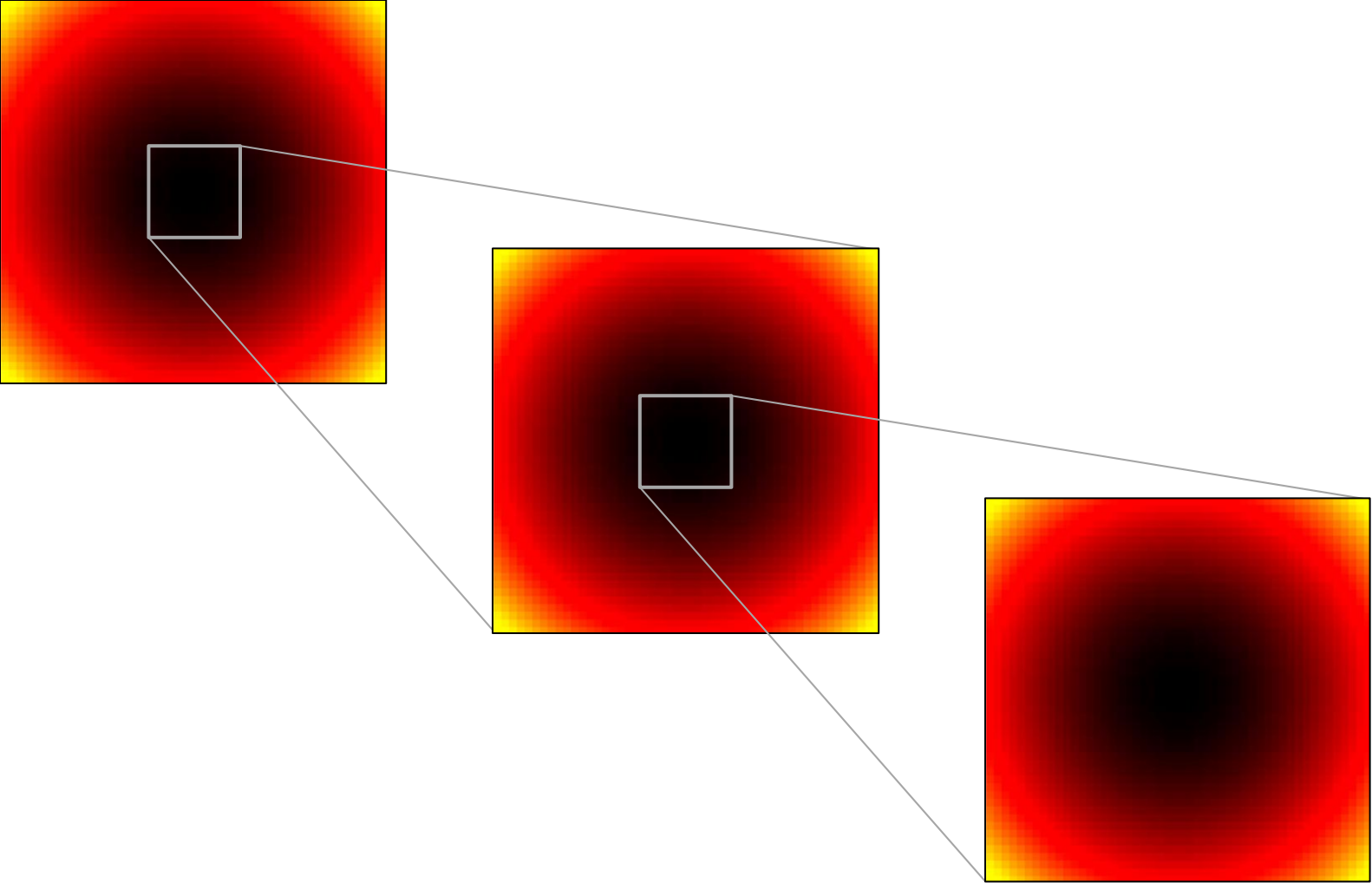


Indefinite zooming-in with zeroes



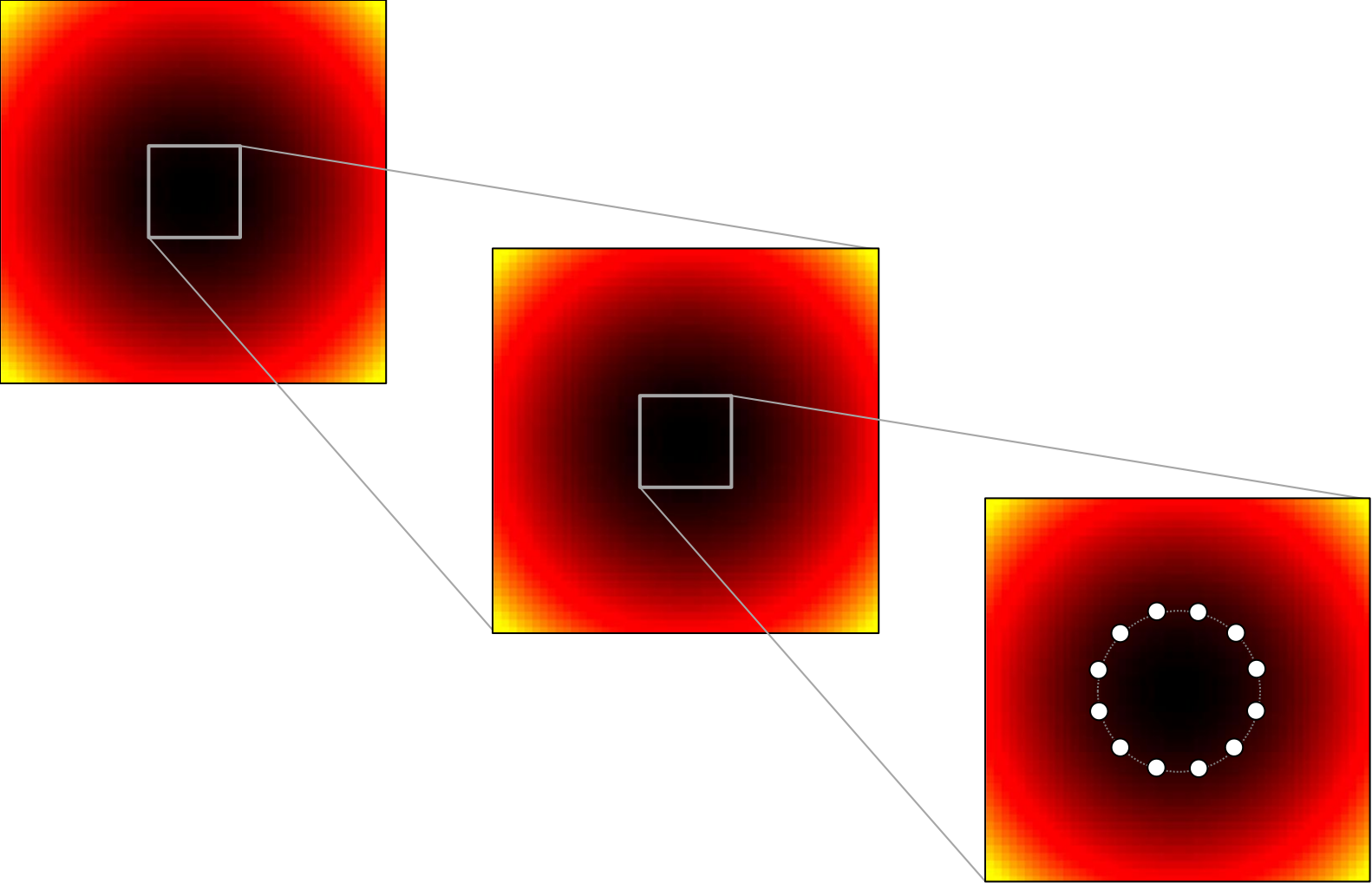
Indefinite zooming-in with zeroes

Parabolic minimum



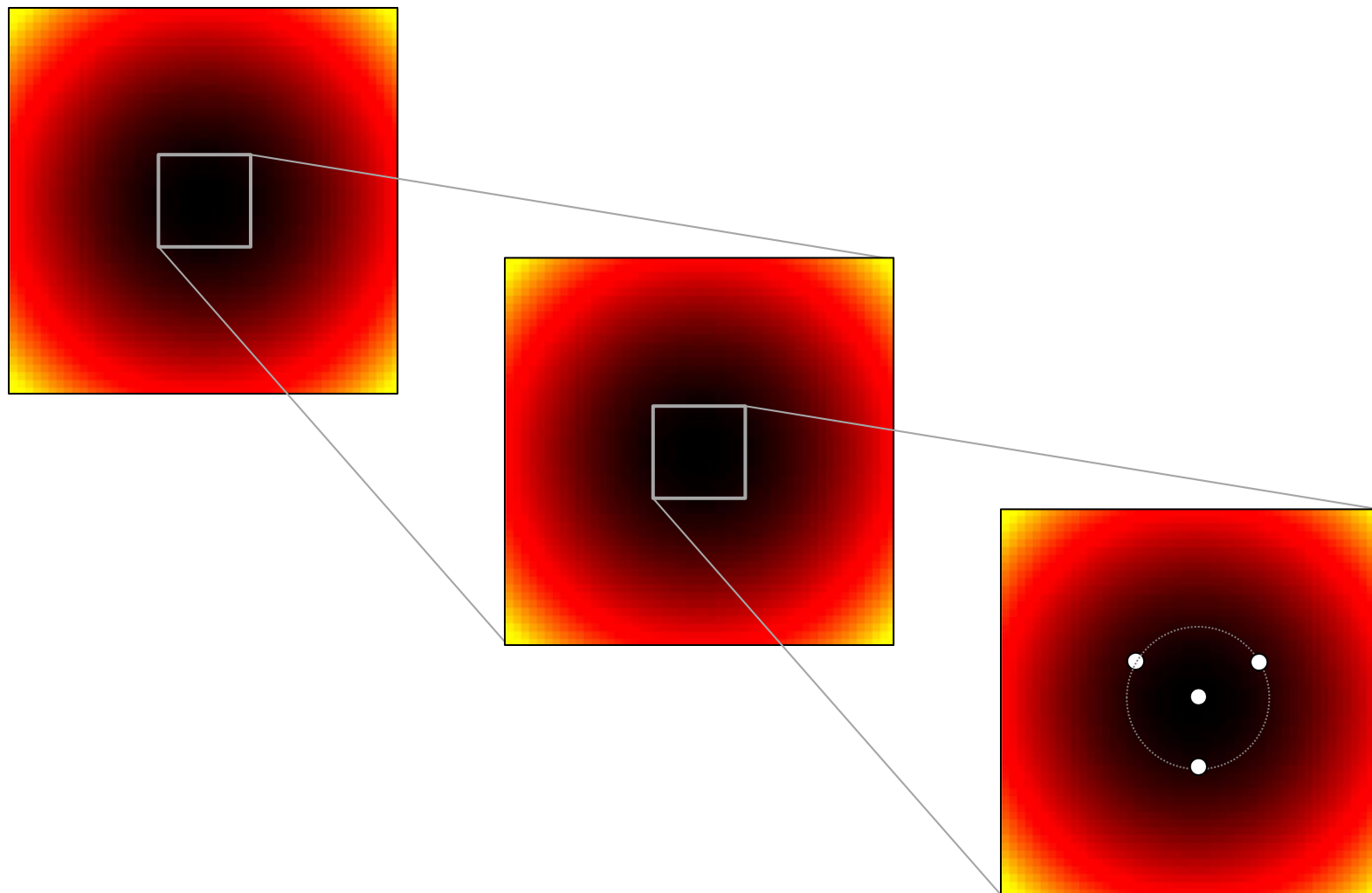
Indefinite zooming-in with zeroes

Parabolic minimum

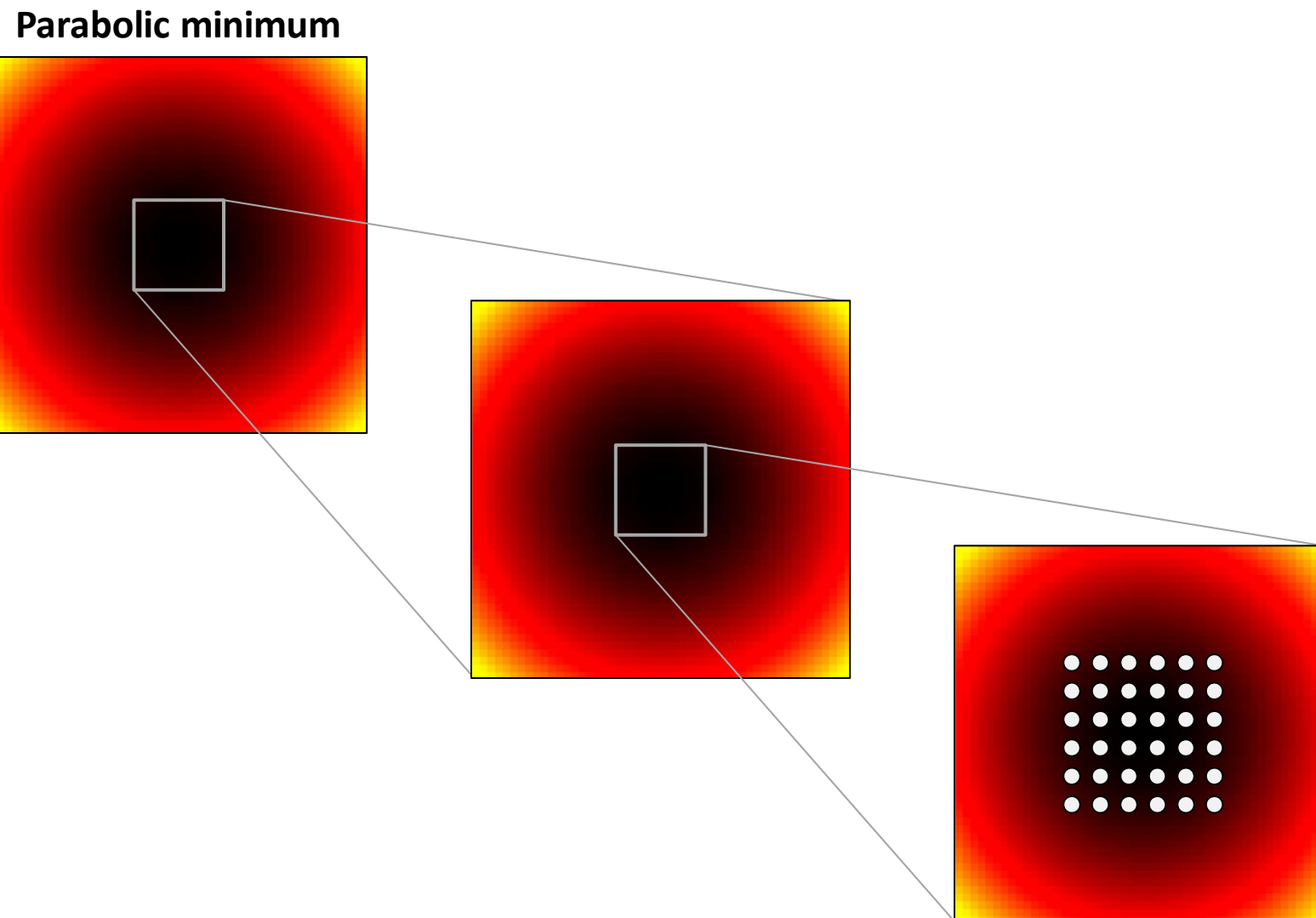


Indefinite zooming-in with zeroes

Parabolic minimum



Indefinite zooming-in with zeroes

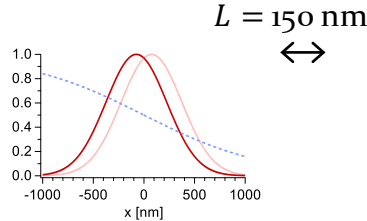
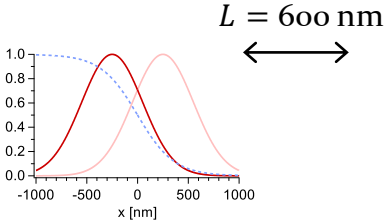
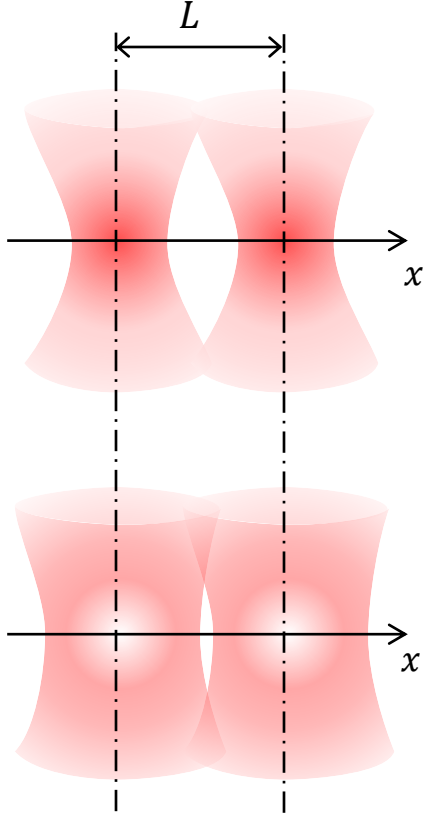


Indefinite zooming-in with zeroes

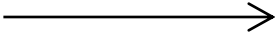
— $I_1(x) = I(x + L/2)$

— $I_2(x) = I(x - L/2)$

