
Somatic sex determination*

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Abstract

***C. elegans* occurs in two natural sexes, the XX hermaphrodite and the XO male, which differ extensively in anatomy, physiology, and behavior. All somatic differences between the sexes result from the differential activity of a “global” sex determination regulatory pathway. This pathway also controls X chromosome dosage compensation, which is coordinated with sex determination by the action of the three SDC proteins. The SDC proteins control somatic and germline sex by transcriptional repression of the *her-1* gene. HER-1 is**

^{*}Edited by Barbara J. Meyer. Last revised February 8, 2005. Published February 10, 2006. This chapter should be cited as: Zarkower, D. Somatic sex determination (February 10, 2006), *WormBook*, ed. The *C. elegans* Research Community, WormBook, doi/10.1895/wormbook.1.84.1, <http://www.wormbook.org>.

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a secreted protein that controls a regulatory module consisting of a transmembrane receptor, **TRA-2**, three intracellular FEM proteins, and the zinc finger transcription factor **TRA-1**. The molecular workings of this regulatory module are still being elucidated. Similarity of **TRA-2** to patched receptors and of **TRA-1** to GLI proteins suggests that parts of the global pathway originally derived from a Hedgehog signaling pathway. **TRA-1** controls all aspects of somatic sexual differentiation, presumably by regulating a variety of tissue- and cell-specific downstream targets, including the cell death regulator **EGL-1** and the male sexual regulator **MAB-3**. Sex determination evolves rapidly, and conservation of sexual regulators between phyla has been elusive. An apparent exception involves DM domain proteins, including **MAB-3**, which control sexual differentiation in nematodes, arthropods, and vertebrates. Important issues needing more study include the detailed molecular mechanisms of the global pathway, the identities of additional sexual regulators acting in the global pathway and downstream of **TRA-1**, and the evolutionary history of the sex determination pathway. Recently developed genetic and genomic technologies and comparative studies in divergent species have begun to address these issues.

1. A global sex determination pathway controls sexual dimorphism

1.1. Two highly dimorphic sexes

The two natural sexes of *C. elegans* are the XX hermaphrodite and the XO male (Figure 1). Hermaphrodites are somatically female but can reproduce either by self-fertilization or by mating with males. Mating generates broods of half XX and half XO progeny. Self progeny mostly are XX, but about 1/500 are XO males, due to meiotic X chromosome nondisjunction, which ensures genetic exchange between individuals. The level of X chromosome nondisjunction, and hence the proportion of males in a population can be greatly increased by *him* (high incidence of males) mutations (Hodgkin et al., 1979). Because the two sexes differ in X chromosome dosage, a dosage compensation system has evolved to equalize expression of X-linked genes between XX and XO animals. This is accomplished by specifically reducing transcription of X-linked genes in XX animals (see X-chromosome dosage compensation). Sexual dimorphism is extensive in *C. elegans*, with most tissues and organs differing between the sexes in anatomy or physiology (Figure 1). Indeed, at least 40% of male and 30% of hermaphrodite cells are sexually specialized (Sulston and Horvitz, 1977). For further description of male- and hermaphrodite-specific development, see Male development and Hermaphrodite cell fate specification, respectively.

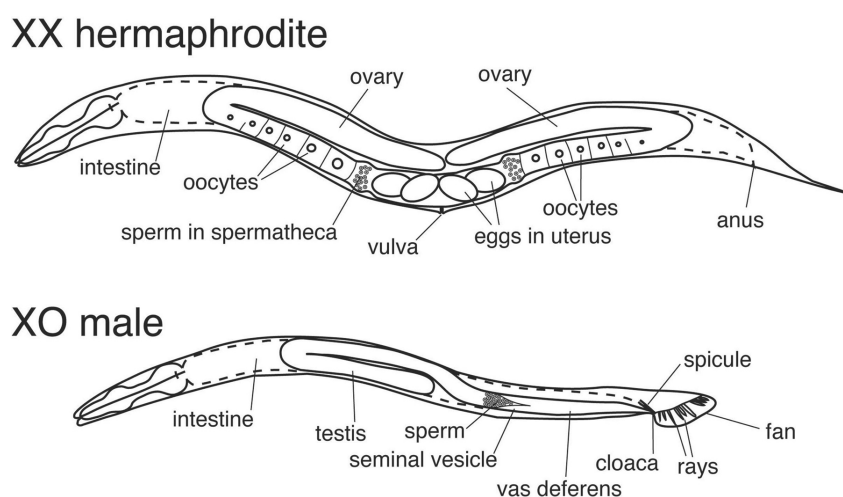


Figure 1. The two sexes of *C. elegans*. The hermaphrodites and males differ strikingly in overall body size and structures such as the somatic gonad and tail. Many other tissues and organs are sexually dimorphic, notably the nervous system and musculature. More detailed description of sex-specific cells and structures can be found elsewhere in the Sex determination section.

1.2. The global somatic sex determination pathway

The sexual fates of all somatic cells are controlled by a regulatory pathway whose activity differs between the sexes (Figure 2). This regulatory pathway is often called a “global” sex determination pathway to distinguish it from more specialized regulatory pathways that control sexual differentiation of particular tissues and lineages. The global pathway also controls dosage compensation, the process that equalizes X-linked gene expression in the two

sexes (see below). The global pathway was originally proposed on the basis of extensive genetic epistasis tests involving mutations affecting sex determination; it has been supported by subsequent molecular analysis (Hodgkin, 1987; Villeneuve and Meyer, 1990). In addition to the regulatory interactions shown in Figure 2, several minor interactions have been described, and these are thought to help reinforce the activity of the pathway in one sex or the other (Goodwin and Ellis, 2002). The same core regulatory pathway also controls sex determination in the germ line (see Vulval development). However, because hermaphrodites need to generate both oocytes and sperm, additional regulators are required in the germ line to temporarily reverse the sex determination decision, as described in Oocyte meiotic maturation and fertilization and Spermatogenesis .