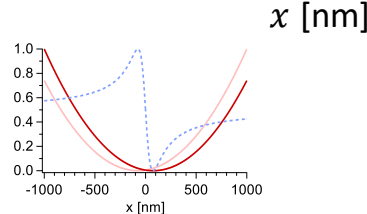
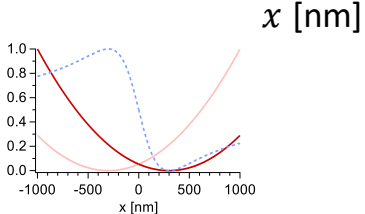
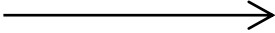
... $p(x) = \frac{I_2(x)}{I_1(x) + I_2(x)}$



Zooming-in
with maxima



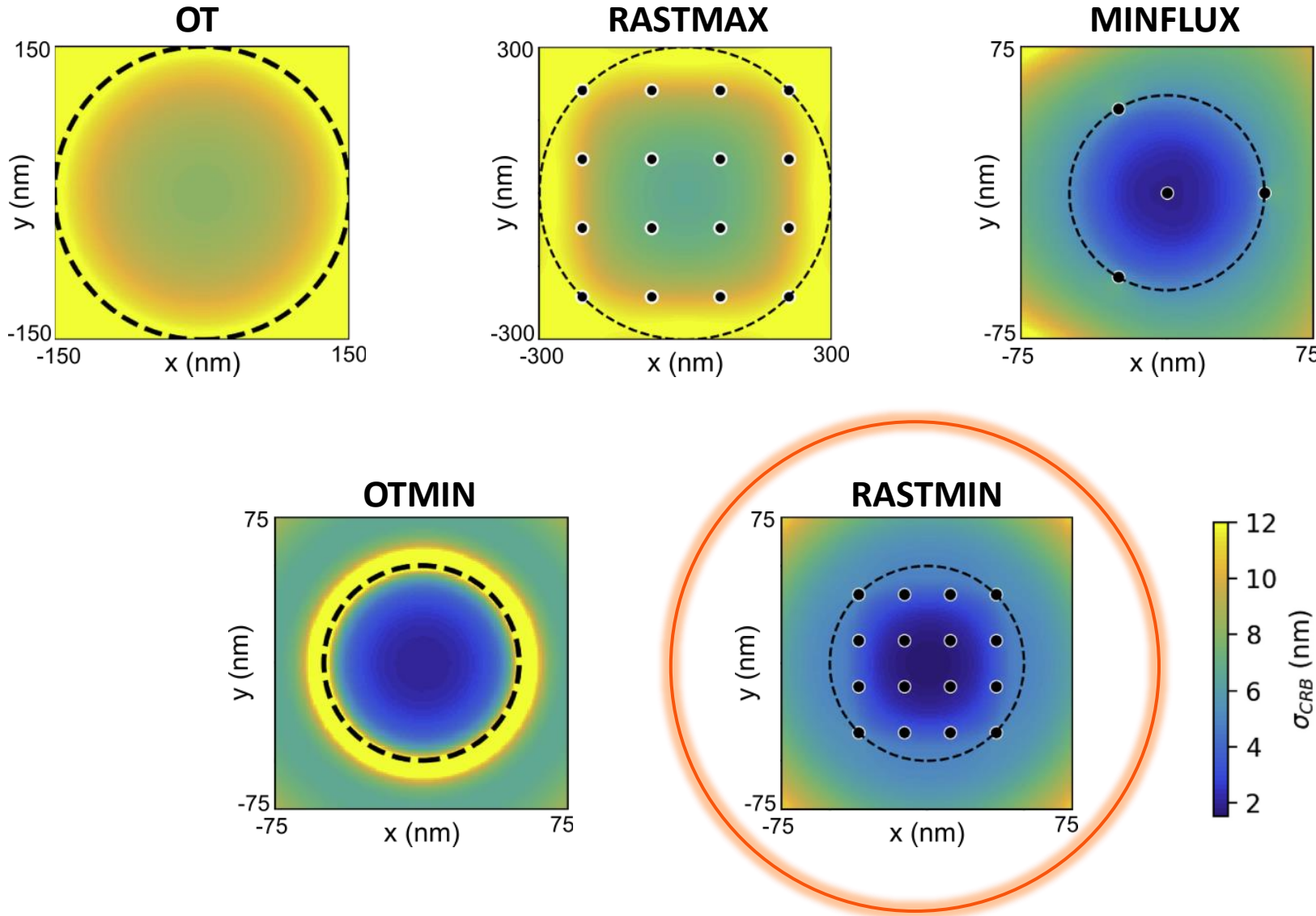
Zooming-in
with minima



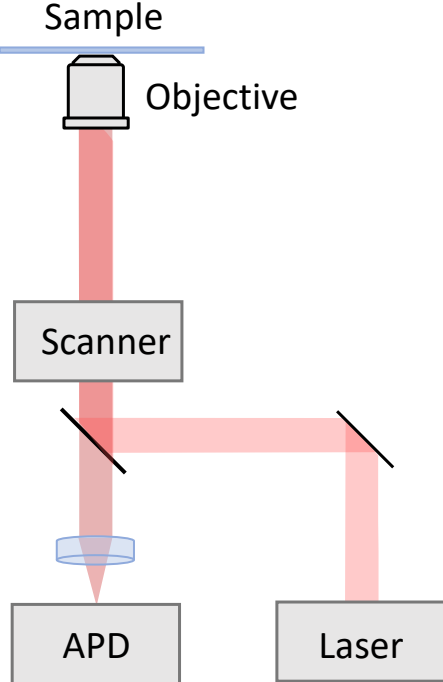
$x \text{ [nm]}$

$x \text{ [nm]}$

Fair benchmarking of known and new methods

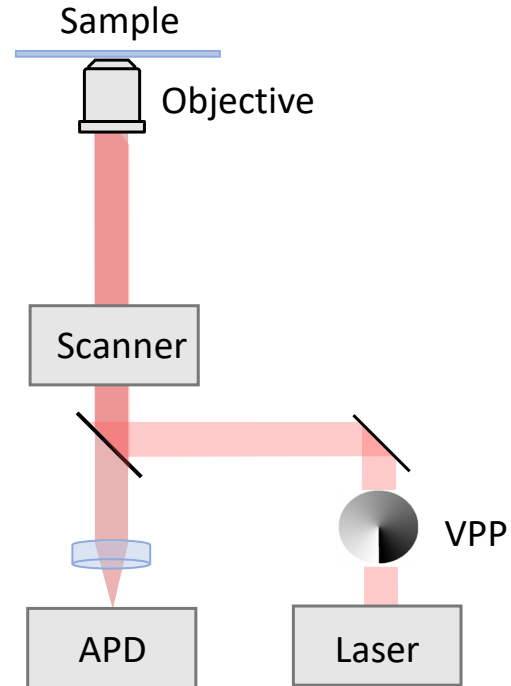


RASTMIN instrumentation



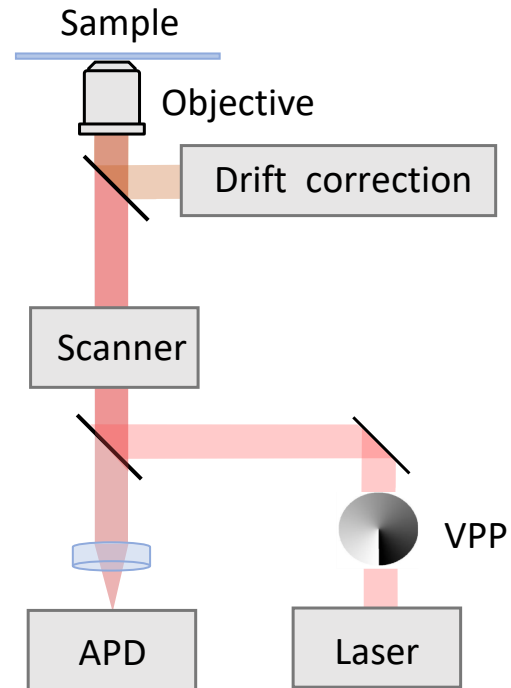
Standard scanning microscope
(confocal, two-photon)

RASTMIN instrumentation



Standard scanning microscope
(confocal, two-photon)
+
Vortex Phase Plate (or SLM)

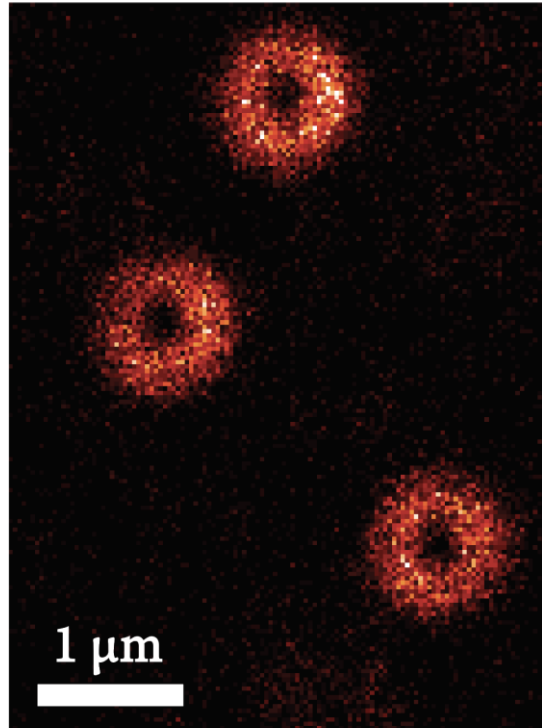
RASTMIN instrumentation



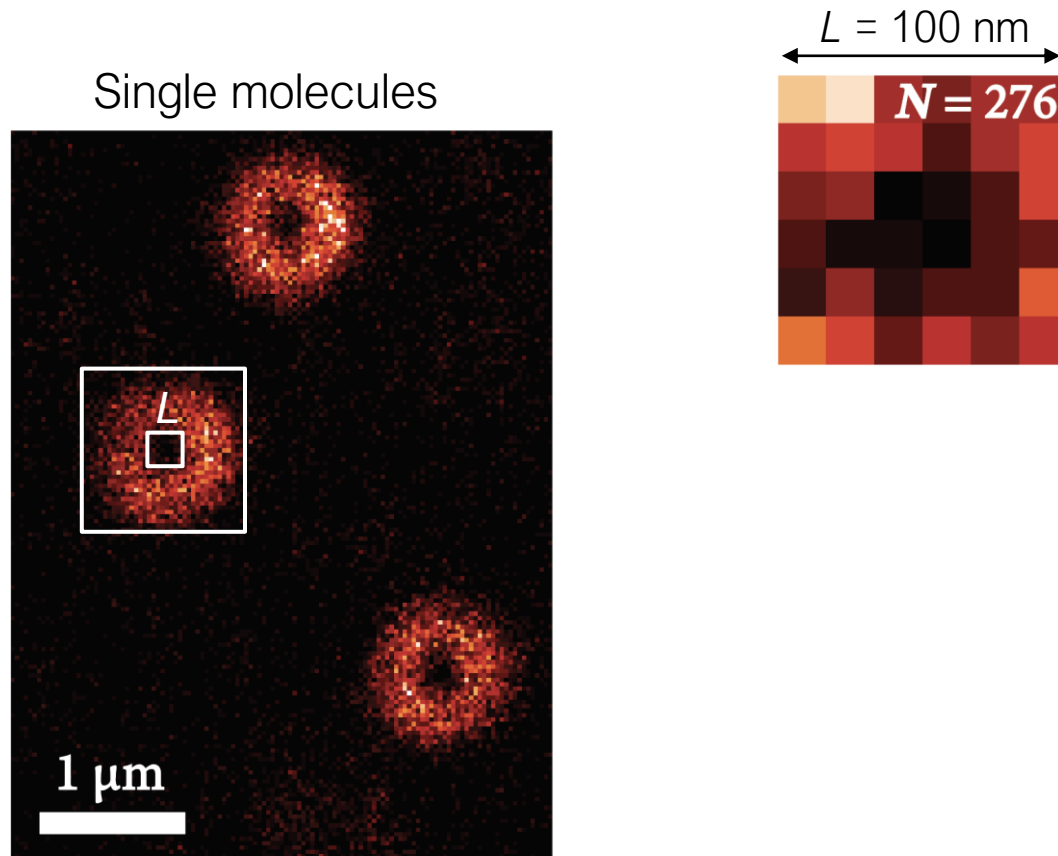
Standard scanning microscope
(confocal, two-photon)
+
Vortex Phase Plate (or SLM)
+
Active drift correction

Single-molecule localization with RASTMIN

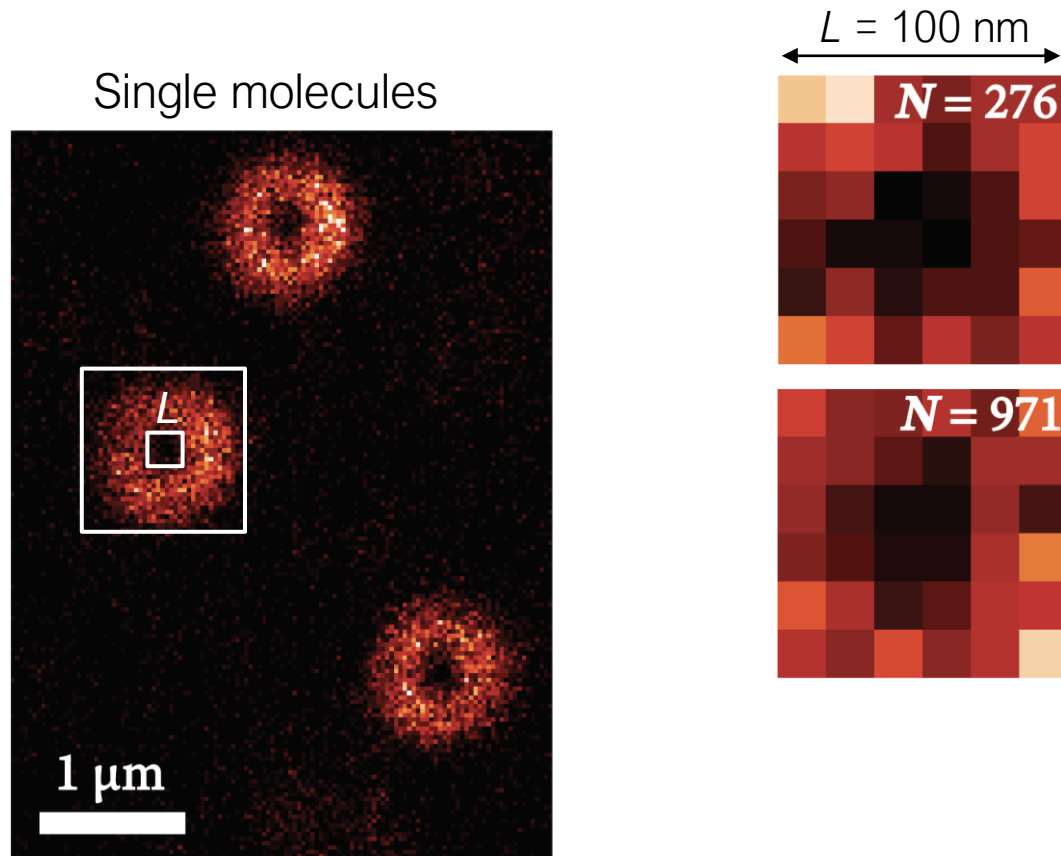
Single molecules



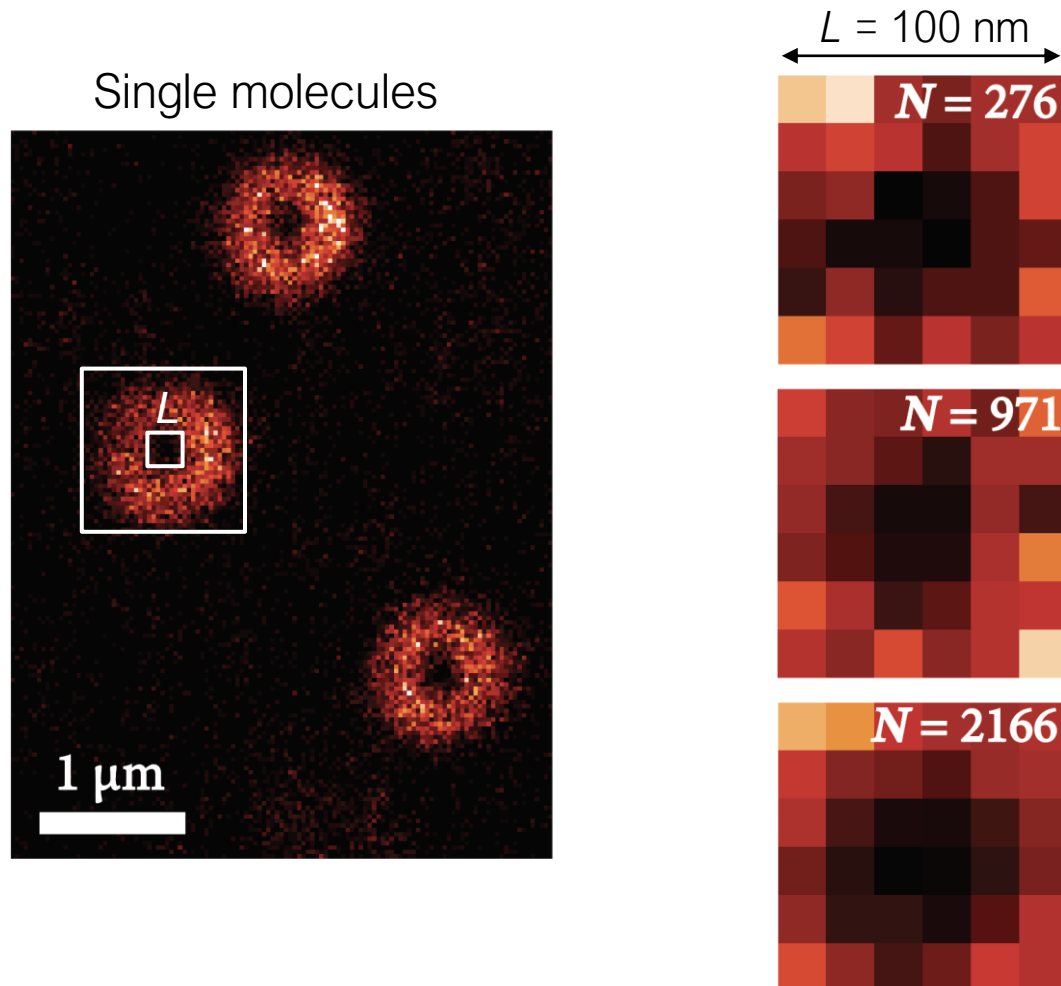
Single-molecule localization with RASTMIN



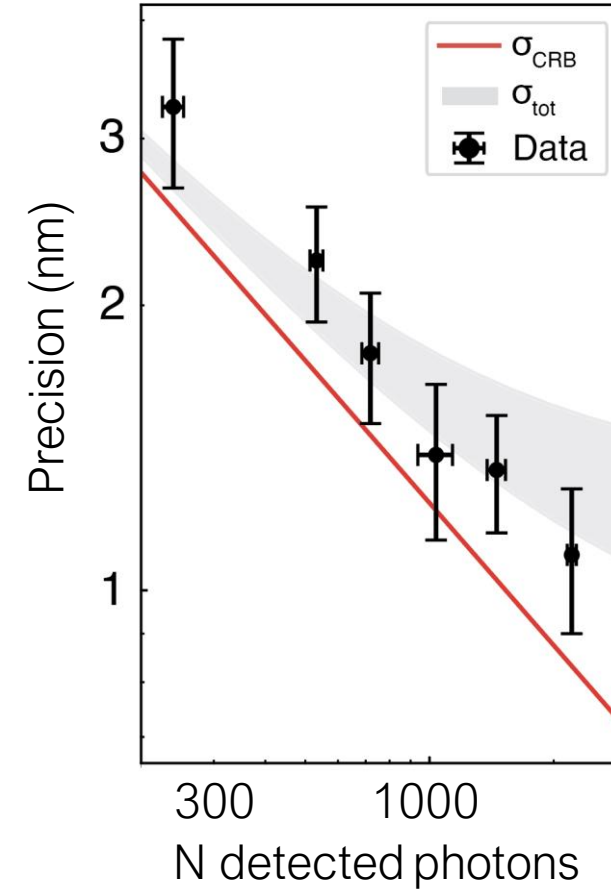
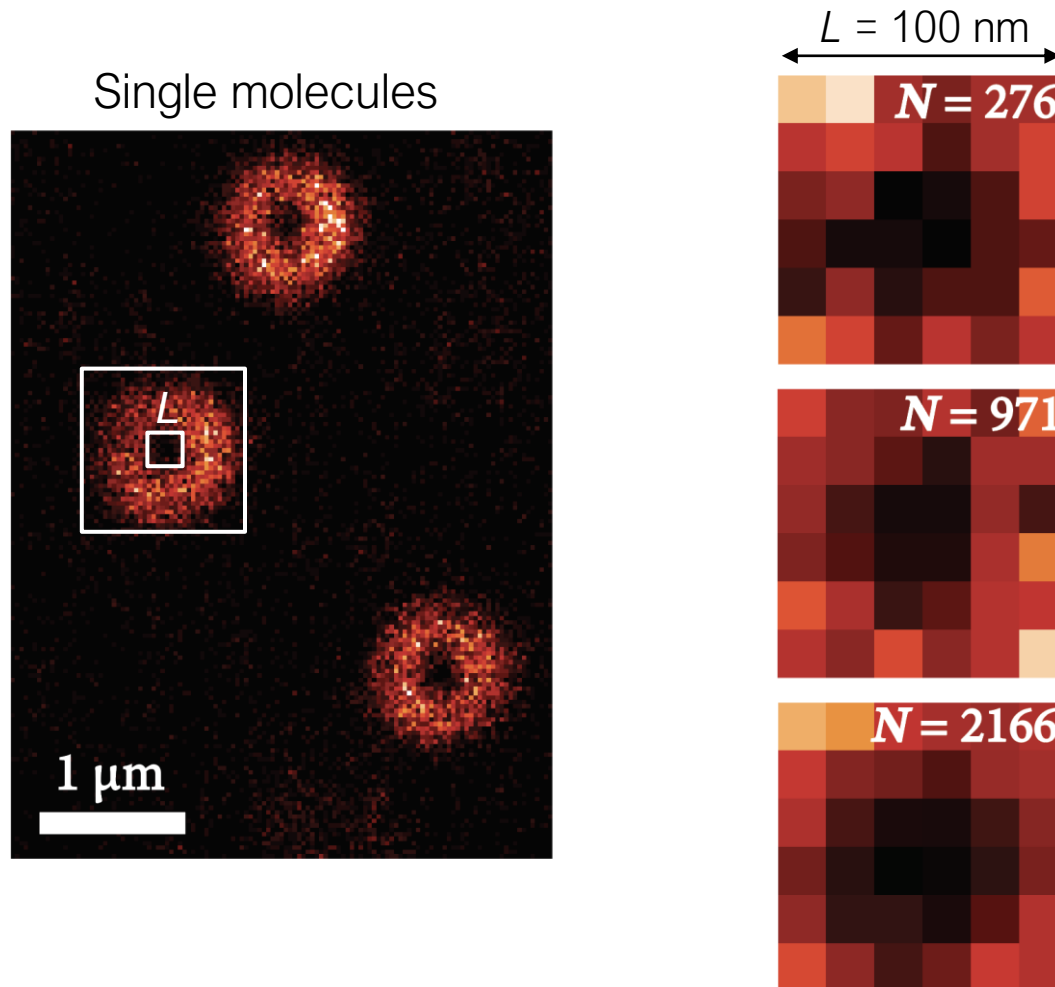
Single-molecule localization with RASTMIN