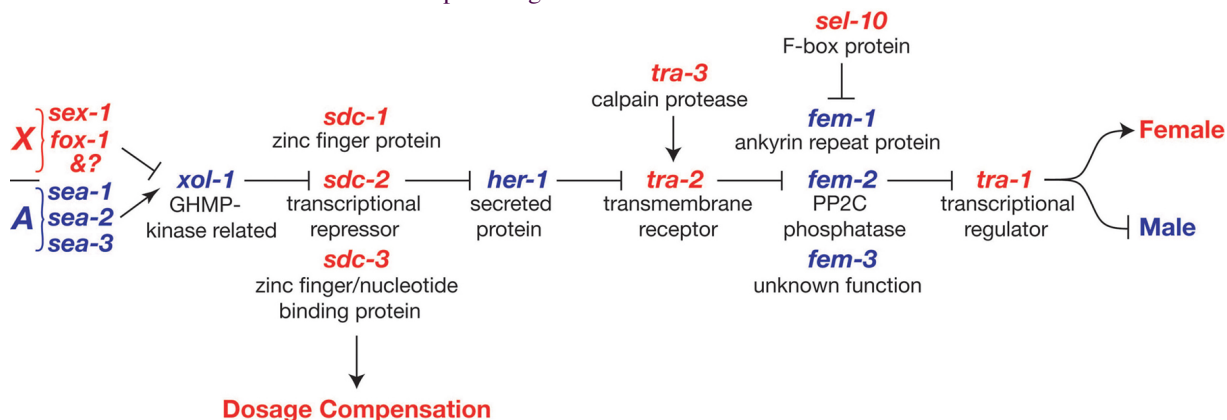


Figure 2. The global somatic sex determination pathway. Feminizing factors are shown in red and masculinizing factors in blue. Arrows represent positive regulatory interactions and bars represent negative interactions. As described in the text, additional minor regulatory interactions (not shown) may fine-tune the pathway. For more information on these genes see Wormbase.

The overall regulatory logic of the somatic sex determination scheme is seemingly straightforward. In essence, the number of X chromosomes controls a series of inhibitory interactions that ultimately sets the activity of the terminal regulator *tra-1* (*transformer 1*). *tra-1* then directs sexual differentiation throughout the soma (Hodgkin, 1987; Hodgkin and Brenner, 1977; Hunter and Wood, 1990).

If its purpose is to regulate *tra-1* activity, why does the pathway appear so needlessly baroque? Part of the explanation is that there are other functions and features, such as the need to coordinate dosage compensation with sex determination (see X-chromosome dosage compensation). Similarly, the *fem* genes promote spermatogenesis independently of their regulation of *tra-1* (Hodgkin, 1986). In addition, the pathway has regulatory interactions that enhance the plasticity and fidelity of the sex determination switch. One example is the ability of the pathway to reverse the sex determination decision in the XX germ line and allow both sperm and oocytes to form from a common stem cell pool. Another is the cell non-autonomous regulation of *tra-2* by *her-1*, described below.

2. The balance of X-linked and autosomal signal elements controls *xol-1*

The sex determination cascade is initiated in the early embryo by the ratio between the number of X chromosomes and sets of autosomes (X:A; Nigon, 1951). The X:A ratio determines the expression level of the upstream regulator *xol-1* (*XO* lethal 1), with a high ratio (XX) repressing *xol-1* expression and a low ratio (XO) permitting high *xol-1* expression. The activity of *xol-1* then controls the rest of the sex determination pathway (Miller et al., 1988; Rhind et al., 1995). XOL-1 is structurally related to GHMP small molecule kinases, but it does not bind ATP and its regulatory activity is not yet understood (Luz et al., 2003).

The X:A ratio regulates the activity of *xol-1* via the combined action of a set of “numerator” or X-signal elements on the X chromosome and “denominator,” or autosomal, signal elements elsewhere in the genome. Genetic dissection of the X chromosome using duplications and deletions revealed that the X-signal is polygenic, involving at least four components (Akerib and Meyer, 1994). Further dissection and mutational analysis have identified two X-signal components, *fox-1* and *sex-1*. FOX-1 is a putative RNA binding protein (Hodgkin et al., 1994; Nicoll et al., 1997; Skipper et al., 1999) that can post-transcriptionally repress a *xol-1* reporter when overexpressed (Nicoll et al., 1997). SEX-1 is related to nuclear hormone receptors and represses *xol-1* transcription, probably by direct binding to the *xol-1* promoter (Carmi et al., 1998).

The X-signal elements are opposed by autosomal signal elements. Three candidate genes were identified in a screen for suppressors of *fox-1 sex-1* lethality. Two of these genes have been cloned and found to encode likely transcriptional regulators: *sea-1* encodes a T-box protein, and *sea-2* a novel protein with zinc fingers and a metalloprotease domain (B. Meyer, pers. comm.; Powell et al., 2005).

The X:A signal system is exquisitely sensitive: although normally required to distinguish between ratios of 0.5 (XO) and 1.0 (XX), it can reliably discriminate a ratio of 0.67 from one of 0.75 (Madl and Herman, 1979).

3. Sex determination and dosage compensation are coordinately regulated

3.1. The SDC proteins connect sex determination and dosage compensation

The global sex determination pathway can be divided into two segments (Figure 2). The first segment (X:A signal to *sdc* genes) coordinately controls both sex determination and dosage compensation, ensuring that these processes always occur in the same mode. The segment from *her-1* onward is dedicated to sex determination.

The three SDC (sex determination and dosage compensation defective) proteins link the gene-specific regulation of *her-1* that controls sex determination with the chromosome-wide process of dosage compensation. The SDC proteins are components of a large protein complex that localizes to the X chromosomes of hermaphrodites early in embryogenesis and reduces their transcription by one-half (reviewed by Meyer, 2000; see X-chromosome dosage compensation). Components of the dosage compensation complex including the SDC proteins also control sex determination by binding to the promoter of *her-1* and repressing its transcription in XX animals to one-twentieth that of XO animals (*her* = hermaphroditization of XO; Chu et al., 2002; Dawes et al., 1999; Trent et al., 1991).