Single-molecule localization with RASTMIN

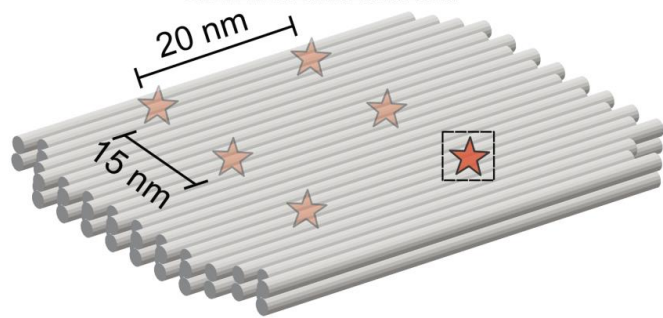
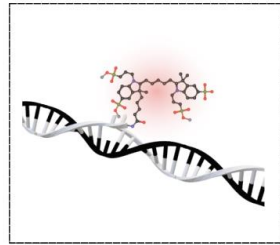


Single-molecule localization with RASTMIN

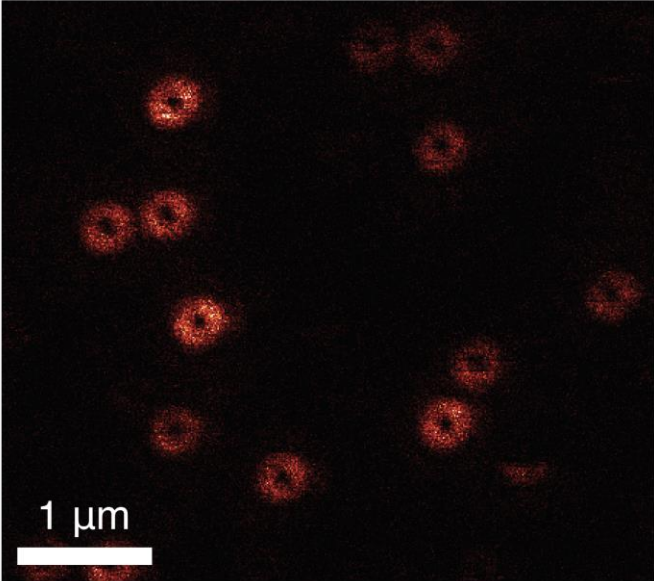
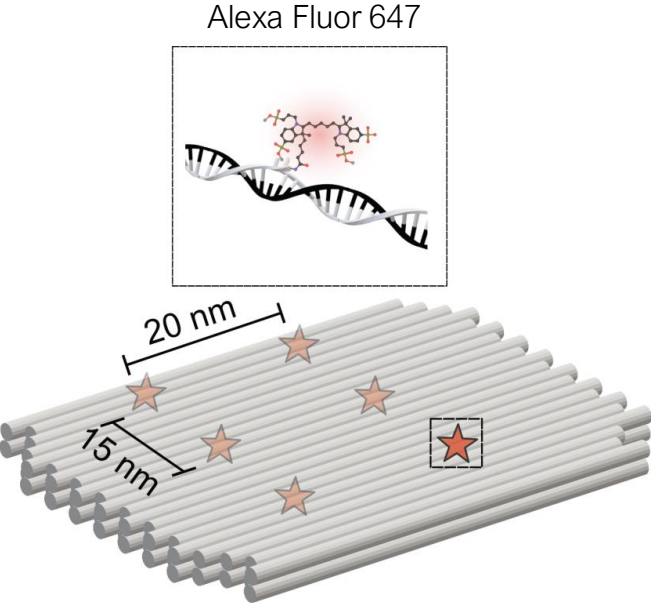


RASTMIN nanoscopy

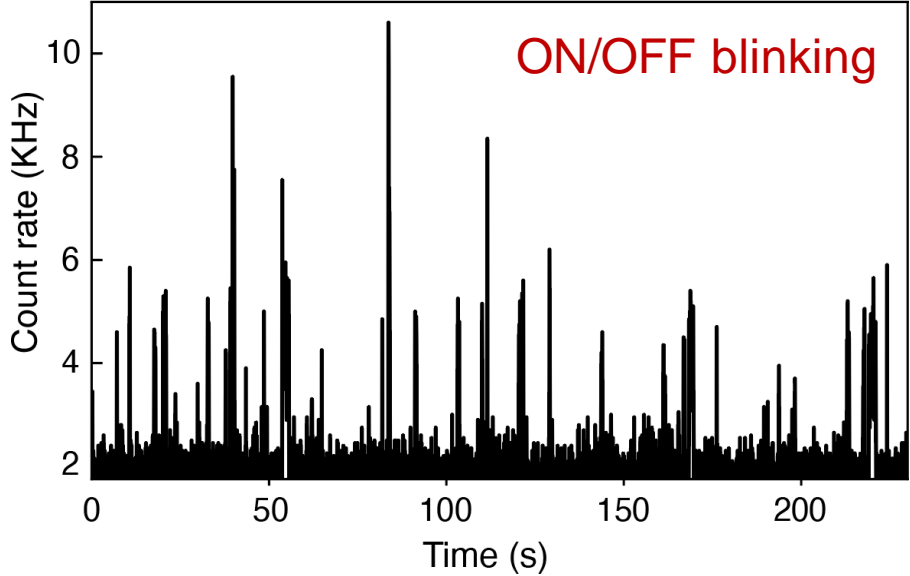
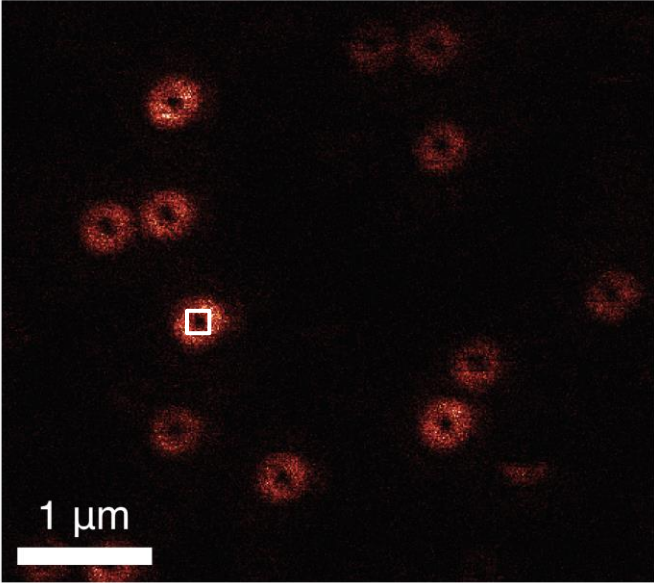
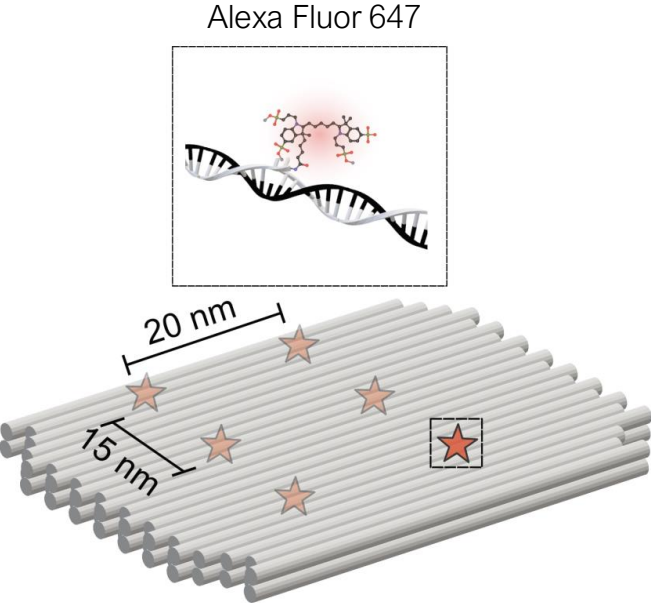
Alexa Fluor 647