3.2. SDC proteins and repression of *her-1*

All three SDC proteins function both in repression of *her-1* and in dosage compensation. However, there are clear indications that these processes are mechanistically distinct. First, the two complexes have different compositions. In particular, *DPY-21* localizes to X but not to *her-1* (Yonker and Meyer, 2003). There also are differences in how the complexes recognize their targets. *SDC-2* plays a key role in X chromosome recognition, whereas *SDC-3* is more important for *her-1* recognition (Yonker and Meyer, 2003). The magnitude of transcriptional repression is much greater for *her-1* regulation than for dosage compensation of X-linked genes (twenty-fold versus two-fold, respectively; Dawes et al., 1999). Finally, the two activities of *SDC-3* are genetically separable, with the ATP binding motif of *SDC-3* required for regulation of *her-1* and the zinc fingers required for dosage compensation (Klein and Meyer, 1993).

SDC-2 is expressed only in hermaphrodites, as a result of male-specific repression by *XOL-1*, and this gives the SDC complex sex-specificity (Dawes et al., 1999). Several potential SDC binding elements have been identified in *her-1*. Two of these contain a 15 bp sequence not found on X; binding of SDC complexes to this element may mediate formation of a sex determination-specific complex (Chu et al., 2002). Two dominant gain-of-function alleles of *her-1* have been shown to alter promoter sequences and result in constitutive *her-1* expression (Perry et al., 1994). These mutations interfere with binding and regulation by the SDC complex (Chu et al., 2002).

4. HER-1 and TRA-2 provide cell non-autonomous control of sexual fate

4.1. HER-1 inactivates TRA-2

her-1 controls the segment of the pathway that is dedicated to sex determination. *HER-1* is a small secreted protein and promotes male development in a cell non-autonomous manner, by inhibiting the function of the transmembrane protein *TRA-2A* (Hunter and Wood, 1992; Perry et al., 1993). A model illustrating likely molecular interactions between the sex-determining proteins from *HER-1* to *TRA-1* is shown in Figure 3.

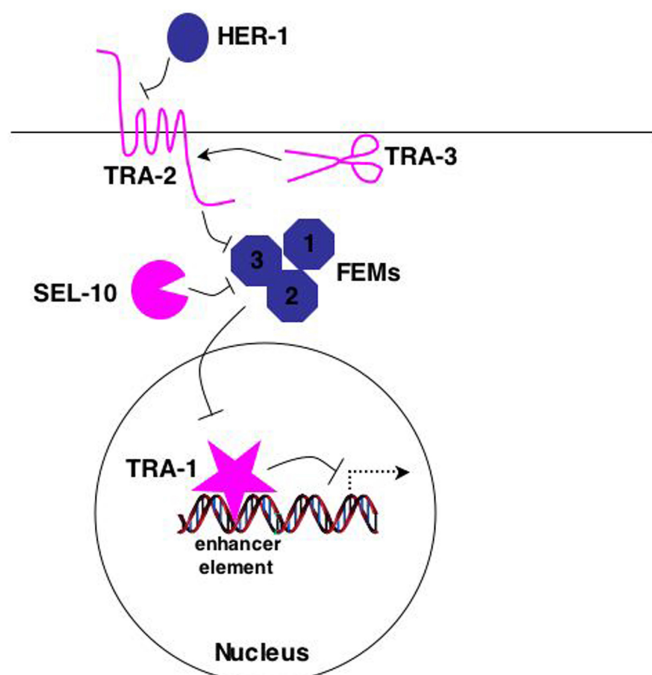


Figure 3. Molecular model of sex determination. Diagram illustrates likely relationships between sex determining proteins from HER-1 to the terminal global regulator TRA-1. Masculinizing proteins are blue, and feminizing proteins are pink. HER-1 is present only in XO males due to XX-specific transcriptional repression by the SDC proteins. In XO animals HER-1 binds the transmembrane receptor TRA-2A, blocking the ability of an intracellular domain of TRA-2A to inhibit FEM activity, and consequently the FEM proteins can inhibit TRA-1A. The TRA-3 protease may cleave TRA-2A to generate a fragment, TRA-2ic, with stronger feminizing activity. In XX animals active TRA-2A or TRA-2ic binds and inhibits FEM-3. As FEM-3 can bind FEM-2, this regulation may inhibit or sequester both proteins. The FEM proteins are targeted for proteasomal degradation by the F box protein SEL-10. When active (in XO animals) the FEM proteins somehow inhibit the activity of the transcriptional repressor TRA-1A. Active TRA-1A in XX animals binds DNA and blocks transcription driven by nearby enhancer elements in genes including *mab-3*, *egl-1* and probably *celh-30*, thereby establishing sexually dimorphic expression of specific genes in specific cell types. A number of molecular details remain poorly understood. In particular, it is not yet clear how TRA-2A inhibits the FEM proteins or how the FEM proteins inhibit TRA-1A.

There is compelling evidence for direct HER-1/TRA-2A binding: TRA-2A expression is necessary and sufficient to confer HER-1 binding to cultured cells, and the probable interaction interfaces of HER-1 and TRA-2A have been identified by a combination of genetic and structural studies (Hamaoka et al., 2004; Kuwabara, 1996). Having a cell non-autonomous step in the sex determination pathway has been suggested to help avoid errors by coordinating the sexual fates of cells. With a largely fixed cell lineage, *C. elegans* is limited in its ability to repair errors in cell fate specification. Secreted HER-1 also allows somatic control of germline sexual fate (Hunter and Wood, 1992).

tra-2 also is regulated by translational repression. This is of particular importance in the germ line (see Vulval development), but also is likely to affect TRA-2 expression in the soma. Disruption of elements in the 3' UTR of *tra-2* can cause weak dominant feminization of the male soma and increased translation of reporters (Doniach, 1986; Goodwin et al., 1993). A candidate to mediate repression via the *tra-2* 3' UTR is the essential gene *laf-1* (Goodwin et al., 1997).

4.2. TRA-2 inactivates the FEM proteins

The three FEM proteins (*fem* = feminization) are a diverse group whose biochemistry is still largely unclear. FEM-1 has ankyrin repeats, but otherwise no obvious conserved motifs, FEM-2 is a serine/threonine protein phosphatase, and FEM-3 is novel (Ahringer et al., 1992; Chin-Sang and Spence, 1996; Pilgrim et al., 1995; Spence et al., 1990). All three are required for male development and, based on genetic epistasis tests, they appear to act between *tra-2* and *tra-1* to control somatic sex (Hodgkin, 1986). TRA-2A inhibits the FEM proteins in hermaphrodites, regulation that is prevented in males by HER-1. Regulation of the *fem* genes is not transcriptional, as they show similar expression levels in both sexes (Ahringer et al., 1992; Gaudet et al., 1996; Pilgrim et al., 1995). Instead regulation is likely to involve protein/protein interactions: *in vitro* and yeast two-hybrid interactions have been demonstrated between TRA-2A and FEM-3 (Mehra et al., 1999) and between FEM-3 and FEM-2 (Chin-Sang and Spence, 1996). Consequently, a simple model is that association of HER-1 with an extracellular domain of

TRA-2A blocks the ability of an intracellular domain of TRA-2A to associate with and inhibit FEM-3 and FEM-2. How these interactions affect FEM protein function, whether the three FEM proteins form a complex *in vivo*, and what substrate FEM-2 dephosphorylates remain to be determined.

TRA-2A function may be enhanced by TRA-3, a calcium-dependent protease that can cleave TRA-2A *in vitro* to produce a peptide (TRA-2ic) that is predicted from transgenic experiments to have feminizing activity (Barnes and Hodgkin, 1996; Sokol and Kuwabara, 2000). Another negative regulator of the FEM proteins in hermaphrodites is SEL-10, one of a large family of F-box proteins. SEL-10 has weak feminizing activity *in vivo*, coimmunoprecipitates all three FEM proteins from transfected cells, and appears to target FEM-1 and FEM-3 for proteasomal degradation (Jager et al., 2004).

5. High versus low *tra-1* activity determines somatic cell sexual fates

5.1. TRA-1 is the nematode GLI protein

The global sex determination pathway appears to act entirely through the terminal regulator *tra-1*. Active *tra-1* promotes hermaphrodite development and prevents male development (Hodgkin, 1983; Hodgkin, 1987). *tra-1* expresses two mRNAs encoding proteins with zinc fingers (Zarkower and Hodgkin, 1992). The larger transcript encodes TRA-1A, with five zinc fingers, and the smaller RNA encodes TRA-1B, which contains only the amino terminus and first two zinc fingers. It is unknown whether both proteins contribute to *tra-1* activity, but TRA-1A binds DNA *in vitro*, while TRA-1B does not (Zarkower and Hodgkin, 1993).

The zinc fingers of TRA-1 are highly similar to those of the *Drosophila* Ci and vertebrate GLI proteins, transcription factors regulated by Hedgehog signaling via the Patched and Smoothed co-receptors (Zarkower and Hodgkin, 1992). Although *C. elegans* lacks obvious Hedgehog or Smoothed homologs, it does have a number of Patched-related proteins, including TRA-2A (Kuwabara et al., 2000). This suggests that part of the global sex determination pathway originally derived from a Hedgehog signaling pathway.

5.2. *tra-1* has distinct functions in sex determination and gonadogenesis

tra-1(null) XX animals are transformed into pseudomales that can be fertile, so robust male development is possible in the absence of *tra-1* function. However, these *tra-1* pseudomales frequently have small and disorganized somatic gonads, indicating that some *tra-1* activity is required for development of the male somatic gonad (Hodgkin, 1987; Schedl et al., 1989). The gonadal defects stem at least in part from roles in the organization and proliferation of the gonadal primordium that are distinct from the sex determination function of *tra-1*.

tra-1 controls gonadal development in concert with at least two other factors. During embryogenesis *tra-1* and *ehn-3* together regulate formation of the somatic gonadal primordium (Mathies et al., 2004), and after hatching, *tra-1* and *fkh-6* redundantly control gonadal cell proliferation (Chang et al., 2004).

5.3. Regulation of *tra-1* activity

How *tra-1* activity is controlled is still poorly understood. Many strong gain-of-function mutations have been isolated (Hodgkin, 1987). All affect a short protein sequence common to TRA-1A and TRA-1B (the “gain-of-function domain”), suggesting that regulation of TRA-1 is primarily post-translational (de Bono et al., 1995). As *tra-1* activity is controlled by the three FEM proteins, an attractive model is that a FEM protein inhibits the activity of TRA-1 via interaction with the gain-of-function domain. Regulation may involve the localization, modification, or degradation of TRA-1 (or a combination of these, as sex-specific differences in processing and nuclear accumulation of the TRA-1 protein(s) have been observed; Segal et al., 2001; A. Spence, pers. comm.).

6. TRA-1 links global and tissue-specific sexual regulation

TRA-1 directs sexually dimorphic development at the cellular level, with diverse consequences in different cell types. Direct transcriptional regulation by TRA-1 in somatic cells has thus far been shown for *mab-3* (Yi et al., 2000) and *egl-1* (Conradt and Horvitz, 1999). Both genes are transcriptionally repressed by TRA-1 in specific hermaphrodite cell types in order to prevent male differentiation. TRA-1 represses *mab-3* transcription in the XX intestine, thereby preventing MAB-3 from repressing yolk protein transcription in hermaphrodites (Yi and Zarkower, 1999; Yi et al., 2000). *egl-1* is a global regulator of programmed cell death (Conradt and Horvitz, 1998); however a TRA-1 binding site is found nearly 6 kb downstream of the *egl-1* coding region. Disruption of this site in

egl-1 gain-of-function mutants results in ectopic *egl-1* transcription in the HSN neurons causes the death of these cells (Conradt and Horvitz, 1999). TRA-1 likely represses the homeobox gene *ceh-30* to control sex-specific death of the four CEM neurons: *ceh-30* is required for CEM survival in males; and mutations in a conserved TRA-1 consensus binding site near *ceh-30* cause ectopic CEM survival in XX animals (H. Schwartz and H.R. Horvitz, pers. comm.).