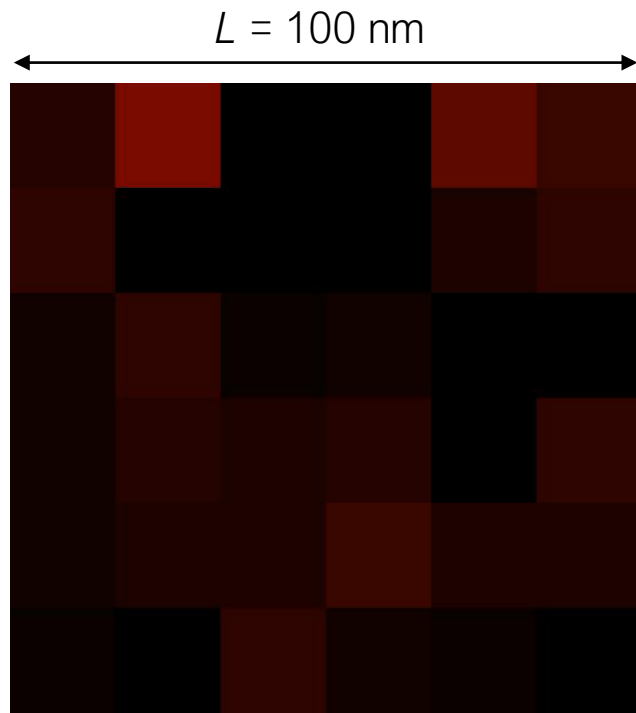
RASTMIN nanoscopy



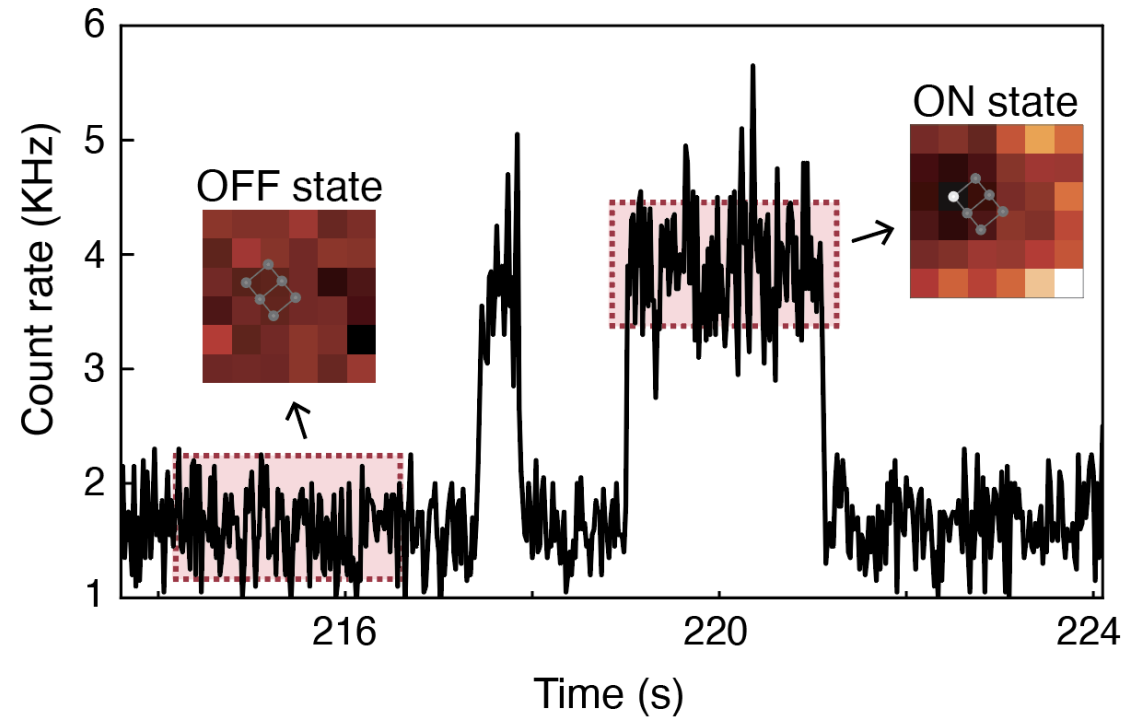
RASTMIN nanoscopy



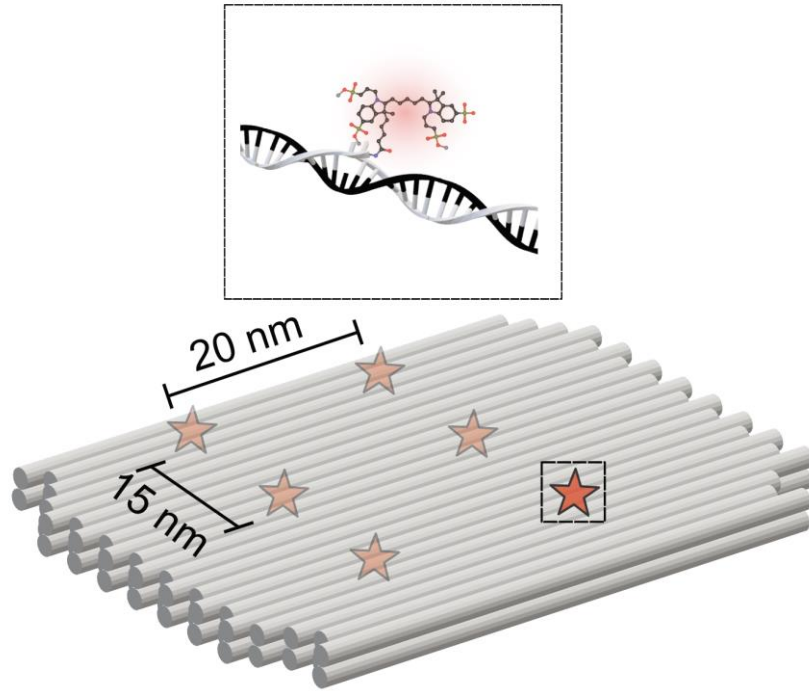
RASTMIN nanoscopy



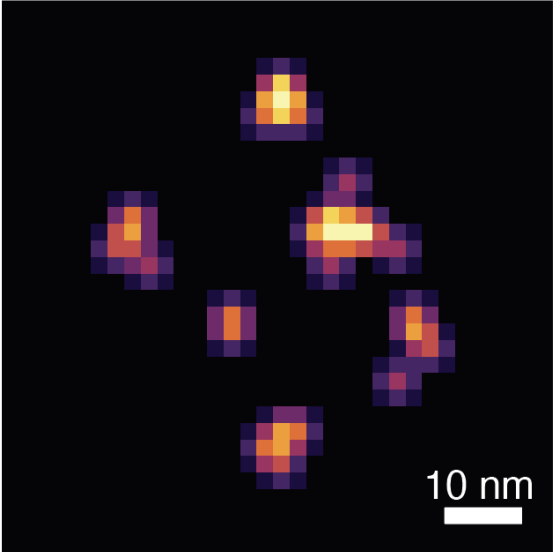
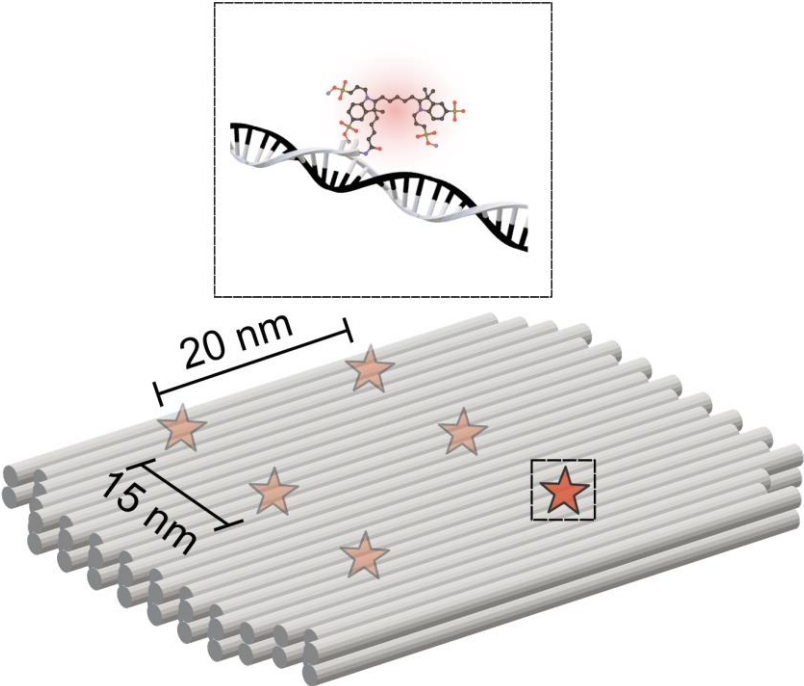
Confocal time-lapse (raw data)



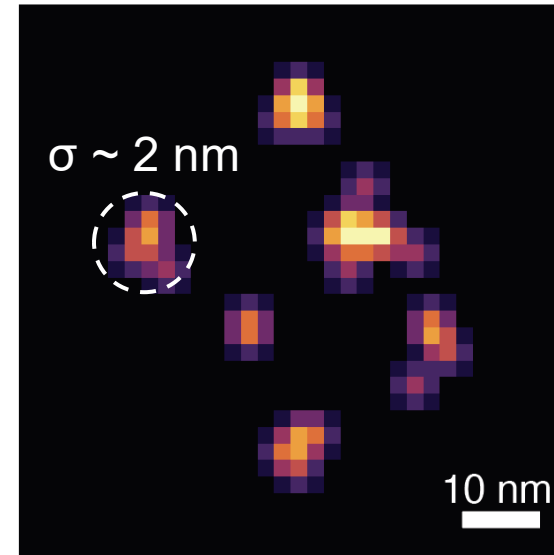
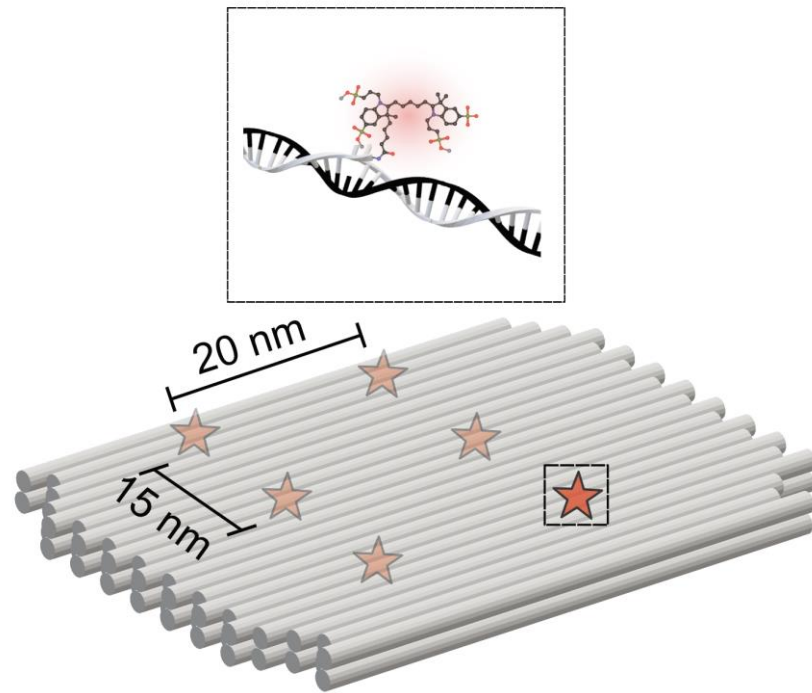
RASTMIN nanoscopy



RASTMIN nanoscopy



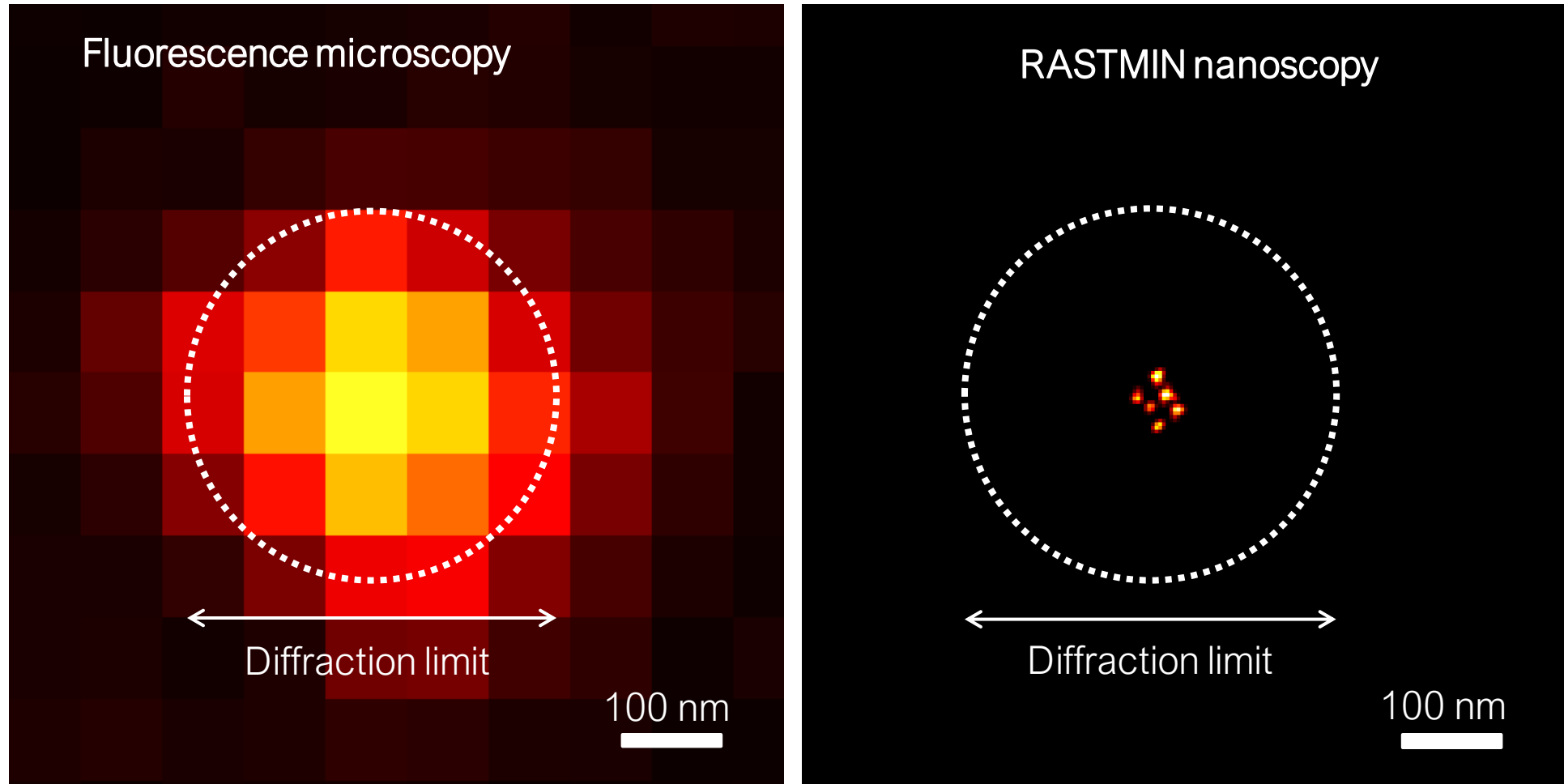
RASTMIN nanoscopy



Masullo et al, *Light: Science and Applications* 11, 199 (2022)

Masullo et al, *Light: Science and Applications* 11, 70 (2022)

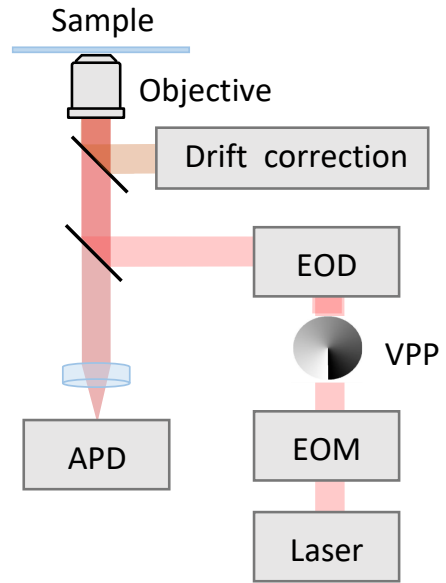
RASTMIN nanoscopy



Masullo et al, *Light: Science and Applications* 11, 199 (2022)

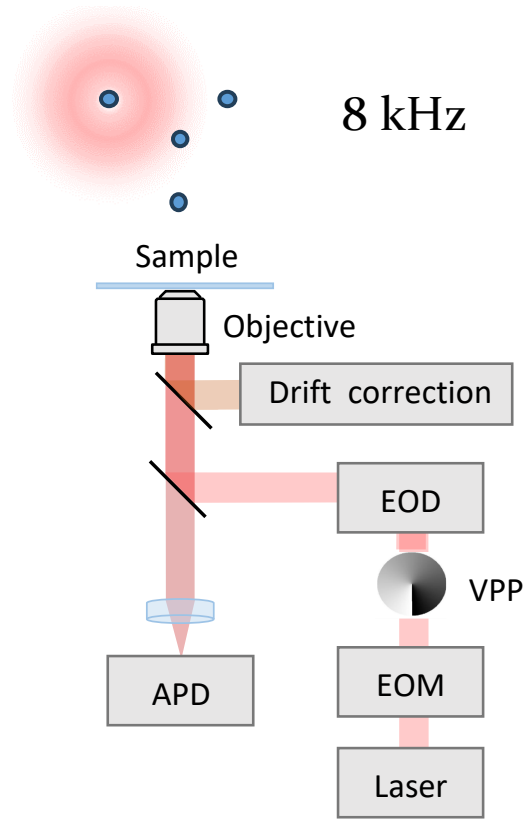
Masullo et al, *Light: Science and Applications* 11, 70 (2022)