From the limited data available, a picture emerges in which TRA-1 is expressed widely but exerts highly selective effects on target gene expression in particular cell types. This specificity may arise from short-range transcriptional repression, in which TRA-1 binding to DNA only blocks the activity of *cis* elements within a limited distance (Yi et al., 2000). By repressing some enhancer elements in a target gene without affecting others, a globally-expressed regulator like TRA-1 can control expression of each target gene selectively in particular cells or tissues. It has been suggested that short-range repression facilitates “sampling” of new target genes by the global regulator during evolution and thereby provides superior evolutionary flexibility (Cai et al., 1996; Gray et al., 1994; Zarkower, 2001).

7. Downstream sexual regulators: the interface between sex determination and sexual differentiation

The critical role of TRA-1 in establishing sexual dimorphism and its function as a likely transcriptional regulator have been known for some time. Despite this, surprisingly little is known of the genes that function directly downstream of TRA-1 in specific cell lineages, tissues, and organs to generate sexual dimorphism. For example, *tra-1* controls whether the posterior seam cells V5 and V6 generate male V rays or hermaphrodite lateral hypodermis (see [Male development](#)), but no TRA-1 target has been identified in this lineage. Regulation by TRA-1 of *mab-3* and *egl-1* accounts for only a small proportion of sexual dimorphism; many additional direct targets likely await discovery.

The establishment of sexual dimorphism can be roughly divided into two steps, first the sex determination decision, and second the execution of sexual differentiation. In general, mutations that abrogate sex determination cause sex reversal, and this is true of all the genes in the global pathway. By contrast, mutations in regulators of sexual differentiation cause defects in this process, but not sex reversal. TRA-1 and its targets are at the interface between these processes. *mab-3* illustrates that the distinction between regulators of sex determination and sexual differentiation can be hazy. *mab-3* mutant males have a feminized intestine that produces yolk, but the V ray lineages in the *mab-3* male tail are defective rather than feminized (Shen and Hodgkin, 1988). Another gene at this interface is the gonadal regulator *fkh-6*, which has a sex-determining function in the early male somatic gonad, and a differentiation role later in the hermaphrodite gonad (Chang et al., 2004).

8. Some aspects of sex determination may be evolutionarily conserved

Primary sex-determining mechanisms are extremely varied and evolve rapidly (see [The evolution of nematode sex determination](#)). The lability of sex determination is clearly demonstrated by the ease with which the process can be brought under the control of different genes in the pathway. Hodgkin has shown that any of seven autosomal sex determination genes can be genetically manipulated to become the primary determinant of sex, and that any of the five autosomes can thereby become a sex chromosome (Hodgkin, 2002). Given the facility with which new sex determination pathways can be created in the laboratory, they would be expected to be highly diverse in nature. This is the case: none of the sex-determining genes in [Figure 2](#) has been found to have a homolog playing a similar role outside the nematode phylum.

So far the best candidates for conserved sexual regulators are downstream genes related to *Drosophila doublesex*. These are termed DM domain (*doublesex* and *mab-3* related) genes, after the conserved DNA binding motif they share (Erdman and Burtis, 1993; Raymond et al., 1998; Zhu et al., 2000). In *C. elegans* these include *mab-3* and *mab-23*, both of which regulate multiple aspects of male differentiation and behavior (Lints and Emmons, 2002; Shen and Hodgkin, 1988; Yi et al., 2000). Genes encoding DM domain proteins have been shown to regulate sexual development in mammals (Raymond et al., 2000) and fish (Matsuda et al., 2002; Nanda et al., 2002), and are likely, based on sexually dimorphic expression and sex-linkage, to play similar roles in other vertebrates with different sex determination mechanisms (Zarkower, 2001). Thus it appears that at least some downstream sexual regulators have an ancient involvement in sexual dimorphism that is more stable than that of the upstream primary sex determination pathway. While *mab-3* and *mab-23* primarily function in sexual differentiation rather than sex determination, comparison with *Drosophila* and vertebrate homologs suggests that the ancestral role of

these genes may have been in sex determination. As more sexual regulators downstream of **TRA-1** are identified, other conserved factors may be found.

9. Open questions

Much progress has been made in understanding the regulation of somatic sex determination in *C. elegans*, but new questions have been raised and some old ones remain unanswered. Questions needing more experimental attention can be divided into four categories:

9.1. Molecular mechanisms of the global pathway

How the cascade of inhibitory interactions that converts X chromosome dosage into **TRA-1** activity functions is known in outline, but not in great detail. Do the X and autosomal signal elements antagonize one another directly or indirectly? How does the interaction of **TRA-2A** with **FEM-3** inactivate the FEM proteins, and how does **HER-1** binding to **TRA-2A** prevent this inactivation? How do the FEM proteins repress *tra-1* activity? **FEM-2** is a phosphatase, so does regulation of **TRA-1** involve sex-differential phosphorylation, and if so what is the kinase? How is **TRA-1** processed to generate sex-specific peptides, and what are their activities?

9.2. Other regulators

Most of known sex determining genes were found in genetic screens for defects in sexual differentiation. These screens have probably identified all of the genes whose loss of function phenotype is severe sex reversal. However, although many of these screens were conducted in sensitized genetic backgrounds, other regulators must have been missed, particularly essential genes and those that act redundantly. Two lines of evidence make it clear that such genes exist. First, there are biochemical activities unaccounted for, such as the putative kinase whose phosphorylation is reversed by **FEM-2**. Second, there is genetic evidence for additional regulators. As an example, mutating *xol-1* enhances the masculinization of XX *tra-(null)* mutants, suggesting that another regulator(s) acts downstream of *xol-1* and in parallel with *tra-2* to promote *tra-1* activity (Hodgkin and Brenner, 1977; Miller et al., 1988). Focused molecular screens should allow the identification of such regulatory molecules.

9.3. What genes act downstream of **TRA-1**?

TRA-1 activity directs alternative cell fates throughout the soma. This involves transcriptional repression of dedicated sexual regulators (*mab-3*) and of more general cellular factors (*egl-1*), and can control the physiology of entire organs like the intestine or the fates of individual cells. Is *mab-3* or *egl-1* the more typical **TRA-1** target? Does **TRA-1** act exclusively as a transcriptional repressor or might it also activate transcription? Much work remains to find genes that act downstream of **TRA-1** in other tissues. This will aid in understanding how sexual dimorphism is established in different cell types and how sexual regulation interacts with spatial and temporal regulation during development. The improved availability of reporter genes marking sexually dimorphic tissues and new techniques for global analysis of gene expression and function should accelerate progress.