MINFLUX instrumentation



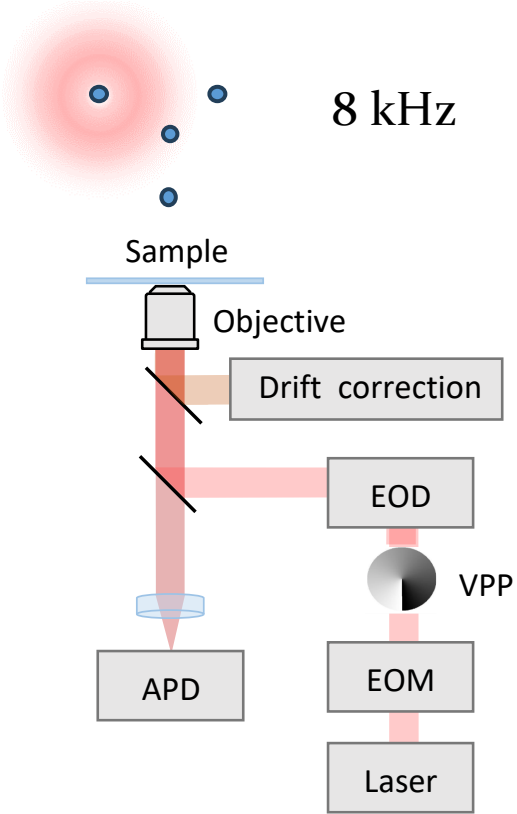
Balzarotti et al. *Science* 355 (2017) 606-612

MINFLUX instrumentation

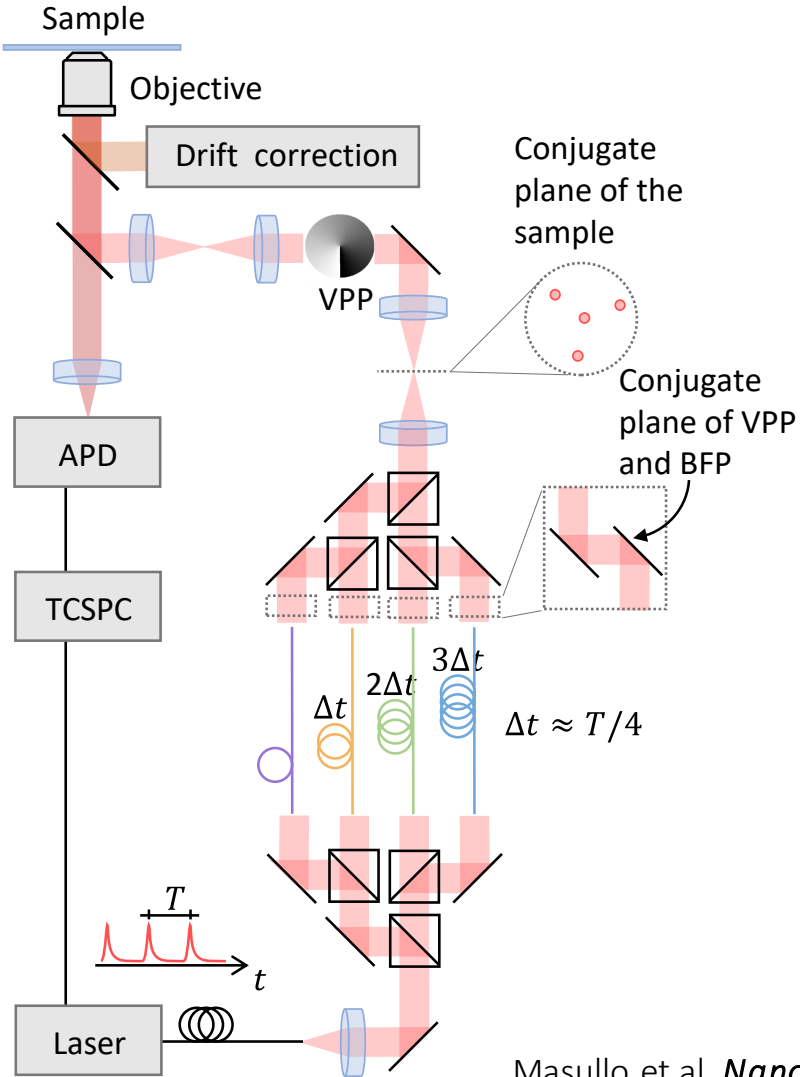


Balzarotti et al. *Science* 355 (2017) 606-612

MINFLUX instrumentation

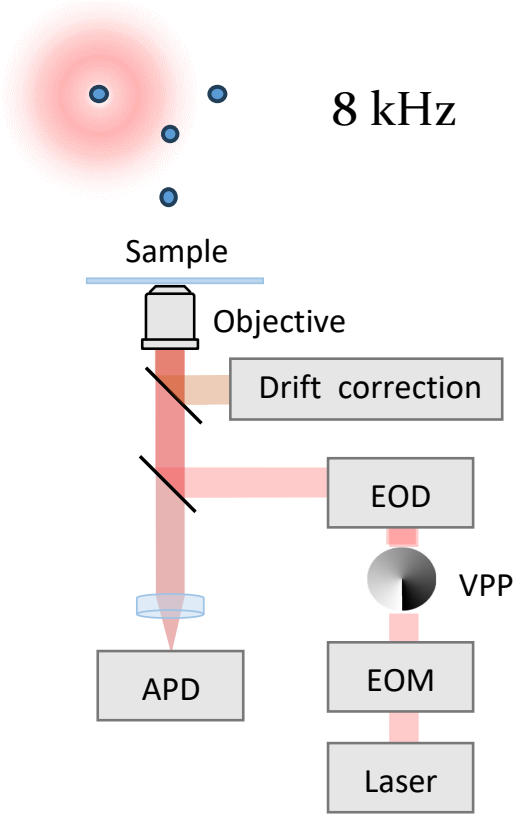


Balzarotti et al. *Science* 355 (2017) 606-612

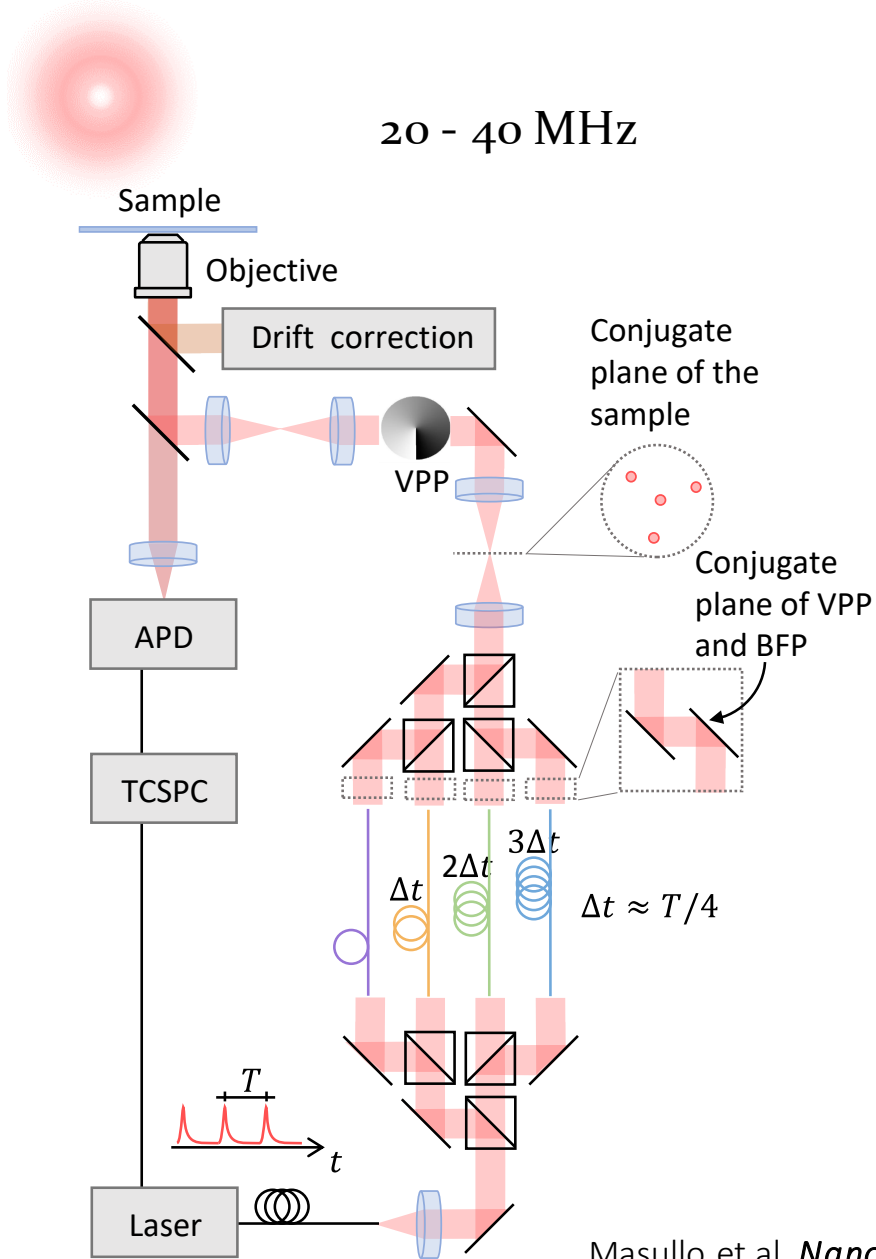


Masullo et al. *Nano Letters* 21 (2021) 840-846

MINFLUX instrumentation

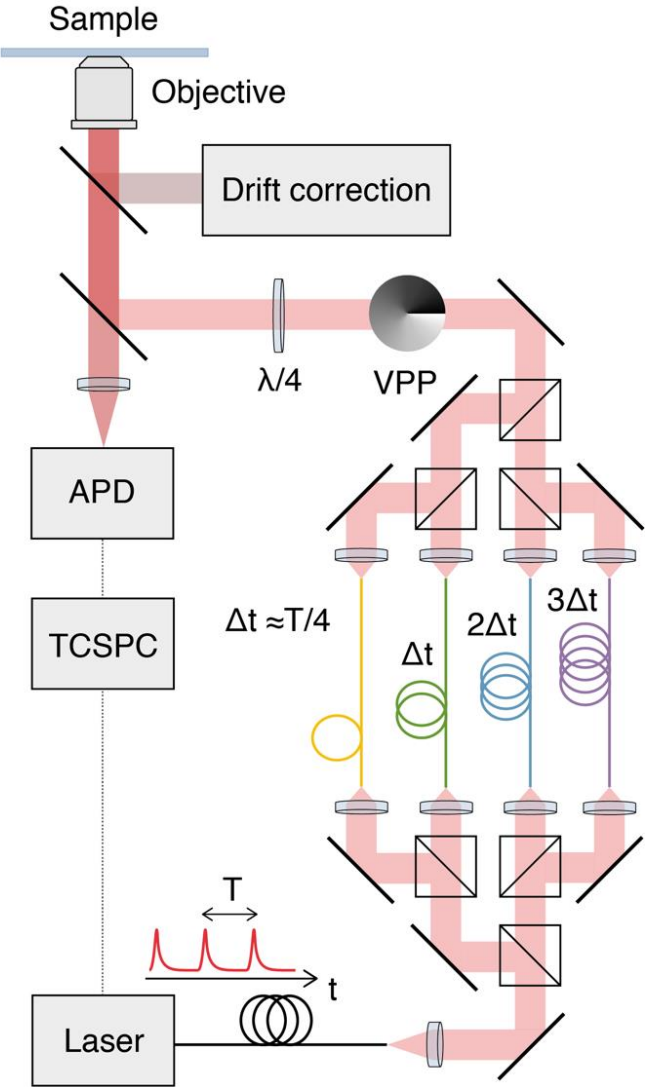
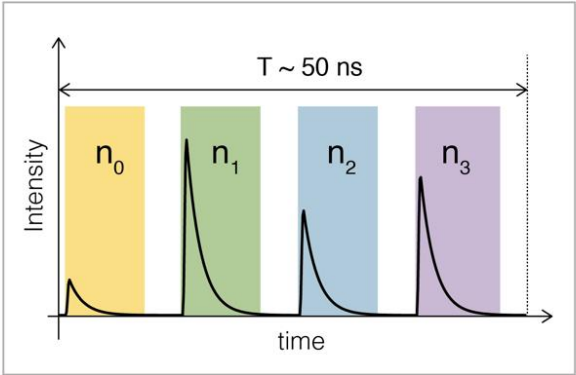
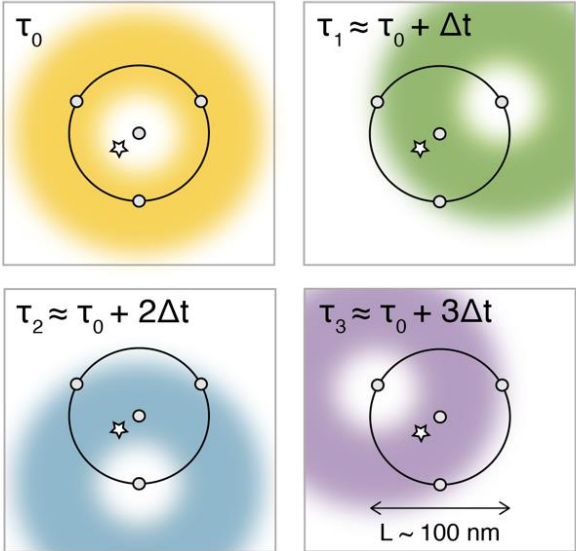