9.4. How has the sex determination pathway evolved?

Evolution of sex determination pathways is covered in detail in [The evolution of nematode sex determination](#). The *C. elegans* sex determination pathway has been proposed to have arisen from a “bottom up” accretion of negative regulators during evolution (Wilkins, 1995). Genetic investigations of somatic sex determination in other nematodes have begun, and these, along with comparative molecular studies, are certain to be informative. As mentioned earlier, the pathway from *her-1* to *tra-1* appears to derive from a hedgehog signaling pathway which might have been co-opted in one step; comparative studies should clarify this question. As more divergent species are studied, it also will become clearer which parts of the *C. elegans* pathway are ancient and which are more recently recruited. In particular, it will be important to test suggestions that DM domain genes like *mab-3* and *mab-23* are descended from very ancient metazoan sex-determining genes.

10. References

Ahringer, J., Rosenquist, T.A., Lawson, D.N., and Kimble, J. (1992). The *Caenorhabditis elegans* sex determining gene *fem-3* is regulated post-transcriptionally. *EMBO J.* 11, 2303–2310. [Abstract](#)

- Akerib, C.C., and Meyer, B.J. (1994). Identification of X chromosome regions in *Caenorhabditis elegans* that contain sex-determination signal elements. *Genetics* *138*, 1105–1125. [Abstract](#)
- Barnes, T.M., and Hodgkin, J. (1996). The *tra-3* sex determination gene of *Caenorhabditis elegans* encodes a member of the calpain regulatory protease family. *EMBO J.* *15*, 4477–4484. [Abstract](#)
- Cai, H.N., Arnosti, D.N., and Levine, M. (1996). Long-range repression in the *Drosophila* embryo. *Proc. Natl. Acad. Sci. USA* *93*, 9309–9314. [Abstract Article](#)
- Carmi, I., Kopczynski, J.B., and Meyer, B.J. (1998). The nuclear hormone receptor *SEX-1* is an X-chromosome signal that determines nematode sex. *Nature* *396*, 168–173. [Abstract Article](#)
- Chang, W., Tilmann, C., Thoenke, K., Markussen, F.H., Mathies, L.D., Kimble, J., and Zarkower, D. (2004). A forkhead protein controls sexual identity of the *C. elegans* male somatic gonad. *Development* *131*, 1425–1436. [Abstract Article](#)
- Chin-Sang, I.D., and Spence, A.M. (1996). *Caenorhabditis elegans* sex-determining protein *FEM-2* is a protein phosphatase that promotes male development and interacts directly with *FEM-3*. *Genes Dev.* *10*, 2314–2325. [Abstract](#)
- Chu, D.S., Dawes, H.E., Lieb, J.D., Chan, R.C., Kuo, A.F., and Meyer, B.J. (2002). A molecular link between gene-specific and chromosome-wide transcriptional repression. *Genes Dev.* *16*, 796–805. [Abstract Article](#)
- Conradt, B., and Horvitz, H.R. (1998). The *C. elegans* protein *EGL-1* is required for programmed cell death and interacts with the Bcl-2-like protein *CED-9*. *Cell* *93*, 519–529. [Abstract Article](#)
- Conradt, B., and Horvitz, H.R. (1999). The *TRA-1A* sex determination protein of *C. elegans* regulates sexually dimorphic cell deaths by repressing the *egl-1* cell death activator gene. *Cell* *98*, 317–327. [Abstract Article](#)
- Dawes, H.E., Berlin, D.S., Lapidus, D.M., Nusbaum, C., Davis, T.L., and Meyer, B.J. (1999). Dosage compensation proteins targeted to X chromosomes by a determinant of hermaphrodite fate. *Science* *284*, 1800–1804. [Abstract Article](#)
- de Bono, M., Zarkower, D., and Hodgkin, J. (1995). Dominant feminizing mutations implicate protein-protein interactions as the main mode of regulation of the nematode sex-determining gene *tra-1*. *Genes Dev.* *9*, 155–167. [Abstract](#)
- Doniach, T. (1986). Activity of the sex-determining gene *tra-2* is modulated to allow spermatogenesis in the *C. elegans* hermaphrodite. *Genetics* *114*, 53–76. [Abstract](#)
- Erdman, S.E., and Burtis, K.C. (1993). The *Drosophila* doublesex proteins share a novel zinc finger related DNA binding domain. *EMBO J.* *12*, 527–535. [Abstract](#)
- Gaudet, J., VanderElst, I., and Spence, A.M. (1996). Post-transcriptional regulation of sex determination in *Caenorhabditis elegans*: widespread expression of the sex-determining gene *fem-1* in both sexes. *Mol. Biol. Cell* *7*, 1107–1121. [Abstract](#)
- Goodwin, E.B., and Ellis, R.E. (2002). Turning clustering loops: sex determination in *Caenorhabditis elegans*. *Curr. Biol.* *12*, R111–120. [Abstract Article](#)
- Goodwin, E.B., Hofstra, K., Hurney, C.A., Mango, S., and Kimble, J. (1997). A genetic pathway for regulation of *tra-2* translation. *Development* *124*, 749–758. [Abstract](#)
- Goodwin, E.B., Okkema, P.G., Evans, T.C., and Kimble, J. (1993). Translational regulation of *tra-2* by its 3' untranslated region controls sexual identity in *C. elegans*. *Cell* *75*, 329–339. [Abstract Article](#)
- Gray, S., Szymanski, P., and Levine, M. (1994). Short-range repression permits multiple enhancers to function autonomously within a complex promoter. *Genes Dev.* *8*, 1829–1838. [Abstract](#)

Hamaoka, B.Y., Dann, C.E., 3rd, Geisbrecht, B.V., and Leahy, D.J. (2004). Crystal structure of *Caenorhabditis elegans* HER-1 and characterization of the interaction between HER-1 and TRA-2A. *Proc. Natl. Acad. Sci. USA* *101*, 11673–11678. [Abstract Article](#)

Hodgkin, J. (1983). Two types of sex determination in a nematode. *Nature* *304*, 267–268. [Abstract Article](#)

Hodgkin, J. (1986). Sex determination in the nematode *C. elegans*: analysis of *tra-3* suppressors and characterization of *fem* genes. *Genetics* *114*, 15–52. [Abstract](#)

Hodgkin, J. (1987). A genetic analysis of the sex-determining gene, *tra-1*, in the nematode *Caenorhabditis elegans*. *Genes Dev.* *1*, 731–745. [Abstract Article](#)

Hodgkin, J. (1987). Sex determination and dosage compensation in *Caenorhabditis elegans*. *Annu. Rev. Genet.* *21*, 133–154. [Abstract Article](#)

Hodgkin, J. (2002). Exploring the envelope. Systematic alteration in the sex-determination system of the nematode *Caenorhabditis elegans*. *Genetics* *162*, 767–780. [Abstract](#)

Hodgkin, J., Horvitz, H.R., and Brenner, S. (1979). Nondisjunction mutants of the nematode *C. elegans*. *Genetics* *91*, 67–94.

Hodgkin, J., Zellan, J.D., and Albertson, D.G. (1994). Identification of a candidate primary sex determination locus, *fox-1*, on the X chromosome of *Caenorhabditis elegans*. *Development* *120*, 3681–3689. [Abstract](#)

Hodgkin, J.A., and Brenner, S. (1977). Mutations causing transformation of sexual phenotype in the nematode *Caenorhabditis elegans*. *Genetics* *86*, 275–287. [Abstract](#)

Hunter, C.P., and Wood, W.B. (1990). The *tra-1* gene determines sexual phenotype cell-autonomously in *C. elegans*. *Cell* *63*, 1193–1204. [Abstract Article](#)

Hunter, C.P., and Wood, W.B. (1992). Evidence from mosaic analysis of the masculinizing gene *her-1* for cell interactions in *C. elegans* sex determination. *Nature* *355*, 551–555. [Abstract Article](#)

Jager, S., Schwartz, H.T., Horvitz, H.R., and Conradt, B. (2004). The *Caenorhabditis elegans* F-box protein SEL-10 promotes female development and may target FEM-1 and FEM-3 for degradation by the proteasome. *Proc. Natl. Acad. Sci. USA* *101*, 12549–12554. [Abstract Article](#)

Klein, R.D., and Meyer, B.J. (1993). Independent domains of the Sdc-3 protein control sex determination and dosage compensation in *C. elegans*. *Cell* *72*, 349–364. [Abstract Article](#)

Kuwabara, P.E. (1996). A novel regulatory mutation in the *C. elegans* sex determination gene *tra-2* defines a candidate ligand/receptor interaction site. *Development* *122*, 2089–2098. [Abstract](#)

Kuwabara, P.E., Lee, M.H., Schedl, T., and Jefferis, G.S. (2000). A *C. elegans* patched gene, *ptc-1*, functions in germ-line cytokinesis. *Genes Dev.* *14*, 1933–1944. [Abstract](#)

Lints, R., and Emmons, S.W. (2002). Regulation of sex-specific differentiation and mating behavior in *C. elegans* by a new member of the DM domain transcription factor family. *Genes Dev.* *16*, 2390–2402. [Abstract Article](#)

Luz, J.G., Hassig, C.A., Pickle, C., Godzik, A., Meyer, B.J., and Wilson, I.A. (2003). XOL-1, primary determinant of sexual fate in *C. elegans*, is a GHMP kinase family member and a structural prototype for a class of developmental regulators. *Genes Dev.* *17*, 977–990. [Abstract Article](#)

Madl, J.E., and Herman, R.K. (1979). Polyploids and sex determination in *Caenorhabditis elegans*. *Genetics* *93*, 393–402. [Abstract](#)

Mathies, L.D., Schvarzstein, M., Morphy, K.M., Billeloch, R., Spence, A.M., and Kimble, J. (2004). TRA-1/GLI controls development of somatic gonadal precursors in *C. elegans*. *Development* *131*, 4333–4343. [Abstract Article](#)

Matsuda, M., Nagahama, Y., Shinomiya, A., Sato, T., Matsuda, C., Kobayashi, T., Morrey, C.E., Shibata, N., Asakawa, S., Shimizu, N., et al. (2002). DMY is a Y-specific DM-domain gene required for male development in the medaka fish. *Nature* *417*, 559–563. [Abstract Article](#)

Mehra, A., Gaudet, J., Heck, L., Kuwabara, P.E., and Spence, A.M. (1999). Negative regulation of male development in *Caenorhabditis elegans* by a protein-protein interaction between TRA-2A and FEM-3. *Genes Dev.* *13*, 1453–1463. [Abstract](#)

Meyer, B.J. (2000). Sex in the worm: counting and compensating X-chromosome dose. *Trends Genet* *16*, 247–253. [Abstract Article](#)

Miller, L.M., Plenefisch, J.D., Casson, L.P., and Meyer, B.J. (1988). *xol-1*: a gene that controls the male modes of both sex determination and X chromosome dosage compensation in *C. elegans*. *Cell* *55*, 167–183. [Abstract Article](#)

Nanda, I., Kondo, M., Hornung, U., Asakawa, S., Winkler, C., Shimizu, A., Shan, Z., Haaf, T., Shimizu, N., Shima, A., et al. (2002). A duplicated copy of DMRT1 in the sex-determining region of the Y chromosome of the medaka, *Oryzias latipes*. *Proc. Natl. Acad. Sci. USA* *99*, 11778–11783. [Abstract Article](#)

Nicoll, M., Akerib, C.C., and Meyer, B.J. (1997). X-chromosome-counting mechanisms that determine nematode sex. *Nature* *388*, 200–204. [Abstract Article](#)

Nigon, V. (1951). Polyploidie experimentale chez un Nematode libre, *Rhabditis elegans*. *Maupas. Bull. Biol. Fr. Belg.* *85*, 187–225.