Balzarotti et al. *Science* 355 (2017) 606-612

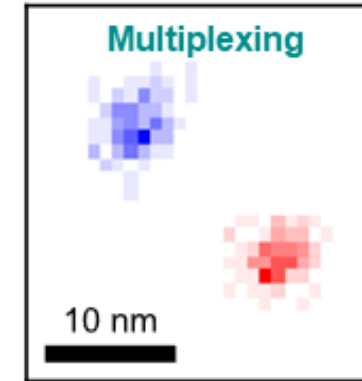
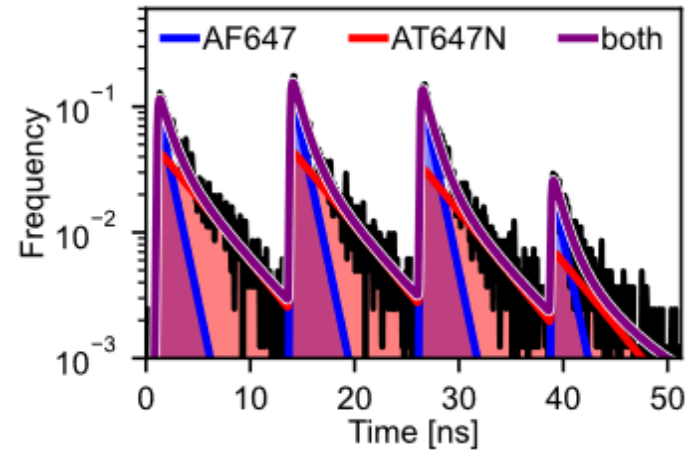
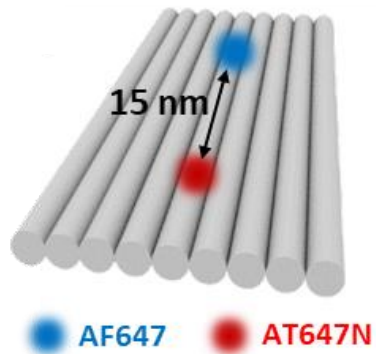


Masullo et al. *Nano Letters* 21 (2021) 840-846

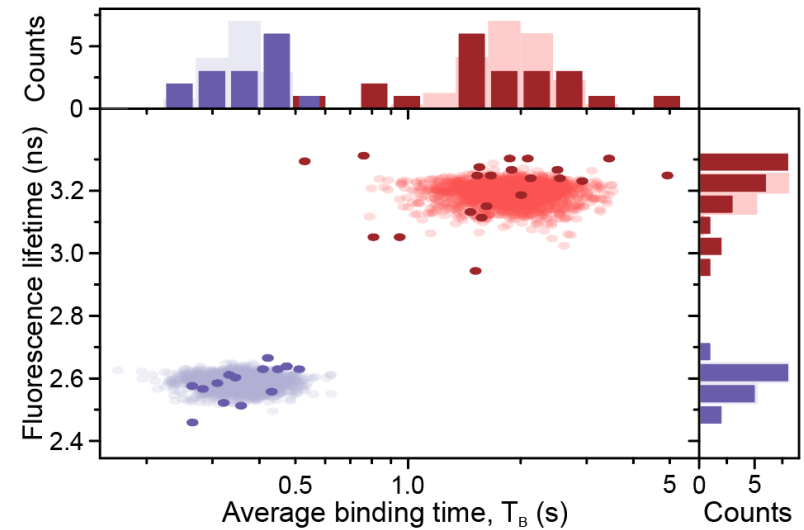
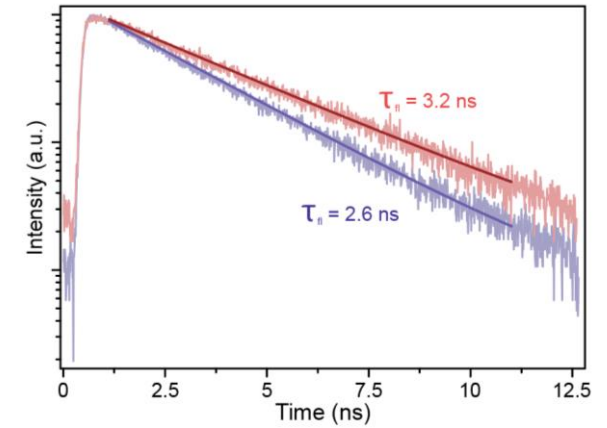
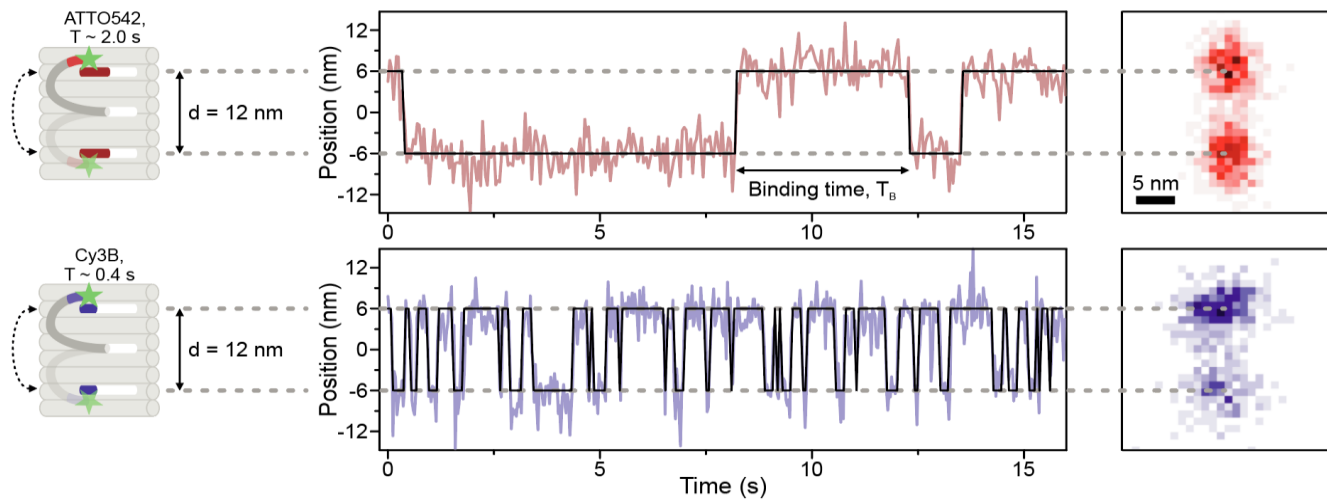
Pulsed-interleaved MINIFLUX (p-MINIFLUX)



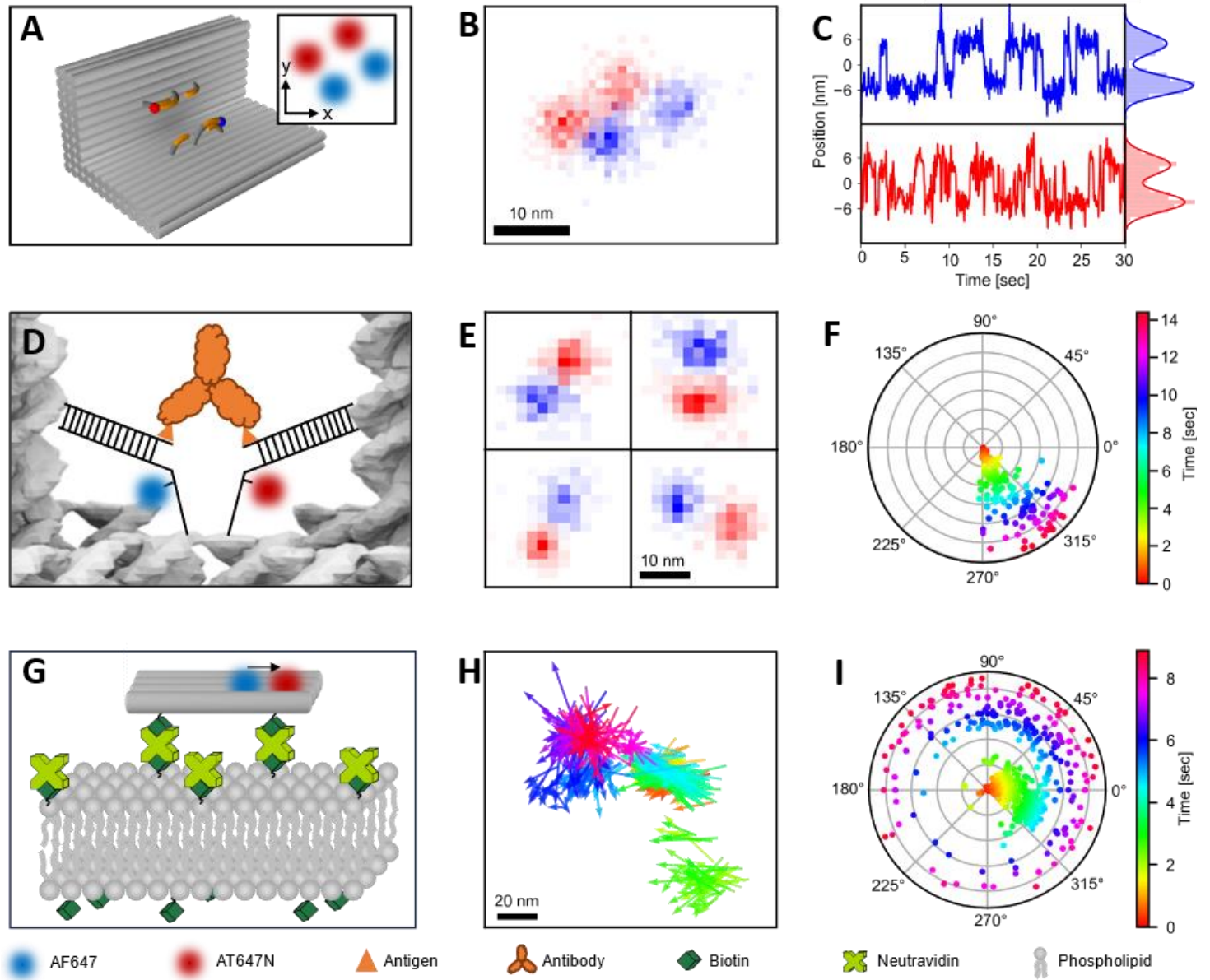
Lifetime multiplexed localization with p-MINFLUX



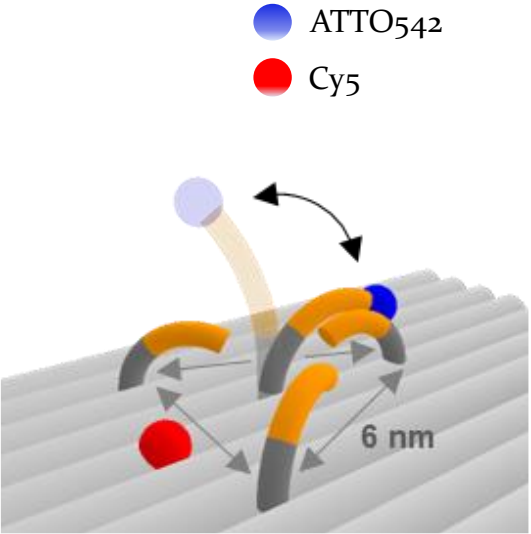
Lifetime multiplexed single-molecule tracking with p-MINFLUX



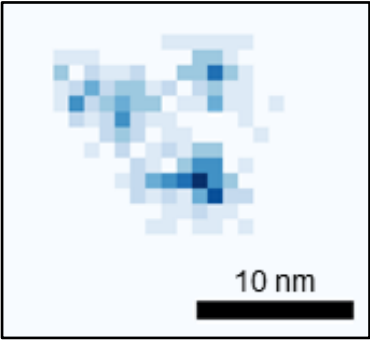
Lifetime multiplexed single-molecule tracking with p-MINFLUX



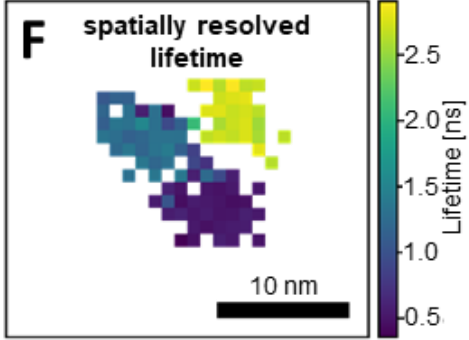
Localizing a dark absorber with p-MINFLUX