Perry, M.D., Li, W., Trent, C., Robertson, B., Fire, A., Hageman, J.M., and Wood, W.B. (1993). Molecular characterization of the *her-1* gene suggests a direct role in cell signaling during *Caenorhabditis elegans* sex determination. *Genes Dev.* *7*, 216–228. [Abstract](#)

Perry, M.D., Trent, C., Robertson, B., Chamblin, C., and Wood, W.B. (1994). Sequenced alleles of the *Caenorhabditis elegans* sex-determining gene *her-1* include a novel class of conditional promoter mutations. *Genetics* *138*, 317–327. [Abstract](#)

Pilgrim, D., McGregor, A., Jackle, P., Johnson, T., and Hansen, D. (1995). The *C. elegans* sex-determining gene *fem-2* encodes a putative protein phosphatase. *Mol. Biol. Cell.* *6*, 1159–1171. [Abstract](#)

Powell, J.R., Jow, M.M., and Meyer, B.J. (2005). The T-box transcription factor SEA-1 is an autosomal element of the X:A signal that determines *C. elegans* sex. *Dev. Cell.* *9*, 339–349. [Abstract Article](#)

Raymond, C.S., Murphy, M.W., O'Sullivan, M.G., Bardwell, V.J., and Zarkower, D. (2000). Dmrt1, a gene related to worm and fly sexual regulators, is required for mammalian testis differentiation. *Genes Dev.* *14*, 2587–2595. [Abstract Article](#)

Raymond, C.S., Shamu, C.E., Shen, M.M., Seifert, K.J., Hirsch, B., Hodgkin, J., and Zarkower, D. (1998). Evidence for evolutionary conservation of sex-determining genes. *Nature* *391*, 691–695. [Abstract Article](#)

Rhind, N.R., Miller, L.M., Kopczynski, J.B., and Meyer, B.J. (1995). *xol-1* acts as an early switch in the *C. elegans* male/hermaphrodite decision. *Cell* *80*, 71–82. [Abstract Article](#)

Schedl, T., Graham, P.L., Barton, M.K., and Kimble, J. (1989). Analysis of the role of *tra-1* in germline sex determination in the nematode *Caenorhabditis elegans*. *Genetics* *123*, 755–769. [Abstract](#)

Segal, S.P., Graves, L.E., Verheyden, J., and Goodwin, E.B. (2001). RNA-Regulated TRA-1 nuclear export controls sexual fate. *Dev. Cell* *1*, 539–551. [Abstract Article](#)

Shen, M.M., and Hodgkin, J. (1988). *mab-3*, a gene required for sex-specific yolk protein expression and a male-specific lineage in *C. elegans*. *Cell* *54*, 1019–1031. [Abstract Article](#)

Skipper, M., Milne, C.A., and Hodgkin, J. (1999). Genetic and molecular analysis of *fox-1*, a numerator element involved in *Caenorhabditis elegans* primary sex determination. *Genetics* *151*, 617–631. [Abstract](#)

Sokol, S.B., and Kuwabara, P.E. (2000). Proteolysis in *Caenorhabditis elegans* sex determination: cleavage of TRA-2A by TRA-3. *Genes Dev.* *14*, 901–906. [Abstract](#)

Spence, A.M., Coulson, A., and Hodgkin, J. (1990). The product of *fem-1*, a nematode sex-determining gene, contains a motif found in cell cycle control proteins and receptors for cell-cell interactions. *Cell* *60*, 981–990. [Abstract Article](#)

Sulston, J.E., and Horvitz, H.R. (1977). Post-embryonic cell lineages of the nematode, *Caenorhabditis elegans*. *Dev. Biol.* *56*, 110–156. [Abstract Article](#)

Trent, C., Purnell, B., Gavinski, S., Hageman, J., Chamblin, C., and Wood, W.B. (1991). Sex-specific transcriptional regulation of the *C. elegans* sex-determining gene *her-1*. *Mech. Dev.* *34*, 43–55. [Abstract Article](#)

Villeneuve, A.M., and Meyer, B.J. (1990). The regulatory hierarchy controlling sex determination and dosage compensation in *Caenorhabditis elegans*. *Adv. Genet.* *27*, 117–188. [Abstract](#)

Wilkins, A.S. (1995). Moving up the hierarchy: a hypothesis on the evolution of a genetic sex determination pathway. *Bioessays* *17*, 71–77. [Abstract Article](#)

Yi, W., Ross, J.M., and Zarkower, D. (2000). Mab-3 is a direct *tra-1* target gene regulating diverse aspects of *C. elegans* male sexual development and behavior. *Development* *127*, 4469–4480. [Abstract](#)

Yi, W., and Zarkower, D. (1999). Similarity of DNA binding and transcriptional regulation by *Caenorhabditis elegans* MAB-3 and *Drosophila melanogaster* DSX suggests conservation of sex determining mechanisms. *Development* *126*, 873–881. [Abstract](#)

Yonker, S.A., and Meyer, B.J. (2003). Recruitment of *C. elegans* dosage compensation proteins for gene-specific versus chromosome-wide repression. *Development* *130*, 6519–6532. [Abstract Article](#)

Zarkower, D. (2001). Establishing sexual dimorphism: conservation amidst diversity? *Nat. Rev. Genet.* *2*, 175–185. [Abstract Article](#)

Zarkower, D., and Hodgkin, J. (1992). Molecular analysis of the *C. elegans* sex-determining gene *tra-1*: a gene encoding two zinc finger proteins. *Cell* *70*, 237–249. [Abstract Article](#)

Zarkower, D., and Hodgkin, J. (1993). Zinc fingers in sex determination: only one of the two *C. elegans* Tra-1 proteins binds DNA *in vitro*. *Nucleic Acids Res.* *21*, 3691–3698. [Abstract](#)

Zhu, L., Wilken, J., Phillips, N.B., Narendra, U., Chan, G., Stratton, S.M., Kent, S.B., and Weiss, M.A. (2000). Sexual dimorphism in diverse metazoans is regulated by a novel class of intertwined zinc fingers. *Genes Dev.* *14*, 1750–1764. [Abstract Article](#)