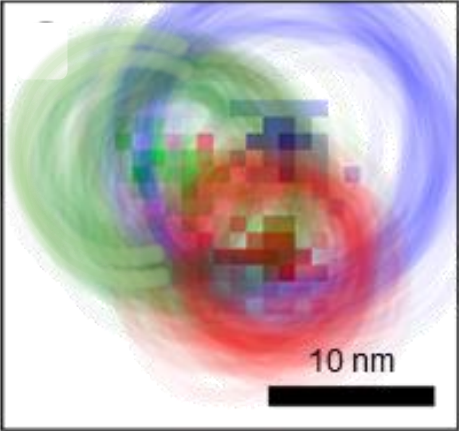
Localizations



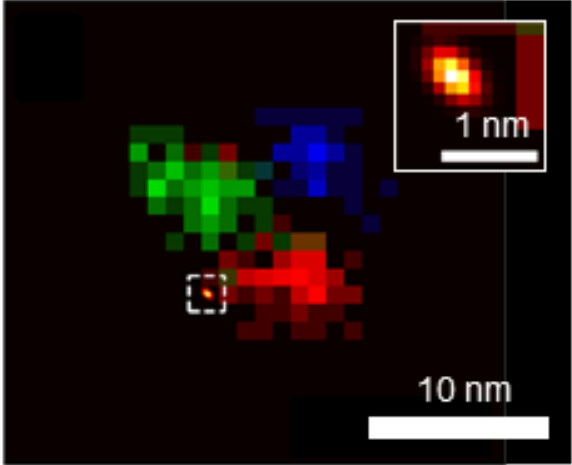
Lifetime



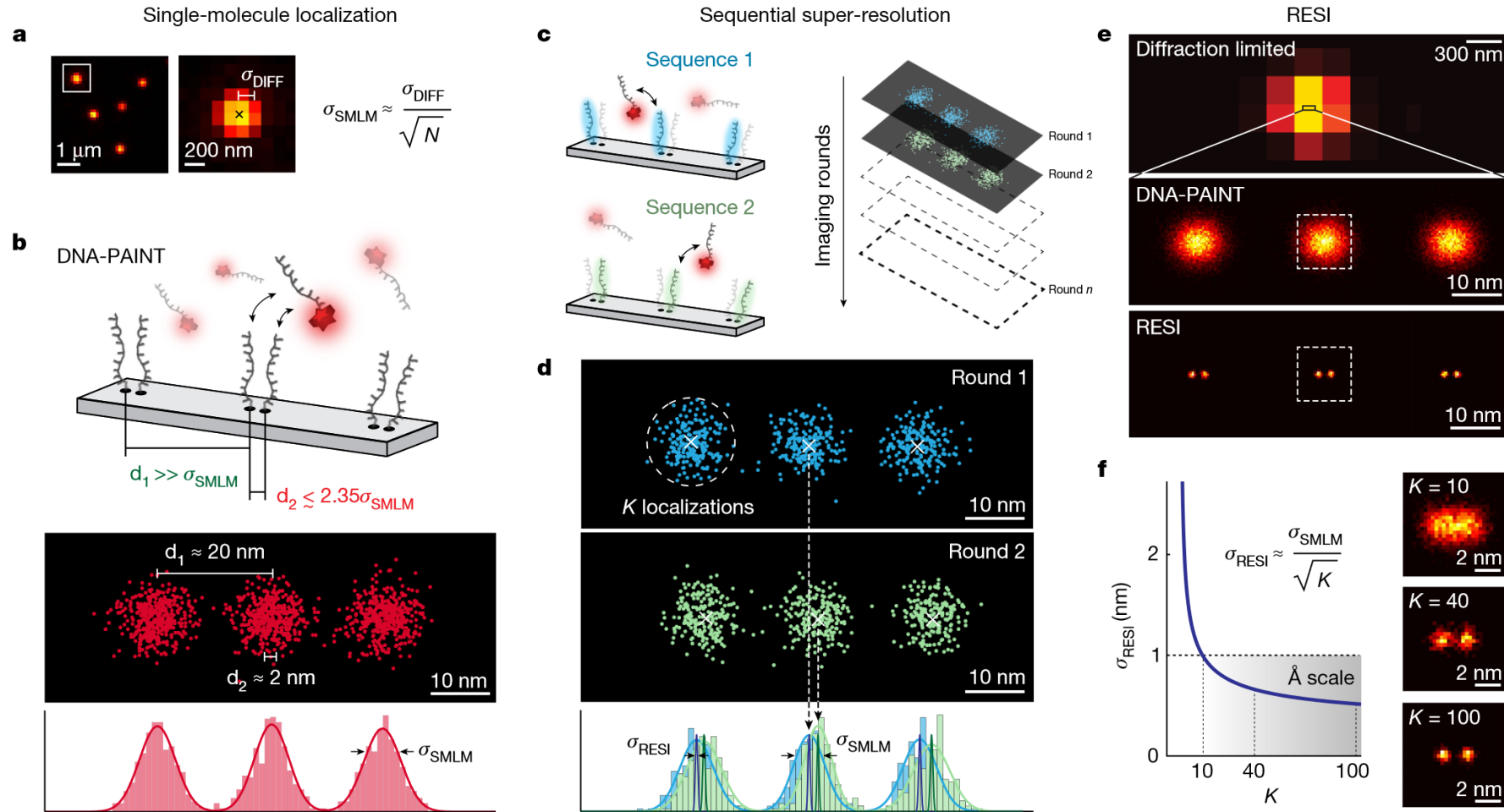
Acceptor triangulation



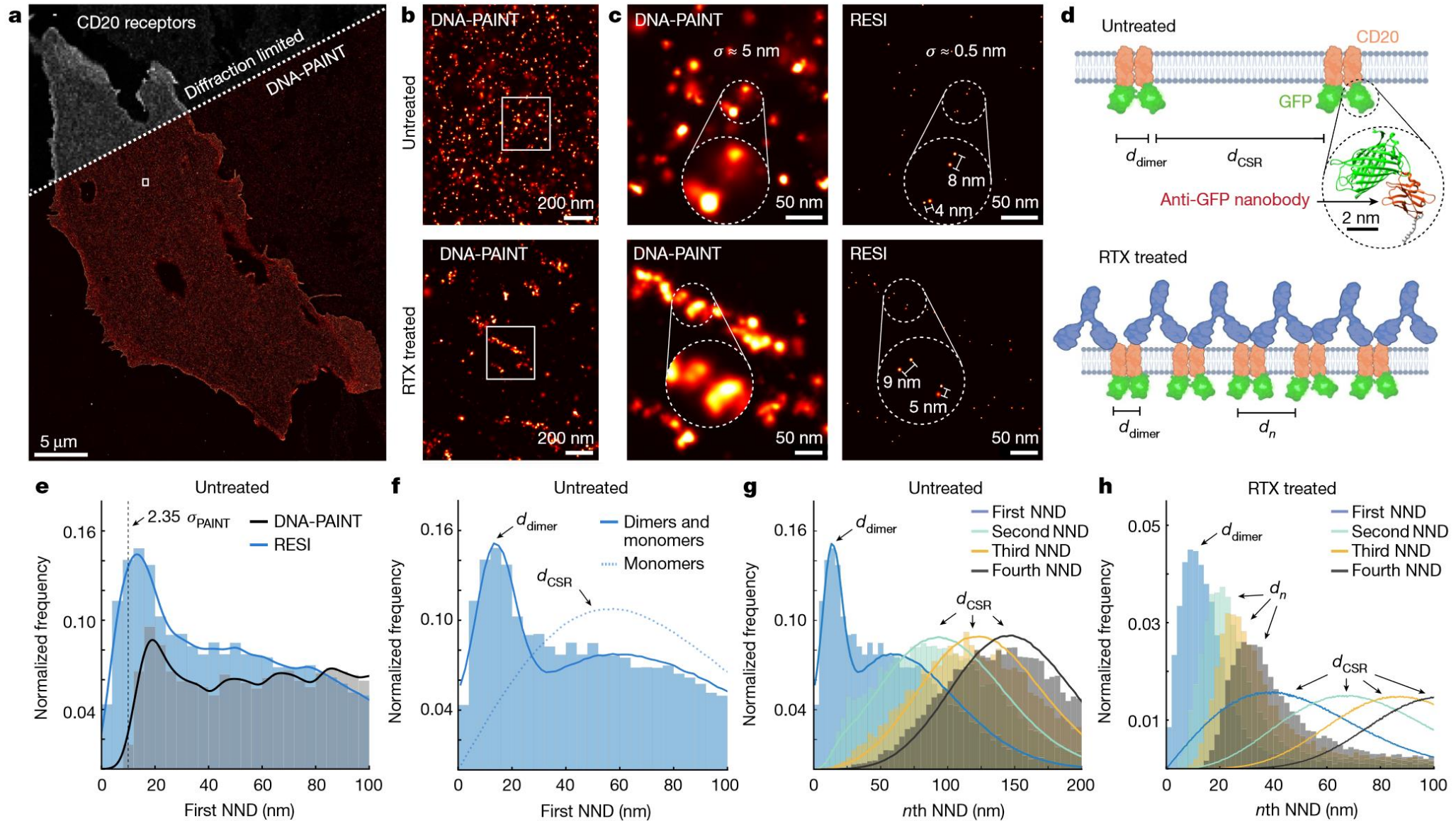
Acceptor localization



Resolution Enhancement by Sequential Imaging (RESI)



Resolution Enhancement by Sequential Imaging (RESI)

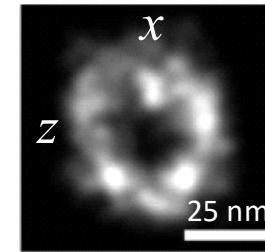
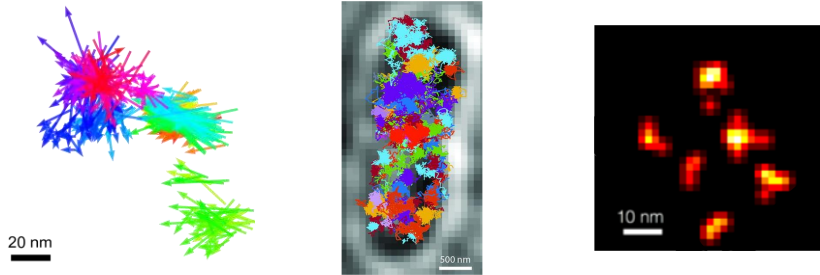


Sub-10 nm resolution fluorescence nanoscopy

<https://stefani-lab.ar/>

fernando.stefani@df.uba.ar

SML-SSI



SIMPLER

Sub-10 nm 3D TIRF nanoscopy

p-MINFLUX: fast / accurate tracking

RASTMIN / MINFLUX:

the high photon-efficiency is losing significance for highest resolution: tricks to get more photons, label size limit,...

Another edge: 10-20 nm resolution with “bad” fluorophores

Combination with camera-based localization for photophysical studies (absorption-emission)

STED-FRET



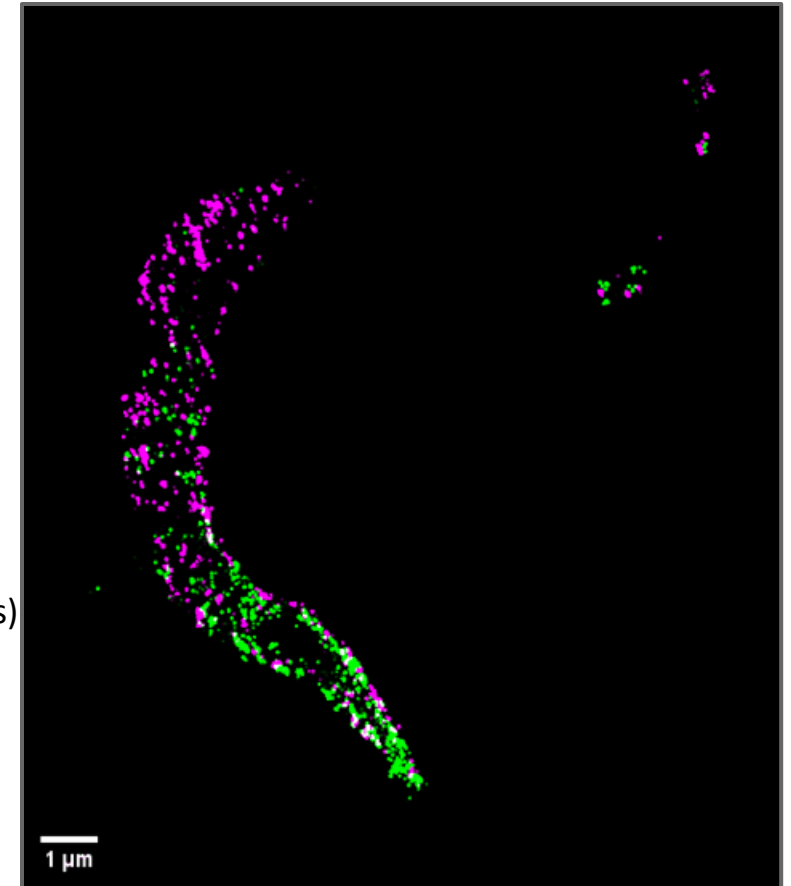
Super-resolved energy transfer / biomolecular interaction



<https://stefani-lab.ar/>

fernando.stefani@df.uba.ar

Current projects: applications to biomedicine



Luciano Masullo (now at MPI biochem, Munich)

Cecilia Zaza (now at UC London)

Romina Landa (now at Collective.ai, Buenos Aires)

Fernando Caprile (now at Iquall Networks, Buenos Aires)

German Chiarelli (now at Fribourg University))

Julián Gargiulo (now at UNSAM, Buenos Aires)

Ianina Violi (now at UNSAM, Buenos Aires)

Valeria País (now at ICFO, Barcelona)

Emiliano Cortés (now Prof. at LMU, Munich)

Martín Bordenave (now at Satellogic, Buenos Aires)

Federico Barabas (now at Spotify, Stockholm)

